Abstract:

A61K 39/395 (2006.01) C12P 21/08 (2006.01)
A61K 39/00 (2006.01) C07K 16/00 (2006.01)

The invention relates to the preparation and use of biologically active human soluble tumor necrosis factor (TNF) monomers or trimers, and of immunologically active antibodies or ligands for human TNF. The methods disclosed provide for the treatment of current and newly developing human diseases for which human TNF is known to be involved with (e.g., rheumatoid arthritis, ankylosing spondylitis, juvenile rheumatoid arthritis, psoriatic arthritis, atherosclerosis, metabolic syndrome, Alzheimer's Disease, HIV, Type II diabetes) mediated by human TNF are further disclosed.
COMPOSITIONS FOR SELECTIVE REDUCTION OF CIRCULATING BIOACTIVE SOLUBLE TNF AND METHODS FOR TREATING TNF-MEDIATED DISEASE

INCORPORATION-BY REFERENCE OF MATERIAL SUBMITTED IN ELECTRONIC FORM

Applicant hereby incorporates by reference the Sequence Listing material filed in electronic form herewith. This file is labeled GGP1 iPCT_ST25.txt", was created on January 23, 2014, and is 2 KB in size.

BACKGROUND OF THE INVENTION

Tumor necrosis factor (TNF; previously referred to as tumor necrosis factor-a) is a proinflammatory cytokine that plays a major role in the pathogenesis of rheumatoid arthritis and associated inflammatory diseases, such as ankylosing spondylitis, juvenile rheumatoid arthritis, and psoriatic arthritis. The human proform, transmembrane-bound TNF (tmTNF), is a 26-kDa homotrimer comprising three non-covalently associated monomers, each monomer having N-terminal sequence imbedded in the cell membrane. Each monomer of tmTNF has a 233 amino acid sequence (UniProtKB/Swiss-Prot entry Accession No. P01375). Soluble TNF (sTNF) is a homotrimer formed by enzymatic cleavage from its pro-form tmTNF. Each monomer of the sTNF trimer has a 157 amino acid sequence (SEQ ID NO: 1), which is the same sequence as aa77 to 233 of the published Acc No. P01374.

Both forms of active TNF (tmTNF and sTNF) exist as homotrimers and engage trimeric receptors that recognize receptor-binding sites in the grooves between the TNF monomers in assembled homotrimers. The grooves between the monomers comprise amino acid sequence from two contiguous monomers. The receptor binding regions of both forms of TNF are identical.

Trimer integrity is essential for biological function. For tmTNF, trimeric structure is established intracellularly before tmTNF insertion into the cell membrane and is maintained in tmTNF by the anchoring of the protein stems passing through the membrane plus further lipid anchoring by palmitoylated amino acid side chains at the membrane.
boundary. In contrast, sTNF active trimers dissociate freely into inactive monomers and dimers that reform as active sTNF homotrimers in a steady-state equilibrium between the three forms.

Anti-TNF biologies have provided a major advance in the management of the above-noted inflammatory diseases with anti-TNF monoclonal antibodies REMICADE (Infliximab; Janssen Biotech, Inc.) and HUMIRA (Adalimumab, Abbott Laboratories), and a chimeric solubilized TNF receptor fused to Fc, i.e., ENBREL (Entanercept, Biogen, Inc) being widely used. This therapeutic and marketing success is marred by the rare but statistically significant occurrence of serious infections and malignancies, likely related to concomitant blockade of tmTNF function impairing immune defenses. These adverse occurrences have included the development of tuberculosis, systemic fungal infection and other intracellular infections due principally to intracellular pathogens such as Mycobacterium tuberculosis, Listeria monocytogenes and Histoplasma capsuiatum, and certain forms of cancer. These results were unsurprising since these agents block pro-inflammatory sTNF but also block tmTNF, which is essential for juxtacrine cellular control of such intracellular infections and malignancies.

Because the receptor binding regions of both forms of TNF are identical, there has been little hope for the development of new monoclonal antibodies selectively blocking receptor engagements of one form versus the other. Antibodies to short sequences of TNF have not lead to useful therapeutics. For example, in 1987, Socher et al. in exploring antibodies to full or partial synthetic sequences of TNF, observed a high polyclonal antibody response to the TNF fragment 1-15 that appeared to block bioactivity and receptor binding of TNF. However, this 16-year old observation has not lead to the development of additional therapeutic reagents, likely because the TNF receptor is a discontinuous surface region not associated with TNF amino acids 1-15. Subsequent researchers in 2001 coupled TNF amino acids 4-23 conjugated to papillomavirus-like particles, and observed an induction of polyclonal antibodies, and an attenuation of experimental arthritis. Other researchers in 2007 used the same fragment TNF aa4-23 coupled to a virus-like particle-based composition and induced antibodies that attenuated experimental arthritis. No suppression of resistance to infection occurred, in contrast with full length TNF immunization. Because these TNF fragments were not directed to receptor binding regions of TNF, these publications displayed no further teachings or
suggestion of therapeutic use of the resulting polyclonal antibodies; and further research has not been published since that date.

One more recent attempt to selectively suppress the pro-inflammatory activity of sTNF while preserving tmTNF function required for innate immunity involved the design of synthetic dominant-negative TNF monomer variants that formed trimers that were inactive. These were shown to attenuate experimental arthritis without suppressing innate immunity to infection, emphasizing the major role of sTNF in pathogenesis of arthritis. Another approach has been the search for small-molecule drugs that interact with the inter-monomer contact regions. One molecule, SP304, bound such a contact region with μM affinity to effect trimer disruption in vitro.

Despite the plethora of literature in the field of anti-TNF treatment for a variety of inflammatory disorders, there remains a need in the art for new and useful compositions and methods for generating therapeutic or prophylactic immunogenic compositions for these diseases which do not result in adverse side effects due to suppression of cellular immunity.

SUMMARY OF THE INVENTION

As described herein the inventor has provided selective anti-TNF monomer-specific biologic compositions and various methods of use thereof which do not affect the structure or bioactivity of tmTNF or increase the treated subject's susceptibility to infection by an intracellular pathogen.

In one aspect, an isolated or synthetic antibody or ligand is provided that specifically binds to an epitope of a dissociated monomer of human TNF. The binding of the antibody or ligand to the monomer disrupts or prevents assembly of the monomer into bioactive trimeric human sTNF. In one embodiment, the antibody or ligand binds specifically to monomer-specific epitope A2 of sequence PSDKPVAH, amino acids 8-15 of SEQ ID NO: 1 or PSDKPVAHV, amino acids 8-16 of SEQ ID NO: 1. In still another embodiment, the antibody or ligand binds specifically to monomer-specific epitope F of sequence EPIYLGGVF, amino acids 116 to 124 of SEQ ID NO: 1. In one embodiment, the antibody is a bi-specific antibody directed to epitopes A2 and F.

In another aspect, a pharmaceutical composition comprises one or more isolated or synthetic antibody or ligand that specifically binds to an epitope of a dissociated monomer
of human TNF, the binding disrupting or preventing assembly of the monomer into bioactive trimeric human sTNF, and a pharmaceutically acceptable carrier or diluent. In certain embodiments, the composition contains one or two of the above-described antibodies.

In yet a further aspect, methods for preparing or generating isolated or synthetic antibodies or ligands that specifically bind to an epitope of a dissociated monomer of human TNF are provided.

In still another aspect, a method for treating a subject having a disease mediated by soluble human TNF (sTNF) comprises reducing the amount, concentration or bioactivity of sTNF in the blood of a subject having the disease without affecting the amount, concentration or bioactivity of tmTNF. This is accomplished by disrupting, preventing or reducing the in vivo assembly or reassembly of dissociated monomers of TNF into bioactive trimeric human sTNF without affecting the amount, concentration or bioactivity of tmTNF. In certain embodiments, this method employs the monomer-specific antibodies, bi-specific antibodies, ligands and compositions described above and herein. In one embodiment, the disease is rheumatoid arthritis (RA), juvenile rheumatoid arthritis, ankylosing spondylitis (AS), psoriatic arthritis or psoriasis.

In still another aspect, sTNF elevations are also implicated in initial HIV infection, and the reoccurrence of latent HIV infection and type II diabetes. Therefore, in still another aspect, a method for preventing a subject infected with HIV-1 and treated with anti-retroviral drugs from developing a new infection (or rebound infection due to latent HIV) comprises administering to the subject treated with anti-retroviral therapy (ART) with an isolated or synthetic selective anti-TNF monomer-specific antibody or ligand, or pharmaceutical composition, as described herein, after the ART is discontinued.

In yet another aspect, a method for treating a subject with type II diabetes comprises administering periodically to a subject in need thereof an isolated or synthetic selective anti-TNF monomer-specific antibody or ligand, or pharmaceutical composition, as described above, optionally in combination with known anti-diabetic therapies.

In other aspects, the TNF monomer-specific antibodies, ligands, bi-specific antibodies, or compositions described herein are provided for use in the treatment of a disease or disorder mediated by soluble human TNF, including any disease identified herein. In other aspects, use of the antibodies, ligands, or compositions described herein
in preparation of a medicament for treatment of a disease or disorder mediated by soluble human TNF, including any disease identified herein, is provided.

Other aspects and advantages of these methods and compositions are described further in the following detailed description.

5

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 illustrates the immunogen and epitope detector sequences for TNF interface A and TNF Interface F, which were used to identify the sequences and margins of the three monomer-specific TNF epitopes A1, A2 and F described herein as well as the binding activities of the anti-TNF monomer-specific antibodies generated thereto. These sequences are discussed in Example 1 below.

