USE OF INTERLEUKIN-4 RECEPTOR ANTIBODIES AND COMPOSITIONS THEREOF

VERWENDUNG VON INTERLEUKIN-4 REZEPTOR ANTIKÖRPERN UND ZUSAMMENSETZUNGEN DAVON

UTILISATION D’ANTICORPS DU RECEPTEUR DE L’INTERLEUKINE-4 ET LEURS COMPOSITIONS

Designated Contracting States:
AT CH CY DE DK ES FI FR GR IE IT LI LU MC NL PT SE TR

Priority:
26.05.2000 US 579808
19.09.2000 US 665343
15.02.2001 US 785934
01.05.2001 US 847816

Date of publication of application:
02.03.2016 Bulletin 2016/09

Document number(s) of the earlier application(s) in accordance with Art. 76 EPC:
10185626.8 / 2 292 665
01952133.5 / 1 283 851

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BACKGROUND OF THE INVENTION

[0001] Interleukin-4 (IL-4), previously known as B cell stimulating factor, or BSF-1, was originally characterized by its ability to stimulate the proliferation of B cells in response to low concentrations of antibodies directed to surface immunoglobulin. IL-4 has been shown to possess a far broader spectrum of biological activities, including growth co-stimulation of T cells, mast cells, granulocytes, megakaryocytes, and erythrocytes. In addition, IL-4 stimulates the proliferation of several IL-2- and IL-3-dependent cell lines, induces the expression of class II major histocompatibility complex molecules on resting B cells, and enhances the secretion of IgE and IgG1 isotypes by stimulated B cells. IL-4 is associated with a TH2-type immune response, being one of the cytokines secreted by TH2 cells.

[0002] Murine and human IL-4 have been identified and characterized, including cloning of IL-4 cDNAs and determination of the nucleotide and encoded amino acid sequences. (See Yokota et al., Proc. Natl. Acad. Sci. USA 83:5894, 1986; Noma et al., Nature 319:640, 1986; Grabstein et al., J. Exp. Med. 163:1405, 1986; and U.S. Patent 5,017,691.)

[0003] IL-4 binds to particular cell surface receptors, which results in transduction of a biological signal to cells such as various immune effector cells. IL-4 receptors are described, and DNA and amino acid sequence information presented, in Mosley et al., Cell 59:335-348, October 20, 1989 (murine IL-4R); Idzerda et al., J. Exp. Med. 171:861-873, March 1990 (human IL-4R); and U.S. Patent 5,599,905. The IL-4 receptor described in these publications is sometimes referred to as IL-4Rα.

Other proteins have been reported to be associated with IL-4Rα on some cell types, and to be components of multi-subunit IL-4 receptor complexes. One such subunit is IL-2Rγc, also known as IL-2RγC. (See the discussion of IL-4R complexes in Sato et al., Current Opinion in Cell Biology, 6:174-179, 1994.) IL-4Rα has been reported to be a component of certain multi-subunit IL-13 receptor complexes (Zurawski et al., J. Biol. Chem. 270 (23), 13869, 1995; de Vries, J. Allergy Clin. Immunol. 102(2):165, August 1998; and Callard et al. Immunology Today, 17(3):108, March 1996).

[0005] IL-4 has been implicated in a number of disorders, examples of which are allergy and asthma. Studies of biological properties of IL-4 continue, in an effort to identify additional activities associated with this pleiotrophic cytokine, and to elucidate the role IL-4 may play in various biological processes and diseases.

SUMMARY OF THE INVENTION

[0006] In a first aspect, the present invention provides a human monoclonal antibody capable of inhibiting an IL-4 induced biological activity that competes with a reference antibody for binding to a cell that expresses human IL-4 receptor (IL-4R), wherein:

a) the light chain of the reference antibody comprises the sequence of SEQ ID NO:6 and the heavy chain of the reference antibody comprises the sequence of SEQ ID NO:8; or
b) the light chain of the reference antibody comprises the sequence of SEQ ID NO:26 and the heavy chain of the reference antibody comprises the sequence of SEQ ID NO:24.

[0007] The antibody may be additionally capable of inhibiting an IL-13- induced biological activity.

[0008] In a second aspect, the present invention provides the antibody of the first aspect for use in a method of treating dermatitis, atopic dermatitis, contact dermatitis, or asthma. The antibody for may be in the form of a composition that additionally comprises a diluent, excipient, or carrier.

[0009] The IL-4R may comprise amino acids 1 to 197 of SEQ ID NO:2.

[0010] In a third aspect, the present invention provides a nucleic acid encoding the light chain of the antibody of the first aspect and, in a fourth aspect, the invention provides a nucleic acid encoding the heavy chain of the antibody of the first aspect. The invention also provides, in a fifth aspect, a host cell comprising the nucleic acid of the third or fourth aspects.

BRIEF DESCRIPTION OF THE DRAWINGS

[0011] FIGURES 1A-C present the nucleotide sequence of the coding region of a human IL-4 receptor cDNA. The amino acid sequence encoded by the cDNA is presented as well. The cDNA clone was isolated from a cDNA library derived from a human T cell line T22. The encoded protein comprises (from N- to C-terminus) an N-terminal signal peptide, followed by an extracellular domain, a transmembrane region (underlined), and a cytoplasmic domain, as discussed further below. The DNA and amino acid sequences of Figures 1A to 1C are also presented in SEQ ID NOS:1 and
The present invention provides a human monoclonal antibody capable of inhibiting an IL-4 induced biological activity and the use thereof in methods for treating certain conditions induced by IL-4, and for inhibiting biological activities of interleukin-4 (IL-4) in vivo. One method comprises administering such an IL-4 antagonist antibody to a patient afflicted with such a condition. Compositions for use in such methods for treating IL-4-induced conditions also are provided.

Among the conditions that may be treated in accordance with the present invention are lung disorders in which IL-4 plays a role, conditions in which IL-4-induced epithelial barrier disruption plays a role in particular contact dermatitis, and atopic dermatitis.