Fig. 2A is a bar graph illustrating the TNF monomer-specific epitopes by immunizing a rat with sTNF amino acids 1-23. Antibody binding responses were measured on recombinant TNF (rTNF), a mixture of monomers, dimers and trimers, and on synthetic peptides employing the indicated terminally truncated TNF peptides. The synthetic detector peptides included: TNF amino acids 1-10 of SEQ ID NO: 1, which demonstrated no binding at all; TNF amino acids 5-12 of SEQ ID NO: 1, minimal to no binding; the A1 epitope SSRTPSDKP, TNF amino acids 4-12 of SEQ ID NO: 1; TNF amino acids 4-11 of SEQ ID NO: 1 SSRTPSDK; TNF amino acids 9-15 of SEQ ID NO: 1 SDKPVAH; the A2 epitope sequence of amino acids 8-15 of SEQ ID NO: 1; TNF amino acids 8-14 of SEQ ID NO: 1 PSDKPVA; and TNF amino acids 16-23 of SEQ ID NO: 1 VVANPQAE, which exhibited no binding. Clearly only two overlapping epitopes were detected by rat antiserum to TNF amino acids 1-23, which were epitope A1, TNF amino acids 4-12 and A2, TNF amino acids 8-15.

Fig. 2B is a bar graph illustrating the margins of the A2 epitope PSDKPVAHV detected in mouse serum using the same procedure described in Fig. 2A, with synthetic peptides. The synthetic detector peptides included: TNF amino acids 10-17 of SEQ ID NO: 1 DKPVAHVV, which demonstrated minimal binding; TNF amino acids 9-16, minimal binding; the A2 epitope PSDKPVAHV, TNF amino acids 8-16 of SEQ ID NO: 1; TNF amino acids 8-15 of SEQ ID NO: 1 PSDKPVAH; and TNF amino acids 8-14 of SEQ ID NO: 1 PSDKPVA, no binding. When rat and rabbit sera were used, the boundaries of epitope A2 are amino acids 8-15 of SEQ ID NO: 1. The immune system of the mouse

5
sees only TNF amino acids 8-16 and does not bind to TNF amino acids 8-15, as shown in the graph.

Fig. 2C is a bar graph illustrating the results of determining the margins of the monomer-specific TNF epitopes by immunizing a rat or mouse with sTNF amino acids 112-128 of SEQ ID NO: 1. KPWYEPIYLGGVFQLEK (interface region underlined; F epitope in bold). Antibody binding responses were measured on recombinant TNF (rTNF), a mixture of monomers, dimers and trimers, and on synthetic peptides employing the indicated four terminally truncated TNF peptides. The synthetic peptides were IYLGGVF, amino acids 118-124 of SEQ ID NO: 1; PIYLGGVF, amino acids 117-124 of SEQ ID NO: 1, EPIYLGGVF, amino acids 116-124 of SEQ ID NO: 1 (epitope F) and EPIYLGGV, amino acids 116-123 of SEQ ID NO: 1. The greatest binding was to the aal 16-124 peptide, thereby indicating the margins of the epitope (referred to as epitope F).

Fig. 3A illustrates data from the sandwich assay using 200 ng/mL sTNF, biotin-labeled antibody and non-biotin labeled antibody, as described in Example 2 below. The binding curves show that commercial REMICADE anti-TNF antibody, when labeled with biotin and mixed with sTNF (trimers, dimers and monomers), binds multimeric forms of TNF. The biotinylated antibody-TNF in the mixture still has available TNF trimer epitopes that can bind and form a sandwich with the unlabeled plated REMICADE antibody( ). In contrast, Protein A/Protein G purified IgG (o) was obtained from rats immunized with TNF amino acids 1-23 of SEQ ID NO: 1. This purified IgG contains a mixture of monomer-specific anti-TNF that selectively bind epitope A1 and monomer-specific anti-TNF that selectively bind epitope A2. The purified IgG (o) does not sandwich in the assay, because once these antibodies bind the TNF monomers in the TNF mixture, the labeled monomer-specific antibody-TNF complexes have no available monomer-specific epitopes to bind to the plated unlabeled monomer-specific antibody on the plate. Labeled TNF monomer-specific antibody- monomers complexes are simply washed from the plate without binding. REMICADE antibody that binds trimeric TNF was used as a positive control in this assay. Thus neither anti-A1 nor anti-A2 antibodies bind the trimeric form of sTNF.

Fig. 3B illustrates data from a similar sandwich assay to that of Fig. 3A, using as reagents: commercial REMICADE anti-TNF antibody ( ); affinity purified IgG from monomer-specific antisera to the TNF epitope F that selectively binds only epitope F:
EPIYLGGVF (X) and Protein A/G purified IgG from monomer-specific antisera to the TNF epitope F that selectively binds only epitope F: EPIYLGGVF (□). Thus, in contrast to the commercial REMICADE anti-TNF antibodies, the anti-F antibodies do not bind the trimeric form of sTNF.

Fig. 4A illustrates the results of an assay of antibody inhibition of sTNF-induced cytotoxicity in target cells, using antiserum generated to TNF amino acids 1-23 that contains antibodies that selectively bind the monomer specific epitopes PSDKPV AH and SSRTPSDKP (epitopes A2 and A1, respectively). The titers of antisera with 200 pg/mL TNF in all wells are displayed under the bars, from 1x10^(-6), 2.5x10^(-5), 1x10^(-5), 2.5x10^(-4), 1x10^(-4), and 2.5x10^(-3). The last bar is sTNF in 50% NRS.

Fig. 4B illustrates functional blocking of sTNF cytotoxicity in actinomycin-treated WEHI cells in the assay described in Example 2 by antiserum to TNF epitope F. The indicated dilutions (50%, 16.7%, 5.6%, 1.85%, 0.48%) of monomer-specific antiserum generated to the immunogen KPWYEPIYLGGVFQLEK, amino acids 112-128 of SEQ ID NO: 1 (the F beta sheet interface sequence of TNF), in rats were compared for their ability to inhibit sTNF bioactivity with 200 pg/mL TNF and 50% NRS (normal rat serum). As is shown in this figure, inhibition of TNF cytotoxicity was shown using antiserum diluted from 0.48% to 50%. Thus, the monomer-specific antiserum to epitope F showed the ability to inhibit the cytotoxic effect of sTNF on the cells as evidenced by increasing replication of cells in the presence of the antiserum. Statistical significance was determined by one-way ANOVA and post testing with Dunnett's test. REMICADE antibody, which binds trimeric TNF, was used as a positive control in this assay.

Fig. 5 is a bar graph showing inhibition of sTNF cytotoxicity in WEHI cells by monoclonal antibody generated to TNF epitopes A1 or A2. Cell replication (OD) was measured in WEHI cells grown in the presence of no TNF and the cells showed good replication. WEHI cells grown in the presence of 0.2 ng/ml full length TNF 1-157 of SEQ ID NO: 1 demonstrated that TNF inhibited replication. Cells grown in TNF plus the commercial REMICADE anti-TNF antibody at 1 μg/mL, showed that the antibody returned replication to the same levels demonstrated in the absence of TNF. TNF plus the inventor's monoclonal monomer-specific antibody A1-4H6 to TNF epitope A1 (amino acids 4-12 of SEQ ID NO: 1) at 10 μg/mL, failed to inhibit WEHI cell replication, which remained at the levels of cells exposed to TNF alone. WEHI cells were cultured in the
presence of TNF plus the inventor's monomer-specific monoclonal antibody A2-8D12 to TNF epitope A2 (amino acids 8-16 of SEQ ID NO: 1) at 10 μg/mL; and TNF plus the inventor's monomer-specific monoclonal antibody A2-10H10 to TNF epitope A2 (amino acids 8-16 of SEQ ID NO: 1) at 0.25 μg/mL. Data from these two latter monomer-specific monoclonal antibodies showed highly statistically significant inhibition of sTNF cytotoxicity (i.e., reduction of cell killing). The second monomer-specific monoclonal antibody A2-10H10 showed high sensitivity, a 40X increase in potency in this assay over the other monomer-specific anti-A2 antibody A2-8D12. These results demonstrate that monomer-specific anti-A2 monoclonal antibodies inhibit sTNF and that one such antibody A2-10H10 exhibits a higher affinity than the other, as demonstrated by the 40 fold lower dose. No inhibition even at a high dose was demonstrated by the monomer-specific anti-A1 monoclonal antibody. These data demonstrate that the effects of the TNF amino acid 1-23 polyclonal antisera were due to the monomer-specific anti-A2 antibodies only.

**DETAILED DESCRIPTION OF THE INVENTION**

The inventor has provided selective anti-TNF monomer-specific biologic compositions and various methods of use based on the determination that antibodies and/or other ligands directed to selected epitopes partially or fully within the internal interface contact region of TNF free monomers block their association with other monomers and cause progressive disruption of bioactive sTNF trimer formation. It is advantageous to have an antibody or ligand, e.g., a monoclonal antibody or bi-specific antibody, that selectively blocks the activity of sTNF but not tmTNF for the treatment of rheumatoid arthritis (RA), juvenile rheumatoid arthritis, ankylosing spondylitis (AS) and psoriatic arthritis (PA), psoriasis, and other inflammatory diseases.

1. **ANTIBODIES/LIGANDS**

Thus, this invention provides an isolated or synthetic antibody or ligand that specifically binds to an epitope of a dissociated monomer of human TNF, the binding disrupting or preventing assembly of monomers into bioactive trimeric human sTNF.