The present invention provides the human monoclonal antibody of the invention for use in methods comprising the binding of IL-4 to a native IL-4 receptor on a cell, or which may be inhibited or suppressed by preventing IL-4 from binding to an IL-4 receptor. Conditions that may be treated include, but are not limited to, medical disorders that play a role in the particular disease to be treated.

Indications

The present invention provides the human monoclonal antibody of the invention for use in methods comprising administering the antibody to a patient afflicted with any of a number of conditions induced by IL-4 biological activity. IL-4-induced conditions include conditions caused or exacerbated, directly or indirectly, by IL-4. Other factors or cytokines also may play a role in such conditions, but IL-4 induces or mediates the condition to some degree, i.e., at least in part.

The biological activities of IL-4 are mediated through binding to specific cell surface receptors, referred to as interleukin-4 receptors (IL-4R). IL-4-induced conditions include those arising from biological responses that result from the binding of IL-4 to a native IL-4 receptor on a cell, or which may be inhibited or suppressed by preventing IL-4 from binding to an IL-4 receptor. Conditions that may be treated include, but are not limited to, medical disorders characterized by abnormal or excess expression of IL-4, or by an abnormal host response to IL-4 production. Further examples are conditions in which IL-4-induced antibody production, or proliferation or influx of a particular cell type, plays a role. IL-4-induced disorders include those in which IL-4 induces upregulation of IL-4 receptors or enhanced production of another protein that plays a role in a disease (e.g., another cytokine).

The antibody may be for use in a method for treating a mammal, including a human patient, who has such a medical disorder comprising administering the antibody to the mammal or otherwise contacting endogenous IL-4 with the antibody, e.g., in an ex vivo procedure. Conditions that may be treated in accordance with the present invention include, but are not limited to, conditions in which IL-4-induced barrier disruption plays a role (e.g., conditions characterized by decreased epithelial barrier function in the lung or dermatitis, contact dermatitis or atopic dermatitis, IL-4-induced pulmonary conditions (including asthma). IL-4 antagonists also find use as adjuvants to allergy immunotherapy and as vaccine adjuvants, especially when directing the immune response toward a TH1 response would be beneficial in treating or preventing the disease in question.
Septic/reactive arthritis

[0019] IL-4 antagonists may be employed in treating septic arthritis, which also is known as reactive arthritis or bacterial arthritis. Septic arthritis can be triggered by (result from, or develop subsequent to) infection with such microbes as Staphylococcus aureus, Chlamydia trachomatis, Yersinia e.g., Y. enterocolitica, Salmonella, e.g., S. enteritidis, Shigella and Campylobacter. S. aureus has been reported to be the major human pathogen in septic arthritis, responsible for the majority of cases.

[0020] IL-4 and IL-4-dependent Th2 responses play roles in promoting septic arthritis. IL-4 antagonist(s) may be employed to inhibit IL-4 and also to suppress the Th2 response in patients having septic arthritis or at risk for developing septic arthritis.

[0021] IL-4 increases bacterial burden and bacterial persistence in joints, by inhibiting clearance of the bacteria. IL-4 antagonists may be employed to assist in the clearance of bacteria associated with reactive arthritis, thereby reducing clinical manifestations such as swelling in joints. IL-4 antagonists may be administered to a human patient afflicted with septic arthritis, to reduce IL-4-mediated joint inflammation. In one approach, an antagonist is injected into a joint, e.g., into synovial fluid in the knee.

[0022] The use of IL-4 antagonists may benefit patients having (or at risk for) septic arthritis by suppressing a TH2 response and promoting a TH1 response against the infection. TH2 cytokines may contribute to bacterial persistence in the joint, whereas a TH1 response plays a role in eliminating the bacteria.

[0023] The antagonists may be administered to patients infected with bacteria or other microbes such as those listed above, to prevent development of septic arthritis. Antagonist(s) may be administered after diagnosis with such an infection, but before development of clinical symptoms of septic arthritis.

Whipple’s Disease

[0024] Tropheryma whippelii is the causative bacterium for Whipple’s Disease, also known as intestinal lipodystrophy and lipophagia granulomatosis. The disease is characterized by steatorrhea, frequently generalized lymphadenopathy, arthritis, fever, and cough. Also reported in Whipple’s Disease patients are an abundance of “foamy” macrophages in the jejunal lamina propria, and lymph nodes containing periodic acid-schiff positive particles appearing bacilliform by electron microscopy (Steadman’s Medical Dictionary, 26th Edition, Williams & Wilkins, Baltimore, MD, 1995).

[0025] The use of IL-4 antagonist(s) may benefit patients having (or at risk for developing) Whipple’s Disease, by restoring a normal balance between the TH1 and TH2 components of the patient’s immune response. Increased production of IL-4 (a TH2-type cytokine) and decreased levels of certain TH1-type cytokines have been associated with Whipple’s Disease. TH2 cytokines may contribute to bacterial persistence, whereas a TH1 response plays a role in clearing the causative bacteria. IL-4 antagonists may be administered to patients infected with T. whippelii, whether or not the patient exhibits clinical symptoms of Whipple’s Disease.

Dermatitis herpetiformis

[0026] Dermatitis herpetiformis, also known as Duhring’s disease, is a chronic skin condition characterized by blistering skin lesions, cutaneous IgA deposits, and itching. Patients have an immunobullous skin disorder with an associated gluten sensitive enteropathy, which is mediated by a Th2 immune response. IL-4 antagonist(s) may be administered to inhibit IL-4 and the Th2 response, thus promoting healing of current lesions and reducing or preventing the formation of blisters on the extensor body surfaces.

Hypertrophic scarring

[0027] IL-4 antagonist(s) may be administered to patients who have, or are susceptible to developing, hypertrophic scarring. In one method, an IL-4 antagonist is administered to a burn patient. An immune response to burns and other injury is believed to play a role in the pathogenesis of hypertrophic scarring. Increased production of TH2-type cytokines, including IL-4, and reduced levels of certain TH1-type cytokines have been reported in burn patients who have hypertrophic scarring. The use of IL-4 antagonists may benefit patients having (or at risk for developing) hypertrophic scarring, by suppressing a TH2-type immune response.