The inventor determined that antibodies directed to certain epitopes partially or fully within the internal interface contact region of free TNF monomers would block their association with other monomers and cause progressive disruption of trimer formation. In
contrast to the known publications on TNF, the inventor determined that there were two overlapping epitopes in the TNF sequence of amino acids 1-15 of SEQ ID NO: 1. One epitope A1 which spanned amino acids 4-12 of SEQ ID NO: 1 was monomer-specific, but did not disrupt trimer formation. The other epitope A2, which spanned amino acids 8-15 or 8-16 of SEQ ID NO: 1, was monomer-specific and did disrupt trimer formation.

Further the inventor identified a new unrecognized epitope F, amino acids 116-124 of SEQ ID NO: 1 in the F β sheet of TNF, which was monomer-specific and did disrupt trimer formation. The discovery and new uses of antibodies or ligands that specifically bind these epitopes is discussed in detail below and in the examples.

Early x-ray crystallography studies established that the 157 amino acid (SEQ ID NO: 1) sTNF monomers formed an "elongated, anti-parallel β pleated sheet sandwich with "jelly-roll" topology". Three monomers in intimate but non-covalent association constituted the active trimer\(^1\). Five stretches of amino acid sequences formed the interface β sheet contact surfaces: A, aal-18; A', aa35-39; C, aa54-67; F, aal 14-126 and H, aal49-157, all of SEQ ID NO: 1, where A, A', C, F and H refer to a β sheet naming convention\(^11\). The inventor explored all five regions for potential B cell epitopes and detected and mapped antibodies to two epitopes partially (the A β sheet) and one epitope fully (the F β sheet) within an interface region, all being outside the known regions of the receptor binding sites of TNF\(^2\)\(^3\)\(^4\)\(^5\). See, Example 1 below.

In one embodiment, the selective anti-TNF monomer-specific antibody or ligand binds the epitope A2 of sequence PSDKPVAH, amino acids 8-15 of SEQ ID NO: 1 or sequence PSDKPVAHV, amino acids 8-16 of SEQ ID NO: 1. In another embodiment a selective anti-TNF monomer-specific antibody or ligand binds the A1 epitope of sequence SSRTPSDKP, amino acids 4-12 of SEQ ID NO: 1. In another embodiment, the TNF monomer-specific antibody or ligand binds the F epitope of sequence EPIYLGGVF, amino acids 116 to 124 of SEQ ID NO: 1.

As described below, antibodies to A1 epitope, while TNF monomer-specific, have been found to be inactive in blocking TNF function when used alone. Antibodies to these A2 and F epitopes specifically bind TNF monomers but not trimers, disrupt assembly of sTNF trimers and inhibit sTNF function \textit{in vitro}. These monomer-specific anti-A2 and anti- F epitope antibodies or ligands do not bind transmembrane TNF (tmTNF) and do not affect the structure or bioactivity of tmTNF. Additionally, these antibodies or ligands do
not bind intact bioactive trimeric human sTNF. Monoclonal antibodies to the A2 and F epitopes are useful for therapeutic use as safer sTNF-selective anti-TNF biologies for rheumatoid arthritis and related inflammatory diseases with sTNF-related pathologies.

As used herein, the term "antibody" refers to an intact immunoglobulin having two light and two heavy chains. The term "antibody fragment" refers to less than an intact antibody structure, including, without limitation, an isolated single antibody chain, an sc-Fv construct, a Fab construct, a Fab_2 construct, or a light chain variable or complementarity determining region (CDR) sequence, etc. The term "bi-specific" antibody refers to a synthetically or recombinantly produced antibody that contains one heavy and/or one light chain that binds to one epitope, e.g., the A2 epitope, and one heavy and/or one light chain that binds to a second epitope, e.g., the F epitope. The term "ligand" is used to refer to other synthetic molecules or sequences that can be designed to bind to the indicated epitopes.

"High affinity" is the strength of binding of the antibody or ligand in question to the TNF monomer-specific epitope A2 or F. In one embodiment, the antibodies/ligands to A2 or F bind at an affinity of less than 10 nanomolar (nM). In another embodiment, the antibodies/ligands to A2 or F bind at an affinity of less than 1 nanomolar (nM). In another embodiment, the antibodies/ligands to A2 or F bind at an affinity of less than 100 picomolar (pM). In another embodiment, the antibodies/ligands to A2 or F bind at an affinity of less than 10 nM.

Thus, in one embodiment, the antibody or ligand as described herein may be a polyclonal, affinity-purified or high affinity antibody or a fragment thereof. In one embodiment, the antibody or ligand is a monoclonal antibody or a fragment thereof. In another embodiment, the antibody or ligand is an isolated single chain of an antibody.

Still other forms of antibodies, such as a synthetic antibody, a recombinant antibody, a chimeric antibody, a humanized antibody, a human antibody or a fragment thereof can be employed as the ligand or antibody directed to one of the above described epitopes. Suitable fragments of such antibodies may also be employed. In yet another embodiment, the antibody or ligand or fragment thereof further comprises a polyethylene glycol (PEG) molecule. The antibody or ligand or fragment can be associated or fused with PEG by known conventional methodologies.
The production of antibodies or ligands that specifically bind to one of the selected epitopes, can employ conventional techniques. For example, polyclonal antibody compositions are typically produced by immunizing a selected mammal, e.g., a primate, rodent, or human, with a peptide/ polypeptide composition containing a specific epitope. See, e.g., the description of the antisera described in Figs. 2A-2C. The selection of high titer, high affinity polyclonal antibodies can be monitored by standard techniques, such as with an enzyme-linked immunosorbent assay and surface plasma resonance. If desired, the polyclonal antibody molecules can be isolated from the mammal, e.g., from the whole blood, plasma or serum, and further purified from the plasma or serum of the immunized mammal by conventional techniques. Conventional harvesting techniques can include plasmapheresis, protein A/G chromatography, among others. Such polyclonal antibody compositions may themselves be employed as pharmaceutical compositions of this invention.

Alternatively, monoclonal antibodies can be generated to any one of the epitopes by now conventional techniques, using antibody producing cells obtained from the immunized mammals and fused to non-IgG-producing myeloma cells to form hybridomas or from selection from activated immune B cells with extraction by known molecular biological techniques. These monoclonal antibodies can be further used to prepare other forms of antibodies and ligands, e.g., chimeric antibodies, humanized antibodies, human antibodies. Other antibody fragments or ligands can be produced by screening phage display libraries, antibody fragments and mixtures thereof. Techniques for generating these types of antibodies and ligands are well-known in the art and the ligands themselves may be generated using the disclosed amino acid sequences of the above-identified epitopes.

Chimeric antibodies may similarly be developed using known techniques. Chimeric antibodies are molecules in which different portions are derived from different animal species. Single chain antibodies may also be prepared by conventional methods, such as described in US Patent Nos. 4,946,778 and 4,704,692 using the variable portions of the polyclonal or monoclonal antibodies produced according to this invention. Antibody fragments, such as the Fab, F(ab)2 and scFv fragments and libraries thereof may also be employed in generation of the selective anti-TNF monomer-specific antibodies or ligands as described herein.
The production of bi-specific antibodies or ligands that specifically bind to two or more of the selected epitopes, can employ conventional techniques. It is within the skill of the art to develop bi-specific antibodies that bind multiple epitopes. See, e.g., Hornig N, Farber-Schwarz A., "Production of bispecific antibodies: diabodies and tandem scFv." 2012, Methods Mol Biol, 907:713-27; Speiss, C. et al, " Bispecific antibodies with natural architecture produced by co-culture of bacteria expressing two distinct half-antibodies, Jul 7, 2013, Nature Biotechnology, 31:753-758; and Jonathan S Martin and Zhenping Zhu, "Recombinant approaches to IgG-like bispecific antibodies", 2005 Acta Pharmacologica Sinica, 26: 649-658. In one embodiment, a bispecific antibody is developed which is capable of binding to or reacting with epitope A2 and epitope F. It is anticipated that such bispecific antibodies, e.g., the antibody reactive with A2 and F, will enhance avidity and create greater potency than the single anti-A2 or anti-F antibodies or ligands alone.

Other selective anti-TNF monomer-specific antibodies or ligands may be developed by screening recombinant combinatorial immunoglobulin scFv libraries (e.g., phage displays) with one of the above-identified TNF monomer-specific epitopes to isolate immunoglobulin library members that bind to the TNF monomer. See, e.g., Phage Display of Peptides and Proteins, A Laboratory Manual, eds. Kay, BK et al, Elsevier Inc. (1996), among other texts well known in the art. Kits for generating and screening phage display libraries are commercially available, e.g., Pharmacia Recombinant Phage Antibody System, Catalog No. 27-9400-01; Stratagene Phage Display kits, etc. See, e.g., US Patent No., 5,223,409, International Publication No. WO92/09690, WO90/02809, etc.

II. PHARMACEUTICAL COMPOSITIONS

In another aspect, a pharmaceutical composition comprises an isolated or synthetic antibody or ligand that specifically binds to an epitope of a dissociated monomer of human TNF, the binding disrupting or preventing assembly of the monomer into bioactive trimeric human sTNF.

The pharmaceutical composition contains one or more of the selective anti-TNF monomer-specific antibodies or ligands described above with a suitable carrier or diluent. Thus, in one embodiment, the pharmaceutical composition contains an anti-TNF monomer-specific antibody or ligand that specifically binds the A2 epitope having the
sequence PSDKPVAH, amino acids 8-15 of SEQ ID NO: 1 or PSDKPVAHV, amino acids 8-16 of SEQ ID NO: 1. In still another embodiment, the pharmaceutical composition contains a selective anti-TNF monomer-specific antibody or ligand that specifically binds the F epitope having the sequence EPIYLGGVF, amino acids 116 to 124 of SEQ ID NO: 1.