Urticaria

[0028] Urticaria, especially chronic forms thereof such as chronic idiopathic urticaria (CIU), may be treated with an IL-4 antagonist. CIU patients have higher serum levels of IL-4 than controls, and may have a predominantly TH2-type cytokine profile. Mast cells and Th2-type T cells are implicated as primary effector cells in chronic urticaria. IL-4 stimulates
mast cell proliferation. Mast cell degranulation leads to histamine release, subsequent erythema, eosinophilia, redness of skin, and itching. IL-4 antagonists are administered to inhibit IL-4 and reduce the TH2-type response, thereby helping to control a patient’s urticaria.

Ulcereative colitis; other disorders of the gastrointestinal tract

0029 IL-4 is implicated in the pathogenesis of ulcerative colitis. Th2-type cytokines including IL-4 may predominate in the colonic mucosa of patients with this disorder. The use of IL-4 antagonist(s) to suppress the TH2 response may alleviate this condition.

0030 In addition to ulcerative colitis, other disorders of the gastrointestinal tract or digestive system may be treated with IL-4 antagonist(s). Examples of such disorders include, but are not limited to, inflammatory bowel disease (IBD), with ulcerative colitis and Crohn’s Disease being forms of IBD, gastritis, ulcers, and mucosal inflammation.

0031 Any gastrointestinal condition in which IL-4 plays a role may be treated with an IL-4 antagonist. For example, conditions involving IL-4-induced inflammation of part of the gastrointestinal tract may be treated with an IL-4 antagonist. Particular examples described herein are directed to treatment of chronic inflammatory conditions in the gastrointestinal tract.

0032 Other examples described herein are directed to conditions in which IL-4-induced barrier disruption plays a role, e.g., conditions characterized by decreased epithelial barrier function in at least a portion of the gastrointestinal tract. Such conditions may, for example, involve damage to the epithelium that is induced by IL-4, directly or indirectly.

0033 The intestinal epithelium forms a relatively impermeable barrier between the lumen and the submucosa. Disruption of the epithelial barrier has been associated with conditions such as inflammatory bowel disease. See the discussion in Youakim, A. and M. Ahdieh (Am. J. Physiol. 276 (Gastrointest. Liver Physiol. 39): G1279-G1288, 1999). A damaged or "leaky" barrier can allow antigens to cross the barrier, which in turn elicits an immune response that may cause further damage to gastrointestinal tissue. Such an immune response may include recruitment of neutrophils or T cells, for example. An IL-4 antagonist may be administered to inhibit undesirable stimulation of an immune response.

Lung disorders

0034 Methods for treating IL-4-induced pulmonary disorders are described herein. Such disorders include, but are not limited to, lung fibrosis, including chronic fibrotic lung disease, other conditions characterized by IL-4-induced fibroblast proliferation or collagen accumulation in the lungs, pulmonary conditions in which a TH2-type immune response plays a role, conditions characterized by decreased barrier function in the lung (e.g., resulting from IL-4-induced damage to the epithelium), or conditions in which IL-4 plays a role in an inflammatory response.

0035 Cystic fibrosis is characterized by the overproduction of mucus and development of chronic infections. Inhibiting IL-4 and the TH2 response will reduce mucus production and help control infections such as allergic bronchopulmonary aspergillosis (ABPA).

0036 Allergic bronchopulmonary mycosis occurs primarily in patients with cystic fibrosis or asthma, where a TH2 immune response is dominant. Inhibiting IL-4 and the TH2 response will help clear and control these infections.

0037 Chronic obstructive pulmonary disease is associated with mucus hypersecretion and fibrosis. Inhibiting IL-4 and the TH2 response will reduce the production of mucus and the development of fibrous thereby improving respiratory function and delaying disease progression.

0038 Bleomycin-induced pneumopathy and fibrosis, and radiation-induced pulmonary fibrosis are disorders characterized by fibrosis of the lung which is manifested by the influx of Th2, CD4+ cells and macrophages, which produce IL-4 which in turn mediates the development of fibrosis. Inhibiting IL-4 and the TH2 response will reduce or prevent the development of these disorders.

0039 Pulmonary alveolar proteinosis is characterized by the disrupation of surfactant clearance. IL-4 increases surfactant product. Use of IL-4 antagonists will decrease surfactant production and decrease the need for whole lung lavage.

0040 Adult respiratory distress syndrome (ARDS) is a frequent complication in such patients. IL-4 antagonists may alleviate ARDS by reducing inflammation and adhesion molecules, although methods for treating such patients are not limited by a particular mechanism of action. IL-4 antagonists may be used to prevent or treat ARDS.

0041 Sarcoidosis is characterized by granulomatous lesions. Use of IL-4 antagonists to treat sarcoidosis, particularly pulmonary sarcoidosis, is contemplated herein.

0042 Conditions in which IL-4-induced barrier disruption plays a role (e.g., conditions characterized by decreased epithelial barrier function in the lung) may be treated with IL-4 antagonist(s). Damage to the epithelial barrier in the lungs may be induced by IL-4 directly or indirectly. The epithelium in the lung functions as a selective barrier that prevents contents of the lung lumen from entering the submucosa. A damaged or "leaky" barrier allows antigens to cross the...
barrier, which in turn elicits an immune response that may cause further damage to lung tissue. Such an immune response may include recruitment of eosinophils or mast cells, for example. An IL-4 antagonist may be administered to inhibit such undesirable stimulation of an immune response.

IL-4 antagonists may be employed to promote healing of lung epithelium, thus restoring barrier function. IL-4 antagonists may be employed to promote healing of lung epithelium in asthmatics, for example. Alternatively, the antagonist is administered for prophylactic purposes, to prevent IL-4-induced damage to lung epithelium.

**Tuberculosis**

A TH2-type immune response is implicated in playing a role in causing tissue damage (e.g., necrosis of lung tissue) in tuberculosis (TB) patients. Elevated levels of IL-4 are associated with TB. IL-4 production may be particularly elevated in cavitary tuberculosis (i.e., in TB patients who have developed pulmonary cavities, which can be detected/visualized by such techniques as radiographs of the chest).