In another embodiment, a pharmaceutical composition comprises two anti-TNF monomer-specific antibodies or ligands. In one embodiment, the composition comprises an antibody or ligand that specifically binds the A2 epitope PSDKPVAH, amino acids 8-15 of SEQ ID NO: 1 or PSDKVPAHV, amino acids 8-16 of SEQ ID NO: 1 and an antibody or ligand that specifically binds the F epitope having the sequence EPIYLGGVF.

In yet a further embodiment, a pharmaceutical composition containing a bispecific antibody that specifically binds with the A2 epitope and the F epitope is also useful in interfering with soluble TNF trimer formation. Other forms of multi-ligand constructs known to the art may also take advantage of binding to A2 and/or F epitopes to provide trimer disruption. Alternatively, the anti-TNF monomer-specific compositions of this invention may be used in conjunction with, or sequentially with, other therapies or pharmaceutical regimens which are used conventionally to treat the various diseases mediated by sTNF.

These pharmaceutical compositions described herein also contain one or more pharmaceutically acceptable carriers or diluents. As defined herein, the pharmaceutically acceptable carrier suitable for use in an immunogenic proteinaceous composition of the invention are well known to those of skill in the art. Such carriers include, without limitation, water, saline, buffered saline, phosphate buffer, alcoholic/aqueous solutions, emulsions or suspensions. Other conventionally employed diluents, adjuvants and excipients, may be added in accordance with conventional techniques. Such carriers can include ethanol, polyols, and suitable mixtures thereof, vegetable oils, and injectable organic esters. Buffers and pH adjusting agents may also be employed. Buffers include, without limitation, salts prepared from an organic acid or base. Representative buffers include, without limitation, organic acid salts, such as salts of citric acid, e.g., citrates, ascorbic acid, gluconic acid, carbonic acid, tartaric acid, succinic acid, acetic acid, or phthalic acid, Tris, trimethanmine hydrochloride, or phosphate buffers. Parenteral carriers can include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride,
lactated Ringer's or fixed oils. Intravenous carriers can include fluid and nutrient
replenishes, electrolyte replenishers, such as those based on Ringer's dextrose and the
like. Preservatives and other additives such as, for example, antimicrobials, antioxidants,
chelating agents, inert gases and the like may also be provided in the pharmaceutical
carriers. The present invention is not limited by the selection of the carrier. The
preparation of these pharmaceutically acceptable compositions, from the above-described
components, having appropriate pH isotonicity, stability and other conventional
characteristics is within the skill of the art. See, e.g., texts such as Remington: The
Science and Practice of Pharmacy, 22nd ed, Lippincott Williams & Wilkins, publ, 2012;
and The Handbook of Pharmaceutical Excipients, 7th edit., eds. R. C. Rowe et al,

III. METHODS OF USE

A. Treatment of Certain Inflammatory Conditions

A method for treating a mammalian, preferably human, subject having a disease
mediated by soluble human TNF (sTNF) involves reducing the amount, concentration or
bioactivity of sTNF in the blood of a subject having the disease without affecting the
amount, concentration or bioactivity of tmTNF. This reduction occurs by disrupting,
preventing or reducing the in vivo assembly or reassembly of dissociated monomers of
TNF into bioactive trimeric human sTNF. Thus, in one embodiment, this method
comprises administering to a subject in need thereof an isolated or synthetic anti-TNF
monomer-specific antibody or ligand that specifically binds to an epitope of a dissociated
monomer of TNF. The selected antibodies or ligands do not bind intact bioactive sTNF
trimer. The selected antibody or ligand does not bind transmembrane TNF (tmTNF) and
does not affect the structure or bioactivity of tmTNF.

In one embodiment, the method is useful for the treatment of rheumatoid arthritis.
In another embodiment, the method is useful for the treatment of ankylosing spondylitis.
In another embodiment, the method is useful for the treatment of juvenile rheumatoid
arthritis. In still another embodiment, the method is useful for the treatment of psoriatic
arthritis. In still another embodiment, the method is useful for the treatment of psoriasis.
In another embodiment, the method is useful for the treatment of a pathogenic effect of
bioactive, trimeric sTNF produced during inflammation or during the course of an
inflammatory disorder. Still additional embodiments of the methods of the invention involve treatment of other diseases in which sTNF and/or inflammation at low or chronic levels plays a role. In one embodiment, such a disease is HIV-1. In another embodiment, the methods are useful for treating type 2 diabetes. In still other embodiment, the therapeutic selective anti-TNF monomer-specific antibodies or ligands are useful in methods for treating inflammation in the pathology of obesity. In another embodiment, the method is useful for the treatment of metabolic syndrome. In another embodiment, the method is useful for the treatment of atherosclerosis and associated cardiovascular disease. In another embodiment, the method is useful for the treatment of inflammation involved in the pathology of Alzheimer's disease. In another embodiment, the method is useful for the treatment of inflammation involved in the pathology of neurodegenerative diseases. Still other inflammatory diseases\(^{40}\) may be treated with the compositions and methods described herein. Such treatment is not burdened by the immune suppression and morbidity and mortality associated with non-selective agents.

Therefore, in one embodiment the antibody/ligand useful in the method binds the A2 epitope sequence PSDKPVAH or PSDKPVAHV. In another embodiment of the method, the antibody/ligand useful in the method binds the F epitope EPIYLGGVF. The binding of the antibody/ligand to these selected epitopes disrupts or prevents assembly of the monomer into bioactive trimeric human sTNF. In still further embodiments of this method, the subject is administered two of these selective anti-TNF monomer-specific antibodies/ligands. In still other embodiments, the pharmaceutical compositions may include the bispecific antibodies discussed above.

Another aspect of this method involves maintaining a reduced amount or concentration of bioactive trimeric sTNF in the subject's bloodstream over time. Such maintenance can involve repeated administration of one of more of the above-noted selective anti-TNF monomer-specific antibodies, ligands, monoclonal antibodies, bispecific antibodies or pharmaceutical compositions containing same. By use of these methods, the subject's susceptibility to infection by an intracellular pathogen, \textit{e.g.}, tuberculosis, bacterial sepsis, invasive fungal infection, or histoplasmosis, or to a malignancy, \textit{e.g.}, lymphoma or hepatosplenic T-cell lymphoma, is not increased by treatment.
According to these therapeutic methods, the selective anti-TNF monomer-specific antibody or ligand is present in a pharmaceutical composition in a pharmaceutically acceptable carrier or diluent. Any of the pharmaceutical compositions described above, e.g., containing one or two of the antibodies/ligands, and possibly antibodies directed to non-sTNF immunogens, can be employed.

In each of the above-described methods, these compositions of the present invention are administered by an appropriate route, e.g., by the subcutaneous, mucosal, intravenous, intraperitoneal, intramuscular, nasal, or inhalation routes. The presently preferred route of administration is subcutaneous, intravenous or intramuscular.

The amount of the selective anti-TNF monomer-specific antibody, ligand, monoclonal or bispecific antibodies, or constructs described above, with or without other antibodies or ligands to other immunogens, present in each dose, is selected with regard to consideration of the patient's age, weight, sex, general physical condition and the specific disease being treated. The amount of antibody required to produce an exogenous effect in the patient without significant adverse side effects varies depending upon the pharmaceutical composition employed. In patients with a disease medicated by sTNF, generally, each dose will comprise between about 5 to 400 mg/mL injection of the selective anti-TNF monomer-specific antibody in a sterile solution. Another dosage is about 200 mg/mL of the antibody. Still another dosage is about 100 mg/mL of the antibody. Still another embodiment is a dosage of about 50 mg/mL of the antibody. A further embodiment is a dosage of about 10 mg/mL of the antibody. When used together, dosages of each anti-TNF monomer-specific antibody to a different one of the two monomer-specific TNF epitopes may be the same. In another embodiment, due to the synergy between the two combined selected anti-TNF monomer-specific antibodies, a combination dosage is lower than additive single dosages of each antibody alone. For example, the dosage of a bi-specific antibody directed to A2 and F may be less than the dosage of an antibody to one of these epitopes alone. Additional combination with antibodies directed to other than sTNF epitopes may alter the dosage of the anti-TNF monomer-specific antibodies.

In one embodiment, the administration of the selective anti-TNF monomer-specific antibody/ligand is repeated periodically during the course of the disease. In various embodiments in which two of the selective anti-TNF monomer-specific antibodies are
administered in the course of treatment, each antibody/ligand in a pharmaceutically acceptable carrier is administered, either separately, in combination, or sequentially in any order.

The frequency of administration may range from weekly to monthly or bimonthly, and less frequently, and may depend upon the half-life of the antibody and the course of the particular disease. In one embodiment, the dosage is administered once a week. In another embodiment, the dosage is administered once every two weeks. In another embodiment the dosage is administered once a month. In another embodiment, the frequency of dosage administration is once every two or three months. Other dosage ranges may also be contemplated by one of skill in the art, particularly where administration of the antibody composition is in conjunction or sequential with other treatments for the disease.

B. *Treatment of HIV-1 Infection*

In still another aspect, a method for treating a subject to reduce or prevent re-infection or rebound infection with latent HIV-1 in a subject treated with anti-retroviral drugs is provided. According to this method, a subject receiving ART is administered the isolated or synthetic selective anti-TNF monomer-specific antibody or ligand, monoclonal antibody or bispecific antibody to epitopes A2 or F, or pharmaceutical composition containing same. In one embodiment, an anti-TNF monomer-specific antibody that binds A2 or F epitope is administered to a subject receiving anti-retroviral therapy. In another embodiment, an anti-TNF monomer-specific antibody or ligand or composition that binds A2 or F epitope is administered to a subject starting immediately after ART is discontinued. In still another embodiment, the anti-TNF monomer-specific antibodies/ligands/compositions are administered chronically to a subject both before ART treatment is discontinued and chronically after ART treatment is discontinued. The purpose of this method of treatment is to prevent re-infection or rebound infection of the subject with a different latent strain or variant of HIV after ART has successfully controlled the initial HIV infection.

In still other embodiments, the selective anti-TNF monomer-specific antibodies/ligands may be used in concert with other anti-HIV compositions (see e.g., US Patent No. 7,943,140).
According to any of the above methods, the selective anti-TNF monomer-specific antibody/ligands are administered in a pharmaceutically acceptable carrier, either separately, in combination, or sequentially in any order. The compositions, dosages, routes of administration and frequency of administration are anticipated to be as described. However, one of skill in the art, given the teachings of this application may employ other suitable dosages and routes of administration. Particularly for HIV, the administration of the selective anti-TNF monomer-specific antibody/ligand/compositions is expected to be repeated periodically for an indefinite period.

C. Treatment of Diabetes

In still another aspect, sTNF elevations are also implicated in type II diabetes. Yet a further embodiment of this invention involves a method for treating diabetes which may also be practiced utilizing the selective anti-TNF monomer-specific antibodies and pharmaceutical compositions described herein. In one embodiment, this method for treating a human subject with type II diabetes comprises administering periodically to a subject in need thereof an isolated or synthetic selective anti-TNF monomer-specific antibody or ligand, or pharmaceutical composition. The antibody, ligand or pharmaceutical composition may be any of those described specifically above. In one embodiment, the subject is concurrently treated with other diabetes medication. In still another embodiment, the administration of the selective anti-TNF monomer-specific antibody/ligand or composition is repeated periodically after the subject ceases treatment with other diabetes medications, such as insulin or oral drugs such as metformin.

IV. EXAMPLES

The following examples illustrate certain embodiments of the above-discussed compositions and methods. These examples do not limit the disclosure of the claims and specification.
EXAMPLE 1 - TNF EPITOPE MAPPING

The 157 amino acid TNF monomers SEQ ID NO: 1 have an elongated, anti-parallel β pleated sheet structure. When three monomers are associated in a non-covalent trimer, bioactive sTNF is formed. Five stretches of amino acid sequences form the interface β sheet contact surfaces:

A - KPVAVV, aal-18 of SEQ ID NO: 1;
A’ - ALLAN, aa35-39 of SEQ ID NO: 1;
C - GLYLIYSQVLFGKQ, aa54-67 of SEQ ID NO: 1;
F - WYEPIYLGVFQI, aal 14-126 of SEQ ID NO: 1; and
H - QVYFGIAP, aal 49-157 of SEQ ID NO: 1,

where A, A', C, F and H refer to a β sheet naming convention

To attain trimer disruption immunologically, the inventor theorized that antibody binding to epitope sequences that are wholly or partially within the contact area between adjacent monomers (the so-called internal or interface regions) would not bind to intact trimers of sTNF or tmTNF but would only bind to free monomers of TNF. In binding only to the free monomers, these antibodies would disrupt or prevent the ability of the monomers to re-associate and form active trimers.

Therefore, the inventor explored all five regions for potential B cell epitopes and detected and mapped antibodies to two epitopes partially (the A interface β sheet contact surface) and one epitope fully (the F interface β sheet contact surface) within an interface region. Rats or mice were immunized with synthetic peptides derived from the linear sequence from the five known interface regions of TNF identified above. These synthetic peptides sequences were conjugated with KLH, and adjuvants such as Freund's Complete or Incomplete Adjuvant were used. Alternatively the synthetic peptide sequences are incorporated in self adjuvating constructs, such as those described for HIV Tat constructs. Polyclonal antibodies isolated from rats immunized with each of the synthetic peptide sequences were evaluated on rTNF and also on the synthetic TNF peptides using conventional ELISAs. See, e.g., the protocols described for anti-human TNF/TNFSF1A antibody by R&D Systems, catalog number MAB610, clones 28401, pages 1 and 2 (June 17, 2005).

Binding to truncated sequences from larger peptide immunogens was used to delineate epitope margins. When antibodies were detected to a region of TNF, truncated
peptide sequences were used to determine the margins of the epitopes defined above. An antibody that bound exclusively to one of each the specific epitope sequences was referred to as a selective anti-TNF monomer-specific antibody of this invention. See, e.g., Fig. 1.

Table 1 sets out the interface regions, immunogens tested and epitopes detected from the epitope searches:

<table>
<thead>
<tr>
<th>Interface Region</th>
<th>Immunogens of SEQ ID NO: 1 Tested</th>
<th>Epitopes Detected*</th>
<th>Titers</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>aa1-15 and aa1-24</td>
<td>aa4-12: SSRTPSDKP (epitope A1) aa8-15: PSDKPVAH (epitope A2)</td>
<td>400,000 to 1 million</td>
</tr>
<tr>
<td>A'</td>
<td>aa35-39</td>
<td>No significant antibody response</td>
<td>--</td>
</tr>
<tr>
<td>C</td>
<td>aa52-67</td>
<td>No significant antibody response</td>
<td>--</td>
</tr>
<tr>
<td>F</td>
<td>aa112-128</td>
<td>aa116-124:EPIYLLGGVF (epitope F)</td>
<td>50,000 to 100,000</td>
</tr>
<tr>
<td>H</td>
<td>aa145-157</td>
<td>No significant antibody response</td>
<td>--</td>
</tr>
</tbody>
</table>

* Bold amino acids within interface sequences

Figs. 2A, 2B and 2C illustrate the margin identification of the epitopes.

All three epitope sequences are outside of the known regions of the receptor binding sites of TNF$^{1-15}$. The two overlapping B cell epitopes detected in synthetic peptide sequences overlapping interface contact region A are:

PSDKPVAH, amino acids 8-15 of SEQ ID NO: 1 and PSDKPVAHV, amino acids 8-16 of SEQ ID NO: 1; and

SSRTPSDKP, amino acids 4-12 of SEQ ID NO: 1.

Thus, the sTNF aal-23 sequence of SEQ ID NO: 1 contains only two B cell epitopes, each epitope containing interface amino acid(s) essential for antibody binding, but only epitope A2 contains amino acids necessary for trimer formation and biological function.

The entire sequence of the third epitope EPIrLGGVF (F sheet), amino acids 116-124 of SEQ ID NO: 1 was within interface contact region F. Two of the epitope amino acids, Ty$r_{19}$ and Glym, are critical for trimer formation and biological activity$^{13,24}$. 
In the three epitopes shown above, the bolded amino acids are within the internal interface regions. The italicized amino acids are those at which point mutations induce failure to form trimers and loss of biological activity.

Antibodies to each of three monomer-specific epitopes Al: SSRTPSDKP, A2: PSDKPVAH/PSDKVPAHV and F: EPIYLGGVF bind to synthetic rTNF (which comprises trimer, dimer and monomer forms) in the conventional ELISA assay. Antibodies to A1 SSRTPSDKP (both polyclonal antibodies and monoclonal antibodies) bound only two interface amino acids (Lysn and Pro^6), essential for antibody binding to the epitope. Neither of these amino acids have been shown to be critical for trimer formation and bioactivity of sTNF. Antibodies to A2 PSDKPVAH or PSDKVPAHV (both polyclonal antibodies and monoclonal antibodies) also masked Hisis that is critical to trimer formation and bioactivity of sTNF molecules. Only one other monomer specific antibody was detected, within the F monomer interface region. The F epitope spanned TNF amino acids 116-124 and induced polyclonal antibodies that were monomer-specific. These antibodies masked Tyr119 and Gly122, both critical to trimer formation and bioactivity of sTNF molecules.

EXAMPLE 2 - SELECTIVE BINDING ACTIVITY

Natural or synthetic sTNF consists of a mixture of inactive TNF monomers, inactive TNF dimers and bioactive TNF trimers. Each antibody used in the assays and commercially available anti-TNF antibodies (e.g., REMICADE) or commercially available TNF receptor chimera (e.g., ENBREL) binds to synthetic TNF coated on a plate, as in the conventional ELISAs performed in Example 1. However, detection of specific binding of an antibody/ligand to the monomeric, dimeric or trimeric form of sTNF requires appropriate selective assays. To demonstrate the selective binding activity of antibodies or ligands that bind only the three epitopes on a monomer as identified in Example 1, the following assays are performed:

A. Sandwich Assay

In one embodiment, a sandwich assay employs a biotinylated anti-TNF antibody which binds the sTNF in a sample, followed by detection with the same antibody, non-biotinylated, coated on a plate. The unlabeled antibody is plated (e.g., one of the inventor's epitope-binding antibodies or a commercial antibody, e.g., REMICADE
antibody). A sample containing synthetic sTNF mixed with the same anti-TNF antibody, which has been biotinylated, is prepared and then introduced to the plate. The plate is then washed and any bound sTNF sandwiched between the unlabeled bound antibody and the labeled detector antibody is measured using a suitable detector system, e.g., streptavidin/horseradish peroxidase, to generate a detectable signal.

If the antibody used as the capture/detector antibody is an antibody that binds sTNF trimeric form, a sandwich effect will be detected. Only sTNF trimers with the ability to simultaneously bind both the plate-bound antibody and the biotinylated antibody will demonstrate binding in this assay. This is because in the mixture not all binding sites on the trimers will be bound, thereby leaving extra binding sites to be captured on the plate. For example, the commercial REMICADE antibody demonstrates trimer binding in this assay.

In contrast, the inventor's antibodies that specifically bind a single epitope on a sTNF monomer do not demonstrate binding in this assay. Once each monomer is bound by the labeled antibody in the mixture, that epitope is no longer available for binding to the same unlabeled antibody on the plate when the mixture is added to the plate. A sandwich cannot form.