IL-4 antagonists may benefit TB patients (especially those with cavitary TB) by suppressing a TH2-type immune response, or by binding (and inactivating) excess secreted IL-4. Methods for treating such patients are not limited by a particular mechanism of action, however. IL-4 antagonists advantageously may be administered in an amount that restores the desired balance between the TH1 and TH2 components of the immune response, and reduces IL-4-induced tissue damage in a patient.

**Churg-Strauss syndrome**

Churg-Strauss syndrome, a disease also known as allergic granulomatous angitis, is characterized by inflammation of the blood vessels in persons with a history of asthma or allergy, and by eosinophilia. IL-4 antagonist(s) may be administered to alleviate inflammation in patients with this syndrome. The use of IL-4 antagonists to suppress a TH2-type immune response, and to combat eosinophilia, would benefit the patients.

**Pre-eclampsia**

Pre-eclampsia is a toxemia of late pregnancy. The condition is characterized by a sharp rise in blood pressure, generally accompanied by edema and albuminuria, during the third term of pregnancy.

Elevated TH1-type and TH2-type immune responses may play a role in the condition. One method described herein comprises administering an IL-4 antagonist to a pregnant woman who has developed pre-eclampsia. The IL-4 antagonist is administered in an amount, and for a period of time, sufficient to reduce the level of IL-4 (or of TH2-type cytokines collectively) to a level that is considered normal during pregnancy. In general, the IL-4 antagonist is administered repeatedly throughout the duration of the pregnancy.

**Scleroderma**

IL-4 antagonist(s) may be administered to scleroderma patients. The antagonists reduce IL-4-induced collagen synthesis by fibroblasts in the patients. The antagonists may be employed in preventing or reducing fibrosis in skin and lung tissues, as well as other tissues in which fibrosis occurs in scleroderma patients, suppressing collagen synthesis in such tissues, and in treating scleroderma-related pulmonary disease.

**Benign Prostate Hyperplasia**

Benign prostate hyperplasia (BPH), also known as benign prostate hypertrophy, may be treated with IL-4 antagonist(s). While not wishing to be bound by a particular mechanism of action, administration of an IL-4 inhibitor may benefit a patient with BPH by suppressing IL-4-induced inflammation, or by suppressing a TH2-type immune response.

**Grave’s Disease**

Antibodies directed against thyrotropin receptor play an important role in Grave’s Disease, a disorder characterized by hyperthyroidism. Studies of cytokine production in Grave’s Disease patients show a shift toward a TH2-type cytokine response. Use of an IL-4 antagonist to suppress the TH2-type immune response, and suppress antibody production, would benefit Grave’s Disease patients.
Sickle Cell Disease

[0052] Sickle cell disease patients typically experience intermittent periods of acute exacerbation called crises, with the crises being categorized as anemic or vaso-occlusive. IL-4 antagonists find use in treating or preventing sickle cell crisis, especially in patients with elevated IL-4 levels or in whom the immune response has shifted toward a TH2-type response. Sickle cell disease (especially sickle cell crisis) has been associated with increased susceptibility to infectious diseases, including bacterial infections. Administering IL-4 antagonists to sickle cell disease patients may help the patient mount an immune response against infectious diseases.

Sjogren’s syndrome

[0053] The autoimmune disease known as Sjogren’s syndrome or sicca syndrome typically combines dry eyes and dry mouth with a disorder of the connective tissues, such as rheumatoid arthritis, lupus, scleroderma, or polymyositis. The vast majority of patients are middle age (or older) females. Sjogren’s syndrome is an inflammatory disease of glands (e.g., lacrimal and salivary glands) and other tissues of the body. The syndrome typically is associated with autoantibody production.

[0054] IL-4 antagonists may be administered to reduce the inflammatory response (such as inflammation of glands, including lacrimal glands) in such patients. IL-4 antagonists may benefit Sjogren’s syndrome patients by suppressing a TH2-type immune response, or by binding (and inactivating) excess IL-4 at inflammatory lesions. Methods for treating patients are not limited by a particular mechanism of action, however.

Autoimmune lymphoproliferative syndrome

[0055] Manifestations of autoimmune lymphoproliferative syndrome include lymphoproliferation and autoantibody production. Patients with the syndrome reportedly have an inherited deficiency in apoptosis. IL-4 antagonists may benefit patients with this syndrome by suppressing a TH2-type immune response, or by binding (and inactivating) excess IL-4 at sites of inflammation. Methods for treating such patients are not limited by a particular mechanism of action, however.

Autoimmune hemolytic anemia

[0056] Excessive IL-4 secretion, and a deficiency in TH1-type cytokines, are implicated in contributing to the pathogenesis of autoimmune hemolytic anemia. IL-4 antagonists may be administered to benefit the patients by reducing autoantibody production, and by restoring a more normal balance between the TH1 and TH2 components of the immune response.

Autoimmune uveitis

[0057] Uveitis involves inflammation of the uvea (generally considered to include the iris, ciliary body, and choroid, considered together). Excess IL-4 secretion is implicated as playing a role in pathogenesis of this sight-threatening inflammatory eye disease. IL-4 antagonist(s) may be administered to a uveitis patient to reduce disease severity. In one embodiment, IL-4 antagonist(s) may be administered to an individual who has autoimmune uveoretinitis.

Kawasaki Disease

[0058] Also known as the mucocutaneous lymph node syndrome, Kawasaki disease (KD) mainly afflicts young children. The disease is characterized by particular changes in the mucus membranes lining the lips and mouth, and by enlarged, tender lymph glands. Symptoms typically include fever, conjunctivitis, inflammation of the lips and mucous membranes of the mouth, swollen glands in the neck, and a rash covering the hands and feet, leading to hardened, swollen and peeling skin on hands and feet. In children with Kawasaki Disease (KD), inflammation of arteries (vasculitis) may develop. Due to the effect of the disease on the vascular system, KD reportedly is the main cause of acquired heart disease in children.

[0059] IL-4 antagonists may be administered to patients with Kawasaki Disease, to reduce the elevated levels of IL-4 in the patient. Excessive IL-4 secretion and a deficiency in TH1-type cytokines contribute to the pathogenesis of the disease.

Barrett’s esophagus

[0060] Barrett’s esophagus is a condition characterized by alteration (subsequent to irritation) of the cells in the epithelial
tissue that lines the lower portion of the esophagus. Frequent reflux of the stomach contents into the esophagus, over time, can lead to Barrett esophagus. Patients with Barrett esophagus are at risk for developing esophageal cancer (e.g., adenocarcinoma). While not wishing to be bound by a particular mechanism of action, administration of an IL-4 antagonist may benefit a patient with Barrett’s esophagus by suppressing a TH2-type immune response. An IL-4 antagonist may be administered to a patient with esophagitis, to inhibit progression to Barrett’s esophagus.

Nephrosis

[0061] Nephrosis, also known as nephrotic syndrome, is kidney disease that is non-inflammatory and non-malignant. In the condition known as minimal change nephrosis, glomerular damage (believed to arise from structural changes in glomerular visceral epithelial cells) results in abnormalities that include proteinuria. A TH2-type immune response (especially secretion of the TH2-type cytokines IL-4 and IL-13) are implicated as playing a role in pathogenesis of minimal change nephrosis.

Other indications

[0062] Additional examples of conditions that may be treated include but are not limited to the following. IL-4 antagonists may be employed in treating or preventing hyper IgE syndrome, idiopathic hypereosinophil syndrome, allergic reactions to medication, autoimmune blistering diseases (e.g., pemphigus vulgaris or bullous pemphigoid), myasthenia gravis (an autoimmune muscular disease), and chronic fatigue syndrome. IL-4 inhibitors may be employed in treating GVHD; particular methods for treating GVHD combination therapy with other therapeutic agents as described below. IL-4 inhibitors also find use in treating or preventing hepatotoxicity induced by drugs such as diclofenac (a non-steroidal anti-inflammatory drug).

[0063] An IL-4 antagonist may be employed as an adjuvant to allergy immunotherapy treatment. IL-4 antagonists find further use as vaccine adjuvants, such as adjuvants for cancer vaccines and infectious disease vaccines. The use of IL-4 antagonists is especially advantageous when favoring a TH1-type immune response would be beneficial in preventing or treating the condition for which the vaccine is being administered. IL-4 antagonists may be employed when reducing an antibody-mediated immune response and/or promoting a T-cell-mediated immune response is desired.

IL-4 Antagonists

[0064] IL-4 antagonists that may be employed include compounds that inhibit a biological activity of IL-4. The IL-4-induced biological activities to be inhibited by the antibodies of the invention, for use in methods provided herein are activities that directly or indirectly play a role in the condition to be treated.

[0065] Examples of IL-4 antagonists described herein include, but are not limited to, IL-4 receptors (IL-4R), antibodies, other IL-4-binding molecules, and IL-4 muteins as discussed further below. The antibodies may bind IL-4 or may bind an IL-4 receptor, for example.

[0066] Antagonists that bind IL-4 include but are not limited to IL-4 receptors and anti-IL-4 antibodies. Endogenous IL-4 that becomes bound to such an antagonist is thereby prevented from binding its natural receptor on cell surfaces in vivo, and thus cannot manifest IL-4-mediated biological activities.

[0067] Different types of antagonists may act at different sites or by different mechanisms of action. Examples include but are not limited to antagonists that interfere with binding of IL-4 to cell surface receptors or that inhibit signal transduction. The site of action may be intracellular (e.g., by interfering with an intracellular signaling cascade), on a cell surface, or extracellular. Antagonists that act by interfering with the interaction of IL-4 with IL-4R may bind to either IL-4 or to the receptor. An antagonist need not completely inhibit an IL-4 induced activity to find use in the present invention; rather, antagonists that reduce a particular activity of IL-4 are contemplated for use as well.

[0068] The above-presented discussions of particular mechanisms of action for IL-4 antagonists in treating particular diseases are illustrative only, and the methods presented herein are not bound thereby. The mechanisms of action by which IL-4 antagonists ameliorate diseases are not limited to those discussed above.

[0069] In treating particular disorders, an IL-4 antagonist may reduce the amount of active IL-4 at a particular site within the body that is involved in the disorder. Antagonists that bind IL-4 such that it no longer can bind to endogenous cellular receptors functionally reduce the amount of active IL-4 available for inducing biological responses.

[0070] An IL-4 antagonist may alleviate a disorder by reducing the amount of free endogenous IL-4 that is circulating in the body, e.g., in the bloodstream or in a particular tissue. When the action of IL-4 on such tissue plays a role in pathogenesis of the disease, the antagonist serves to block action of IL-4 in the tissue, thereby alleviating the disorder. In a further example, antagonists may inhibit IL-4-induced recruitment of cells to a site or tissue within the body, wherein such recruitment plays a role in causing or exacerbating a disease. The antagonists may inhibit an IL-4-mediated influx of cells involved in an immune or inflammatory response. An antagonist may act by reducing proliferation, activation,
migration, influx, or accumulation of a particular cell type, or by inhibiting a biological response directly or indirectly attributable to a particular cell type. Examples of particular cell types are fibroblasts, mast cells, and eosinophils.  

As discussed above, some conditions may be treated by suppressing a TH2-type immune response. IL-4 is associated with a TH2 response, and is one of the cytokines secreted by T-helper cells of type 2 (TH2 cells). An IL-4 antagonist may be administered to reduce a TH2-type immune response. The IL-4 antagonist may be said to reduce proliferation of TH2 cells, to suppress a TH2 response, to shift the immune response toward a TH1 response, or to favor a TH1-type response. The use of antagonists of other cytokines associated with a TH2-type immune response is discussed below. Antagonists of other TH2-type cytokine(s), such as IL-5, IL-10, or IL-13, may be administered to patients who have a disorder involving elevated levels of such cytokines. Techniques for measuring the amount of such cytokines in a patient, e.g., in the patient’s serum, are well known.  

One embodiment of the invention may be directed to the antibody of the invention for use in a method for inhibiting IL-4-induced damage to epithelium, comprising administering the antibody to an individual who has, or is at risk of developing, a condition in which IL-4-mediated epithelial barrier disruption plays a role. Barrier function studies revealed that IL-4 plays a role in reduction of barrier function in models of lung epithelium, and that a soluble human IL-4 receptor polypeptide (an IL-4 antagonist) inhibits the IL-4-mediated reduction of barrier function (see example 7).  