This assay can therefore be used to distinguish between the anti-TNF monomer-specific antibodies/ligands that bind to A2 or F selectively and that bind only sTNF monomers and those non-selective commercial and known anti-TNF antibodies that bind trimers, both tm-TNF and trimeric sTNF.

When employed in these assays, the inventor determined that antibodies (both polyclonal or monoclonal) to epitopes A1, A2 or F all were able to bind only sTNF monomers. Results from performance of this assay are shown in Figs. 3A and 3B.

B. Functional Assay

A functional assay is one that demonstrates the effects of the antibodies on TNF binding cells, such as actinomycin treated WEHI cells. See, e.g., the protocols for "Neutralization of Human TNF Bioactivity" described for anti-human TNF/TNFSF1A antibody by R&D Systems, catalog number MAB610, clones 28401, pages 1 and 2 (June 17, 2005), incorporated by reference herein. For the generation of the data in Figs. 4B and 5, in place of the cells L929, WEHI cells are used. This assay showed inhibition of TNF activity by the selected tested antibodies. In this assay the antibodies that bind one of the
three epitopes on the monomers as described herein are evaluated to determine if they have the ability to disrupt the formation of bioactive trimers of sTNF and thus inhibit sTNF activity in a dose response curve. Performance of this functional assay and its results are demonstrated in Figs. 4B and 5.

C. Results

In the sandwich assay described above, the selective antibodies that bound one of the three epitopes on the sTNF monomer: A1: SSRTPSDKP, A2: PSDKPVAH or PSDKPVAHV and F: EPIYLGGVF, bound only the monomeric form of sTNF. As described above, the biotinylated selective anti-TNF antibodies that bind one of the epitopes A1, A2 or F would not sandwich with sTNF to cause binding by the same selective non-biotinylated anti-TNF monomer-specific antibody coated plate. See, e.g., Figs. 3A and 3B. In contrast to the results for the anti-A2 and anti-F antibodies/ligands, the commercial anti-TNF REMICADE antibody demonstrated binding in the sandwich assay (see e.g., Fig. 3), showing binding to trimeric TNF. Additionally all polyclonal antibodies to TNF peptides encroaching an interface region were negative in the sandwich assay, showing lack of binding to trimers, demonstrating that binding was restricted to monomers.

However, only two of the antibodies that exhibited selective binding to one of the three epitopes, A1: SSRTPSDKP, A2: PSDKPVAH and F: EPIYLGGVF, produced dose-responsive inhibition of TNF, suppression of TNF inhibition of cell replication in actinomycin treated WEHI cells in the functional assay. See, e.g., Figs. 4A, 4B and 5. Antibodies to A2 and F were capable of disrupting trimer formation and were associated with inhibition of sTNF binding to TNF receptors and inhibition of cytotoxicity of sTNF on actinomycin treated WEHI cells. These assay findings provided evidence that the monomer specific antibodies to epitopes A2 and F disrupt trimer assembly by blocking specific amino acid side chains essential for inter-monomer binding.

Surprisingly, antibodies to the A1 epitope, while monomer specific, were inactive in blocking TNF function.

Throughout this specification, the words "comprise", "comprises", and "comprising" are to be interpreted inclusively rather than exclusively. The words "consist", "consisting", and its variants, are to be interpreted exclusively, rather than inclusively. It should be understood that while various embodiments in the specification
are presented using "comprising" language, under various circumstances, a related embodiment is also be described using "consisting of" or "consisting essentially of language. It is to be noted that the term "a" or "an", refers to one or more, for example, "an antibody" is understood to represent one or more antibodies. As such, the terms "a" (or "an"), "one or more," and "at least one" is used interchangeably herein.

Unless defined otherwise in this specification, technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs and by reference to published texts, which provide one skilled in the art with a general guide to many of the terms used in the present application. All documents listed or referred to herein, including US provisional applications No. 61/768044 and 61/756571, as well as the attached or electronic Sequence Listing, are incorporated herein by reference.

Given the teachings provided in this specification, one of skill in the art can generate antibodies and other antibody fragments, including high affinity polyclonal antibodies, affinity purified and humanized antibodies, monoclonal antibodies and bispecific antibodies that bind specifically to one or more of the epitopes A2 or F by conventional methodologies. Such antibodies and ligands are readily obtained and useful in the methods disclosed herein. Without further description, it is believed that one of ordinary skill in the art can, using the preceding description and the following illustrative examples, make and utilize the compositions of the present invention and practice the claimed methods. While the invention has been described and illustrated herein by references to various specific materials, procedures and examples, it is understood that the invention is not restricted to the particular combinations of material and procedures selected for that purpose. Numerous modifications and variations of the embodiments illustrated above are included in this specification and are expected to be obvious to one of skill in the art. Such modifications and alterations to the compositions and processes described herein are believed to be encompassed in the scope of the claims appended hereto.
References

11. Eck, M.J. & Sprang, S.R. The structure of tumor necrosis factor-alpha at 2.6 A
12. Van Ostade, X., et ah. Localization of the active site of human tumour necrosis
factor-alpha receptor binding site and structure-functional relationship. The Journal of
14. Yang, Z., et ah, Crystal structure of TNFalpha complexed with a poxvirus MHC-
related TNF binding protein. Nature Structural & Molecular Biology 16, 1189-1191
(2009).
Signaling 3, ra83 (2010).
16. Tang, P., et ah, Human pro-tumor necrosis factor is a homotrimer. Biochemistry
17. Utsumi, T., et ah Transmembrane TNF (pro-TNF) is palmitoylated. FEBS letters
500, 1-6 (2001).
18. Corti, A., et ah Oligomeric tumour necrosis factor alpha slowly converts into
19. Steed, P.M., et ah Inactivation of TNF signaling by rationally designed dominant-
20. Zalevsky, J., et ah Dominant-negative inhibitors of soluble TNF attenuate
experimental arthritis without suppressing innate immunity to infection. J Immunol 179,
(2005).
22. Wells, J.A. & McClendon, C.L. Reaching for high-hanging fruit in drug discovery
23. Yamamoto, R., et ah, Histidine-15: an important role in the cytotoxic activity of
35. US Patent No. 7,227,003
37. US Patent No. 7,070,775
38. US Patent No. 5,919,452
39. US Patent No. 5,698,419
WHAT IS CLAIMED IS:

1. An isolated or synthetic antibody or ligand that specifically binds to an epitope of a dissociated monomer of human TNF, said binding disrupting assembly of the monomer into bioactive trimeric human sTNF.

2. The antibody or ligand according to claim 1, wherein the TNF monomer-specific epitope has the sequence PSDKPVAH, amino acids 8-15 of SEQ ID NO: 1.

3. The antibody or ligand according to claim 1, wherein the TNF monomer-specific epitope has the sequence PSDKPVAHV, amino acids 8-16 of SEQ ID NO: 1.

4. The antibody or ligand according to claim 1, wherein the TNF monomer-specific epitope has the sequence EPIYLGGVF, amino acids 116 to 124 of SEQ ID NO: 1.

5. The antibody or ligand according to any of claims 1-4, which does not bind transmembrane TNF (tmTNF) and does not affect the structure or bioactivity of tmTNF.

6. The antibody or ligand according to any of claims 1-4, which does not bind intact bioactive trimeric human sTNF.

7. The antibody or ligand according to any of claims 1-6, which is a polyclonal, affinity-purified antibody or a fragment thereof, a monoclonal antibody or a fragment thereof, an isolated single chain of an antibody, a synthetic antibody, a recombinant antibody, a chimeric antibody, a humanized antibody, a human antibody, a bi-specific antibody, a high affinity antibody or ligand or a fragment thereof.

8. The antibody or ligand according to claim 7, which is a bi-specific antibody reactive with the TNF monomer-specific epitope having the sequence PSDKPVAH or PSDKPVAHV, amino acids 8-15 or 8-16 of SEQ ID NO: 1 and a TNF monomer-specific epitope having the sequence EPIYLGGVF, amino acids 116 to 124 of SEQ ID NO: 1.
9. The antibody or ligand according to claim 7, wherein said antibody fragment is an sc-Fv construct, a Fab construct, a Fab₂ construct, or a light chain variable or complementarity determining region (CDR) sequence.

10. The antibody or ligand according to claim 7, wherein the antibody or fragment further comprises a polyethylene glycol molecule.

11. A pharmaceutical composition comprising an isolated or synthetic antibody or ligand that specifically binds to an epitope of a dissociated monomer of human TNF, said binding disrupting assembly of the monomer into bioactive trimeric human sTNF, and a pharmaceutically acceptable carrier or diluent.

12. The pharmaceutical composition according to claim 11, wherein the antibody or ligand is an antibody or ligand of any of claims 1 to 10.

13. The pharmaceutical composition according to claim 11, comprising one or more antibodies or ligands selected from:
   (a) an antibody or ligand that specifically binds the TNF monomer-specific epitope having the sequence PSDKPVAH, amino acids 8-15 of SEQ ID NO: 1 or PSDKPVAHV, amino acids 8-16 of SEQ ID NO: 1; and
   (b) an antibody or ligand that specifically binds the TNF monomer-specific epitope having the sequence EPIYLGGVF, amino acids 116 to 124 of SEQ ID NO: 1.

14. The pharmaceutical composition according to claim 11, which comprises a bi-specific antibody reactive with the TNF monomer-specific epitope having the sequence PSDKPVAH or PSDKPVAHV, amino acids 8-15 or 8-16 of SEQ ID NO: 1 and a TNF monomer-specific epitope having the sequence EPIYLGGVF, amino acids 116 to 124 of SEQ ID NO: 1.