Particular embodiments of methods described herein comprise administering an IL-4 antagonist to inhibit IL-4-induced damage to epithelium in the gastrointestinal tract or lung. Such methods may be employed to prevent epithelial damage, or to restore epithelial barrier function (i.e., promote repair or healing of the epithelium). The ability of an IL-4 antagonist to inhibit IL-4-induced damage to epithelium may be confirmed in any of a number of suitable assays, such as those described in example 7 below.  

Any inflammation associated with (or subsequent to) an infection also may be treated with an IL-4 antagonist. The antagonist may be administered to inhibit any IL-4-induced component of an inflammatory response resulting from microbial infection in the gastrointestinal tract, for example.  

Combinations of two or more antagonists may be employed in methods and compositions described herein. Examples of IL-4 antagonists are as follows.  

### IL-4 Receptor  

An preferred IL-4 antagonist is an IL-4 receptor (IL-4R). When administered in vivo, IL-4R polypeptides circulate in the body and bind to circulating endogenous IL-4 molecules, preventing interaction of IL-4 with endogenous cell surface IL-4 receptors, thus inhibiting transduction of IL-4-induced biological signals.  

IL-4 receptors are described in U.S. Patent 5,599,905; Idzerda et al., J. Exp. Med. 171:861-873, March 1990 (human IL-4R); and Mosley et al., Cell 59:335-348, October 20, 1989 (murine IL-4R). The protein described in those three references is sometimes referred to in the scientific literature as IL-4Rx. Unless otherwise specified, the terms "IL-4R" and "IL-4 receptor" as used herein encompass this protein in various forms that are capable of functioning as IL-4 antagonists, including but not limited to soluble fragments, fusion proteins, oligomers, and variants that are capable of binding IL-4, as described in more detail below.  

The nucleotide sequence of a human IL-4R cDNA, and the amino acid sequence encoded thereby, are set forth in Figures 1A-1C. The cDNA clone was isolated from a cDNA library derived from a C57BL/10 T cell clone designated T22, as described in Idzerda et al., J. Exp. Med., 171:861, March 1990, and in U.S. Patent 5,599,905. The DNA and amino acid sequences of Figures 1A-1C are presented in SEQ ID NO:1 and SEQ ID NO:2, respectively.  

The encoded human IL-4R protein comprises (from N- to C-termius) an N-terminal signal peptide, followed by an extracellular domain, a transmembrane region, and a cytoplasmic domain. The transmembrane region, which is underlined in Figure 1A, corresponds to amino acids 208 through 231. The cytoplasmic domain comprises amino acids 232 through 800.  

A signal peptide includes amino acids -25 to -1 of SEQ ID NO:2. An alternative signal peptide cleavage site occurs between residues -3 and -2 of SEQ ID NO:2, such that the signal peptide corresponds to residues -25 through -3.  

As is recognized in the pertinent field, the signal peptide cleavage site for a given protein may vary according to such factors as the particular expression system (especially the host cells) in which the protein is expressed. The exact boundaries of the signal peptide, and thus the extracellular domain, of a given recombinant protein thus may depend on the expression system employed. Further, the signal peptide may be cleaved at more than one position, generating more than one species of polypeptide in a preparation of recombinant protein.  

In one example, in which an expression vector comprises DNA encoding amino acids -25 through 207 of SEQ ID NO:2, the expressed recombinant IL-4R includes two species of mature soluble human IL-4R. The expressed polypeptides include a major species corresponding to amino acids -2 to 207 and a minor species corresponding to amino acids 1 to 207 of SEQ ID NO:2. Two alternate forms of the extracellular domain of human IL-4R thus correspond to residues -2 to 207 and 1 to 207 of SEQ ID NO:2. The term “mature” refers to a protein in a form lacking a signal peptide or leader sequence, as is understood in the pertinent art.
Among the IL-4 receptors suitable for use as described herein are IL-4R fragments. Truncated IL-4R polypeptides may occur naturally, e.g., as a result of proteolytic cleavage, post-translational processing, or alternative splicing of mRNA. Alternatively, fragments may be constructed by deleting terminal or internal portions of an IL-4R sequence, e.g., via recombinant DNA technology. Fragments that retain the ability to bind IL-4 may be identified in conventional binding assays. Such fragments may be soluble fragments, as discussed below.

In an example, the antagonist comprises a soluble form of the IL-4R. A soluble IL-4 receptor is a polypeptide that is secreted from the cell in which it is expressed, rather than being retained on the cell surface. The full length human IL-4R protein of SEQ ID NO:2 is a transmembrane protein, which, as described above, comprises an N-terminal signal peptide, followed by an extracellular domain, a transmembrane region, and a C-terminal cytoplasmic domain. Soluble IL-4R polypeptides lack the transmembrane region that would cause retention on the cell, and the soluble polypeptides consequently are secreted into the culture medium. The transmembrane region and intracellular domain of IL-4R may be deleted or substituted with hydrophilic residues to facilitate secretion of the receptor into the cell culture medium.

Particular examples of soluble IL-4R polypeptides lack the transmembrane region but comprise the extracellular domain (the complete extracellular domain or a fragment thereof that is capable of binding IL-4). As one option, the polypeptide comprises all or part of the cytoplasmic domain, as well as the extracellular domain (or fragment of the extracellular domain), but lacks the transmembrane region.

Examples of soluble human IL-4R polypeptides include, but are not limited to, polypeptides comprising amino acid residues x to y of SEQ ID NO:2, wherein x represents 1 or -2 and y represents an integer from 197 to 207. Examples include polypeptides comprising residues 1 to 207 or -2 to 207 of SEQ ID NO:2.

A protein preparation administered as an IL-4 antagonist may comprise more than one form of IL-4R. For example, the preparation may comprise polypeptide molecules consisting of amino acids 1 to 207 of SEQ ID NO:2, as well as polypeptides consisting of amino acids -2 to 207 of SEQ ID NO:2.

IL-4R polypeptides arising from alternative mRNA constructs, e.g., which can be attributed to different mRNA splicing events following transcription, and which yield polypeptide translates capable of binding IL-4, are among the IL-4R polypeptides disclosed herein. Such alternatively spliced mRNAs may give rise to soluble polypeptides.

Further examples of IL-4 receptors that may be employed in the methods described herein are variants having amino acid sequences which are substantially similar to the native interleukin-4 receptor amino acid sequence of SEQ ID NO:2, or fragments thereof. Variant IL-4 receptor polypeptides that are capable of functioning as IL-4 antagonists may be employed in the methods herein described.

Any of a number of conventional assay techniques may be employed to confirm that a given form of IL-4R (e.g., an IL-4R fragment or variant) functions as an IL-4 antagonist. Examples include binding assays or assays that test the ability of a given IL-4R polypeptide to inhibit transduction of an IL-4-induced biological signal. Examples of suitable in vitro assays are described below.

"Substantially similar" IL-4 receptors include those having amino acid or nucleic acid sequences that vary from a native sequence by one or more substitutions, deletions, or additions, but retain a desired biological activity of the IL-4R protein. Examples of nucleic acid molecules encoding IL-4 receptors include, but are not limited to: (a) DNA derived from the coding region of a native mammalian IL-4R gene; (b) DNA that is capable of hybridization to a DNA of (a) under moderately stringent conditions and which encodes an IL-4R having a biological activity of a native IL-4R; or (c) DNA that is degenerate as a result of the genetic code to a DNA defined in (a) or (b) and which encodes an IL-4R having a biological activity of a native IL-4R. Due to code degeneracy, there can be considerable variation in nucleotide sequences encoding the same amino acid sequence.

Variants may be naturally occurring, such as allelic variants or those arising from alternative splicing of mRNA. Alternatively, variants may be prepared by such well known techniques as in vitro mutagenesis.

A variant sequence identified by Idzerda et al., supra, comprises a GTC codon encoding the amino acid valine (Val) at position 50, instead of isoleucine (Ile). The variant sequence is otherwise identical to the sequence of SEQ ID NO:1 and 2. IL-4R fragments, such as soluble fragments, comprise Val at position 50 are provided herein.

In particular examples, an IL-4 receptor DNA or amino acid sequence is at least 80 percent identical to the sequence of a native IL-4R. Preferably, an IL-4R DNA or polypeptide comprises a sequence that is at least 90 percent identical to a native IL-4R DNA or amino acid sequence. One example is a human IL-4R comprising an amino acid sequence that is at least 80 percent identical to the sequence presented in SEQ ID NO:2. Another example is a soluble IL-4R comprising an amino acid sequence at least 80 percent identical to the sequence of the extracellular domain of human IL-4R. Further examples are polypeptides comprising amino acid sequences that are at least 90 percent identical to the sequence presented in SEQ ID NO:2, or a fragment thereof. In a particular example, the polypeptide comprises no more than 10 amino acid substitutions. IL-4R polypeptides that retain the ability to bind IL-4 may be identified in conventional binding assays.

Percent similarity or percent identity may be determined, for example, by comparing DNA or amino acid sequence information using the GAP computer program, version 6.0, available from the University of Wisconsin Genetics Computer Group (UWGCG). The GAP program utilizes the alignment method of Needleman and Wunsch (J. Mol. Biol.
IL-4R polypeptides that vary from native proteins but possess a desired property may be constructed by, for example, substituting or deleting residues not needed for the particular biological activity. Substitutions may be conservative substitutions, such that a desired biological property of the protein is retained. Amino acids may be replaced with residues having similar physicochemical characteristics.

Cysteine residues can be deleted or replaced with other amino acids to prevent formation of incorrect intramolecular disulfide bridges upon renaturation. Other alterations of a native sequence involve modification of adjacent dibasic amino acid residues, to enhance expression in yeast host cells in which KEX2 protease activity is present.

Also described is IL-4R with or without associated native-pattern glycosylation. The glycosylation pattern may vary according to the type of host cells in which the protein is produced. Another option is inactivation of N-glycosylation sites by site-specific mutagenesis. N-glycosylation sites in eukaryotic proteins are characterized by the amino acid triplet Asn-X-Ser or Thr, where X is any amino acid except Pro, and Z is Ser or Thr. In this sequence, asparagine provides a side chain amino group for covalent attachment of carbohydrate. Such a site can be eliminated by substituting another amino acid for Asn or for residue Z, deleting Asn or Z, or inserting a non-Z amino acid between A1 and Z, or an amino acid other than Asn between Asn and A1.

Oligonucleotide-directed site-specific mutagenesis procedures can be employed to provide an altered gene having particular codons altered according to the substitution, deletion, or insertion required. Examples of techniques for making such alterations are described in Walder et al. (Gene 42:133, 1986); Bauer et al. (Gene 37:73, 1985); Craik (BioTechniques, January 1985, 12-19); Smith et al. (Genetic Engineering: Principles and Methods, Plenum Press, 1981 and U.S. Patent Nos. 4,518,584 and 4,737,462).

IL-4 receptors that may be employed also include derivatives, e.g., various structural forms of the primary protein which retain a desired biological activity. Due to the presence of ionizable amino and carboxyl groups, for example, an IL-4R protein may be in the form of acidic or basic salts, or in neutral form. Individual amino acid residues may also be modified by oxidation or reduction. The primary amino acid sequence may be modified by forming covalent or aggregative conjugates with other chemical moieties, such as glycosyl groups, lipids, phosphate, acetyl groups and the like, or by creating amino acid sequence mutants. PEGylated derivatives (modified with polyethylene glycol) are contemplated. Covalent derivatives may be prepared by linking particular functional groups to IL-4R amino acid side chains or at the N- or C-termini. IL-4R derivatives may also be obtained by cross-linking agents, such as M-maleimidobenzoyl succinimide ester and N-hydroxysuccinimide, at cysteine and lysine residues. IL-4R proteins may also be covalently bound through reactive side groups to various insoluble substrates, such as cyanogen bromide-activated, bisoxirane-activated, carboxydiimidazole-activated or tosyl-activated agarose structures, or by adsorbing to polyeofin surfaces (with or without glutaraldehyde cross-linking).