15. A method for treating a subject having a disease mediated by human TNF comprising:
reducing the amount or concentration of bioactive trimeric sTNF in the blood of a subject having the disease without affecting the amount, concentration or bioactivity of tmTNF by disrupting or reducing the in vivo assembly or reassembly of dissociated monomers of sTNF into bioactive trimeric human sTNF.

16. The method according to claim 15, comprising administering to a subject in need thereof an isolated or synthetic antibody or ligand of any of claims 1 to 10.

17. The method according to claim 15, further comprising maintaining a reduced amount or concentration of bioactive trimeric sTNF in the subject's bloodstream over time.

18. The method according to claim 15, wherein the subject's susceptibility to infection by an intracellular pathogen or to a malignancy is not increased by treatment.

19. The method according to claim 18, wherein the infection is tuberculosis, bacterial sepsis, invasive fungal infection, or histoplasmosis, or wherein the malignancy is lymphoma or hepatosplenic T-cell lymphoma.

20. The method according to claim 15, wherein the antibody or ligand is present in a pharmaceutical composition in a pharmaceutically acceptable carrier or diluent.

21. The method according to any of claims 15-20, wherein the composition is administered intravenously or subcutaneously.

22. The method according to any of claims 15-21, wherein said administration is repeated periodically.

23. The method according to claim 15, wherein said disease is rheumatoid arthritis, ankylosing spondylitis, juvenile rheumatoid arthritis, psoriatic arthritis, psoriasis, obesity, metabolic syndrome, atherosclerosis, associated cardiovascular disease, Alzheimer's disease or a neurodegenerative disease.
24. The method according to claim 15, wherein the disease is a pathogenic effect of bioactive, trimeric sTNF produced during inflammation or during the course of an inflammatory disorder.

25. A method for treating a subject infected with HIV-1 comprising administering to the subject in need thereof an isolated or synthetic antibody or ligand, or pharmaceutical composition, of any of claims 1-14.

26. A method for treating a subject with type II diabetes comprising administering periodically to a subject in need thereof an isolated or synthetic antibody or ligand, or pharmaceutical composition, of any of claims 1-14.

27. The method according to claim 26, wherein the subject is being concurrently treated with other diabetes medication.

28. The method according to claim 27, wherein the administration is repeatedly periodically after the subject ceases treatment with other diabetes medication.

29. A composition that reduces the amount or concentration of bioactive trimeric sTNF in the blood of a subject without affecting the amount, concentration or bioactivity of tmTNF by disrupting or reducing the in vivo assembly or reassembly of dissociated monomers of sTNF into bioactive trimeric human sTNF for treating a subject having a disease mediated by human TNF.

30. The composition of claim 29 comprising an isolated or synthetic antibody or ligand of any of claims 1 to 10.

31. An isolated or synthetic antibody or ligand, or pharmaceutical composition, of any of claims 1-14 for treating a subject infected with HIV-1 or having Type II diabetes.
FIG. 1

TNF Interface A (underlined)

Immunogen:

TNF 1-23 VRSSRTPSD\textbf{KPVAHHVVANPQAE}

Epitopes detected:

A1:  TNF 4-12 SSRTPSD\textbf{KP}
A2:  TNF 8-15 (in rats) PSD\textbf{KPVAH}
A2:  TNF 8-16 (in mouse) PSD\textbf{KPVAHV}

TNF Interface F (underlined)

Immunogen:

TNF 112-128 \textbf{KPWYEPILGGVFQLEK}

Epitope detected: TNF 116-124 \textbf{EPIYLGGVF}
FIG. 2A

Antibody binding epitopes within TNF 1-23 in rat antiserum

Titer

1-10 VSRSSRTP3D 5-12 SRTTPSDKP 411 SRTTPSDKP 8-15 SDKPVAH 8-16 SDKPVAH 8-14 SDKPVVA 16-23 VNRPDRE

TNF detector peptides
FIG. 2B

Epitope A2 in mouse antiserum

TNF detector peptides
FIG. 2C

F epitope margins

<table>
<thead>
<tr>
<th>TNF peptides</th>
<th>Titers</th>
</tr>
</thead>
<tbody>
<tr>
<td>118-124 YVYVGF</td>
<td>10,000</td>
</tr>
<tr>
<td>117-124 YVYVGF</td>
<td>20,000</td>
</tr>
<tr>
<td>116-124 EPYVYVGF</td>
<td>40,000</td>
</tr>
<tr>
<td>116-123 EPYVYVGF</td>
<td>60,000</td>
</tr>
</tbody>
</table>
FIG. 3A

Sandwich of Epitope A1 and A2 Antiserum

---●--- Remicade

---○--- Protein A/G purified IgG from TNF 1-23 immunization (A1/A2)
FIG. 3B

Sandwich assay with 200 ng/mL TNF

- Remicade
- Affinity purified IgG from F antiserum
- Protein A/G purified IgG from F antiserum
FIG. 4A

Epitope A1 and A2 Rat Antiserum to TNF 1-23
Inhibition of TNF Cytotoxicity

P < 0.0001, one way ANOVA

Titers of anti-TNF antiserum with TNF in all wells

Dunnett's post testing

**** P < 0.0001; *** P < 0.001; ** P < 0.01; NS not significant
FIG. 4B

Inhibition of TNF Cytotoxicity in WEHI Cells, MTT Assay, by Antiserum to F Interface Immunogen

P < 0.0001, one way ANOVA

Dunnett's multiple comparisons versus TNF alone

** P < 0.01, ****P < 0.0001
FIG. 5

Mab Inhibition of sTNF cytotoxicity in WEHI cells

OD (450 nm)
INTERNATIONAL SEARCH REPORT

A. CLASSIFICATION OF SUBJECT MATTER

IPC(8) - A61K 39/395; A61K 39/00; C12P 21/08; C07K 16/00 (2014.01)
USPC - 424/145.1; 424/133.1; 530/387.9; 530/388.23; 530/387.1

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC(8) - A61K 39/395; A61K 39/00; C12P 21/08; C07K 16/00 (2014.01)
USPC - 424/145.1; 424/133.1; 530/387.9; 530/388.23; 530/387.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

IPC(8) - A61K 39/395; A61K 39/00; C12P 21/08; C07K 16/00 (2014.01) - see keyword below
USPC - 424/145.1; 424/133.1; 530/387.9; 530/388.23; 530/387.1 - see keyword below

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

PubMed(PCT,PGP,PAP,JPAB); PatBase; Medline; Google: tumor necrosis factor, TNF, sTNF, monomer, trimeric, trimNF, trimer, antibody, anti-TNF, assembling, soluble, immunoglobulin, block, inhibit, antagonist, transmembrane, neutralizing, blood, isolate, purify, epitope, human, treat, pharmaceutical, carrier, diluent, composition, TNF, alpha, dissoci

C. DOCUMENTS CONSIDERED TO BE RELEVANT

<table>
<thead>
<tr>
<th>Category</th>
<th>Citation of document, with indication, where appropriate, of the relevant passages</th>
<th>Relevant to claim No.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Y</td>
<td>US 20110195603 A1 (LE et al) 11 August 2011 (11.08.2011), Abstract, para [0003], [0015], [0071], [0079], [0080], [0096], [0142], [0145], [0173], [0189], [0218], [0235], [0258], [0281], [0296], [0380], [0395]. and SEQ ID NO: 1</td>
<td>1-6, 11, 13-14, 29</td>
</tr>
<tr>
<td>Y</td>
<td>YONE et al Epitropic regions for antibodies against tumor necrosis factor alpha. Analysis by synthetic peptide mapping. J Biol Chem. 1995, Vol. 270(33), p. 19509-15, Abstract: pg 19510, col 2, para 4; pg 19511, col 2, middle para, last para, and Fig 3; pg 19512, col 1, para 2; pg 19513, col 2, last para; and pg 19514, col 1, top para</td>
<td>1-6, 11, 13-14, 29</td>
</tr>
<tr>
<td>A</td>
<td>McCoy et al. TNF signaling inhibition in the CNS: implications for normal brain function and neurodegenerative disease. J Neuroinflammation. 2008, Vol. 5:45, PDF file: pg 1-13. Abstract: pg 1, col 1; and pg 5; Fig 1</td>
<td>1-6, 11, 13-14, 29</td>
</tr>
</tbody>
</table>

Further documents are listed in the continuation of Box C.

Date of the actual completion of the international search 14 March 2014 (14.03.2014)

Date of mailing of the international search report 20 MAY 2014

Name and mailing address of the ISA/US Mail Stop PCT, Attn: ISA/US, Commissioner for Patents P.O. Box 1450, Alexandria, Virginia 22313-1450

Facsimile No. 571-273-3021

Authorized officer: Lee W. Young

PCT Helpdesk: 571-272-4300
PCT OSP: 571-272-7774
INTERNATIONAL SEARCH REPORT

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☒ Claims Nos.: 7-10, 12, 16, 21-22, 25-28, 30-31 because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single general inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees must be paid.

Group I, claims 1-6, 11, 13-14, 29, drawn to an isolated or synthetic antibody or a composition comprising the antibody.

Group II, claims 15, 17-20, and 23-24, drawn to a method reating a subject having a disease mediated by human TNF.

The inventions listed as Groups I-II do not relate to a single general inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons:

Groups I-II are related as a product (Group I) and methods of using the product (Groups II - claim 20 only).

******************************************************************************Continued in the extra sheet******************************************************************************

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.

2. ☐ As all searchable claims could be searched without effort justifying additional fees, this Authority did not invite payment of additional fees.

3. ☒ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-6, 11, 13-14, 29

Remark on Protest

☐ The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.

☐ The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.

☐ No protest accompanied the payment of additional search fees.

Form PCT/ISA/210 (continuation of first sheet (2)) (July 2009)
Continuation of:
Box No III (unity of invention is lacking)

Common Technical Features
The inventions of Groups I to IV share the technical feature of disrupting/reducing assembly of the monomers of sTNF into biotactic trimeric human sTNF.

However, these shared technical features do not represent a contribution over prior art as being obvious over US 2011/0195063 A1 to LE et al. (hereinafter ‘La’), in view of an article entitled ‘Eptopic regions for antibodies against tumor necrosis factor alpha. Analysis by synthetic peptide mapping’ by Yone et al. (hereinafter ‘Yone’), J Biol Chem. 1995, Vol. 270(33), p. 19209-15, and further in view of an article entitled ‘TNF, signaling inhibition in the CNS: implications for normal brain function and neurodegenerative disease’ by McCoy et al. (hereinafter ‘McCoy’); J Neuroimmunol. 2008, Vol. 545. PDF file: pg 1-13 as follows:

Regarding claim 1, 1 discloses an isolated or synthetic antibody or ligand (para [0142] - a human anti-TNF antibody can be isolated; para [0145] - producing or isolating antibodies; para [0188] - ‘synthesis of an anti-TNF peptide or A fragment’; para [0380] - The purified chimeric antibody was evaluated for its binding and inhibiting (and/or neutralizing activity)

- that specifically binds to an epitope of an TNF α of SEQ ID NO: 1 (para [0090] - ‘Preferred antibodies to bind epitope’).
- corresponding amino acids of SEQ ID NO: 1, wherein SEQ ID NO: 1 is 100%, identical to the claimed SEQ ID NO: 1, see claims 2-4; - said binding neutralizing at least one TNF activity including disrupting assembly of the monomer into biotactic trimeric human TNF (para [0078] - ‘Anti-TNF antibodies binds a portion of a TNF and inhibits or neutralizes at least one TNF biological activity’; para [0085] - ‘the chimeric antibody…specific for human TNF’; para [0218] - TNF neutralizing activity - blocking production of TNF by assembly of the biotactic trimer of TNF; wherein ‘blocking production of TNF by assembly of the biotactic trimer of TNF is’ disrupting assembly of the monomer into trimeric human TNF, because activated TNF are in trimer form; Please see McCoy: Abstract, and the discussion that follow. 

Le further discloses antibodies known in the art recognize epitopes including amino acid residues 1-18 and 108-127 of human TNF α, (para [0105] - PCT publication No WO2007/02078) discloses TNF ligands, which can bind to monomeric antibodies having the following epitopes: at least one of 1-10, ‘...10-82...; at 1-10-12; ‘...10-82-128’; para [0173] - Putative receptor binding loci of human TNF have been presented... WO91/02078... discloses TNF ligands which can bind to monomeric antibodies having the following epitopes of at least one of 1-120, ‘...10-127...’ both of which comprise the epitopes in the application (see claims 2-4).

Le does not specifically teach wherein the antibody specifically binds to a dissociated monomer, said binding disrupting assembly of the monomer into biotactic trimeric human sTNF. Yone discloses a method of using a set of overlapping octamer peptides (peptides of 8 a.a.) derived from a human TNF alpha for binding to a anti-TNF polyclonal antibody to determine amino acid residues involved in epitopes of antibodies binding, wherein the epitopes identified comprising amino acid residues of 7-11 and 106-12/98-127 (Abstract - relevant epitopes on human tumor necrosis factor α (TNF-α) was evaluated...against 149 sequential, overlapping octamer peptides), raised against recombinant TNF-α reacted with oligopeptides, corresponding to TNF-α residues 7-11...106-127’; pg 951 1, col. 2, middle para, Fig 3; pg 9512, para 2 - residues 98-127, including a 1F12A7 binding region...106-11...inhibited the binding of IF12A7; Fig 4), and further, wherein octamer peptides of 106-127, which include octamer peptides having amino acid residues of 106-1 13 to octamer peptides having amino acid residues of 112-120 (Fig 3; Legend - ‘The numerical assignment of the peptide is such that peptide number 1 represents residues 1-8, number 2 represents residues 2-9’ recognize a monomeric antibody, which does not neutralize TNF-trimer function, or does not interact with trimer TNF (pg 951 1, col 2, last para - A murine non- neutralizing mAb, 1F12A7, recognized one octapeptide with initial residue 104, one of the six major areas of binding by the polyclonal antibody‘; Fig 3(b); pg 9510, col 2, para 4 - ‘1F12A7 7... bound to TNF-α but did not neutralize its cytolytic activity‘; pg 9513, col 2, last para to pg 9514, col 1, top para - ‘non-neutralizing mAb 1F12A7 recognized a linear epitope...106-11...supported by...a peptide composed of residues 98-127 inhibited the binding of 1F12A7 to rTNF-α...mAb 1F12A7...inhibit binding to its receptor...residues 106-111 are not...involved in the TNF-receptor interaction. The peptide composed of residues 98-127 had neither TNF-α agonistic nor antagonistic activity at concentrations up to 0.15 mM‘; pg 9512, Fig 4), indicating this region comprising (an) epitope(s) that is specific for a dissociated monomer of sTNF (Please see Smith et al: Abstract - The active form of TNF was identified by its inhibitory activity in receptor binding assays...Isolated monomers showed low binding affinity and reduced cytotoxicity, whereas trimers showed high binding affinity...and cytotoxicity’). Yone also does not specifically teach an antibody specifically binds to a dissociated monomer, said binding disrupting assembly of the monomer into biotactic trimeric human sTNF. McCoy discloses elevated level of soluble TNF trimer is a hallmark of a number of neurodegeneration diseases (Abstract - Elevation of sTNF is a hallmark of...neurodegenerative conditions including Alzheimer’s...multiple sclerosis’; pg 1, col 1 - TNF is synthesized...transmembrane protein (mTNF)...as a homotrimer and cleaved...to soluble circulating trimer (sTNF); and a targeted soluble TNF (sTNF) monomer inhibitor alters the assembly of biotactic trimeric human TNF and therefore inhibits biotactic trimeric human sTNF interaction with its receptor (pg 5, Fig 1; and Legend - TNF variants (DN-TNFs) exchange with native sTNF monomers to form heterotrimers with drastically reduced abilities to bind TNF receptors, making them selective for sTNF signaling inhibition’.

Although McCoy also does not specifically teach an antibody specifically binds to a dissociated monomer, said binding said binding disrupting assembly of the monomer into biotactic trimeric human TNF, it provides a motivation to screening or generating an antibody that specifically binds to a dissociated monomer, for inhibiting biotactic trimeric human sTNF, based on the combination of McCoy, Yone and Le, especially Yone discloses a monomeric antibody that binds an epitope between amino acid residues of 106-120 (peptides 106-111 cover amino acid residues 106-120, see discussion above), do not inhibit trimeric TNF cytotoxic function (pg 9510, col 2, para 4 - ‘1F12A7 7...bound to TNF-α but did not neutralize its cytolytic activity‘; pg951 1, Fig 3; pg 9513, col 2, last para to pg 9514, col 1, top para), indicating the epitope more specific for dissociated monomer, and Le discloses antibodies that recognize the similar regions (para [0173] - ‘monoclonal antibodies having the following epitopes at least one of 1-20, ‘...108-127...’ and generating an antibody for disrupting assembly of the monomer into biotactic trimeric human sTNF, especially the peptides taught by Yone comprise the epitopes of the application (Abstract - ‘TNF-α residues 7-11...106-127’, wherein ‘TNF-α residues 7-11 include peptides having amino acid residues of 7-14, 8-15...’, and 11-18 of hTNF-α, wherein peptide 8-15 comprising the sequence of ‘PSDKPKVH’ in claim 2, based on the combination of Yone and Le, and further wherein hTNF-α...106-112’ comprising peptide 112-120, as discussed above, which is in the range of the epitope 106-127 taught by Le, and comprises the epitope 116-124 in claim 4 in the application; Please further see pg 950 1, Figs 3; Legend - explanation for the numbering of peptides).
Continuation of:
The previous extra sheet - Box No III (unity of invention is lacking)

It would have been obvious to one of ordinary skill in the art at the time the invention was made to combine the teachings of Le, Yone, and McCoy, to obtain an isolated or synthetic antibody or ligand that specifically binds to an epitope of human TNF, said binding neutralizing at least one TNF activity including disrupting assembly of the monomer into bioactive trimeric human TNF, based on the teaching of Le, and further wherein the antibody specifically binds to a dissociated monomer, said binding disrupting assembly of the monomer into bioactive trimeric human sTNF, based on the combination of McCoy, Yone, and Le, and inherent properties of antibodies that bind to an epitope selected from the group consisting of: 1) PSDKPVAH, amino acids 8-15 of SEQ ID NO: 1, which is disclosed by the combination of Le and Yone, and 2) 116-124 of the SEQ ID NO: 1, based on the combination of Le and Yone with a further epitope mapping, in order to selectively inhibit sTNF function for treating disorders associated with increased level of bioactive trimeric human sTNF with expected success without undue experimentation.

Without a shared special technical feature, the inventions lack unity with one another.

Groups I-II therefore lack unity under PCT Rule 13 because they do not share a same or corresponding special technical feature.

Note re item 4. Claims 7-10, 12, 16, 21-22, 25-28, 30-31 are not drafted in accordance with the second and third sentences of Rule 6.4 (a). These claims are improper multiple dependent claims.

Note:
Claim 20 is objected to as lacking a proper antecedent basis for the "the antibody or ligand" limitation. For the purposes of this ISR, claim 20 is construed as follows:

20. The method according to claim 15, further comprising administering an antibody or ligand, wherein the antibody or ligand is present in a pharmaceutical composition in a pharmaceutically acceptable carrier or diluent.