Other derivatives of IL-4R described herein include covalent or aggregative conjugates of IL-4R or its fragments with other proteins or polypeptides, such as by expression of recombinant fusion proteins comprising heterologous polypeptides fused to the N-terminus or C-terminus of an IL-4R polypeptide. For example, the conjugated peptide may be a heterologous signal (or leader) polypeptide, e.g., the yeast α-factor leader, or a peptide such as an epitope tag. IL-4R-containing fusion proteins may comprise peptides added to facilitate purification or identification of IL-4R (e.g., poly-His). Specific derivatives of poly-His fusion constructs that is biologically active are soluble human IL-4R (e.g., comprising residues 2 to 207 or 1-207 of SEQ ID NO:2) His His and soluble human IL-4R His His His His His. An amino acid sequence of IL-4 receptor can also be linked to the Flag® peptide Asp-Tyr-Lys-Asp-Asp-Asp-Lys (DYKDDDDK) (SEQ ID NO:3) as described in Hopp et al., Bio/Technology 6:1204, 1988, and U.S. Patent 5,011,912. The Flag® peptide is highly antigenic and provides an epitope reversibly bound by a specific monoclonal antibody, enabling rapid assay and facile purification of expressed recombinant protein. Reagents useful for preparing fusion proteins in which the Flag® peptide is fused to a given polypeptide are commercially available (Sigma, St. Louis, MO).

Oligomers that contain IL-4R polypeptides may be employed as IL-4 antagonists. Oligomers may be in the form of covalently-linked or non-covalently-linked dimers, trimers, or higher oligomers. Oligomers comprising two or more IL-4R polypeptides are contemplated for use, with one example being a homodimer. Other oligomers include heterodimers, heterotrimers, and the like, which comprise an IL-4R polypeptide as well as at least one polypeptide that is not derived from the IL-4R of SEQ ID NO:2.

One example is directed to oligomers comprising multiple IL-4R polypeptides joined via covalent or non-covalent interactions between peptide moieties fused to the IL-4R polypeptides. Such peptides may be peptide linkers (spacers), or peptides that have the property of promoting oligomerization. Leucine zippers and certain polypeptides derived from
antibodies are among the peptides that can promote oligomerization of IL-4R polypeptides attached thereto, as described in more detail below.

[0104] In particular examples, the oligomers comprise from two to four IL-4R polypeptides. The IL-4R moieties of the oligomer may be in any of the forms described above, e.g., variants or fragments. Preferably, the oligomers comprise soluble IL-4R polypeptides.

[0105] As one alternative, an oligomer is prepared using polypeptides derived from immunoglobulins. Preparation of fusion proteins comprising certain heterologous polypeptides fused to various portions of antibody-derived polypeptides (including the Fc domain) has been described, e.g., by Ashkenazi et al. (PNAS USA 88:10535, 1991); Byrn et al. (Nature 344:677, 1990); and Hollenbaugh and Aruffo ("Construction of Immunoglobulin Fusion Proteins", in Current Protocols in Immunology, Suppl. 4, pages 10.19.1 - 10.19.11, 1992).

[0106] One example is directed to a dimer comprising two fusion proteins created by fusing IL-4R to the Fc region of an antibody. A gene fusion encoding the IL-4R/Fc fusion protein is inserted into an appropriate expression vector. IL-4R/Fc fusion proteins are expressed in host cells transformed with the recombinant expression vector, and allowed to assemble much like antibody molecules, whereupon interchain disulfide bonds form between the Fc moieties to yield divalent IL-4R.

[0107] The term "Fc polypeptide" as used herein includes native and mutein forms of polypeptides derived from the Fc region of an antibody. Truncated forms of such polypeptides containing the hinge region that promotes dimerization are also included. Fusion proteins comprising Fc moieties (and oligomers formed therefrom) offer the advantage of facile purification by affinity chromatography over Protein A or Protein G columns.

[0108] One suitable Fc polypeptide, described in PCT application WO 93/10151, is a single chain polypeptide extending from the N-terminal hinge region to the native C-terminus of the Fc region of a human IgG1 antibody. Another useful Fc polypeptide is the Fc mutein described in U.S. Patent 5,457,035 and in Baum et al., (EMBO J. 13:3992-4001, 1994). The amino acid sequence of this mutein is identical to that of the native Fc sequence presented in WO 93/10151, except that amino acid 19 has been changed from Leu to Ala, amino acid 20 has been changed from Leu to Glu, and amino acid 22 has been changed from Gly to Ala. The mutein exhibits reduced affinity for Fc receptors.

[0109] In other examples, IL-4R may be substituted for the variable portion of an antibody heavy or light chain. If fusion proteins are made with both heavy and light chains of an antibody, it is possible to form an oligomer with as many as four IL-4R extracellular regions.

[0110] Soluble recombinant fusion proteins comprising an IL-4R and various portions of the constant region of an immunoglobulin are described in EP 464,533, along with procedures for preparing such fusion proteins and dimers thereof. Among the fusion proteins described in EP 464,533 are those comprising the extracellular portion of human IL-4R and an Fc polypeptide.

[0111] Alternatively, the oligomer is a fusion protein comprising multiple IL-4R polypeptides, with or without peptide linkers (spacer peptides). Among the suitable peptide linkers are those described in U.S. Patents 4,751,180 and 4,935,233.

[0112] Another method for preparing oligomeric IL-4R involves use of a leucine zipper. Leucine zipper domains are peptides that promote oligomerization of the proteins in which they are found. Leucine zippers were originally identified in several DNA-binding proteins (Landschulz et al., Science 240:1759, 1988), and have since been found in a variety of different proteins. Among the known leucine zippers are naturally occurring peptides and derivatives thereof that dimerize or trimerize. Examples of leucine zipper domains suitable for producing soluble oligomeric proteins are described in PCT application WO 94/10308, and the leucine zipper derived from lung surfactant protein D (SPD) described in Hoppe et al. (FEBS Letters 344:191, 1994). The use of a modified leucine zipper that allows for stable trimerization of different proteins. Among the known leucine zippers are naturally occurring peptides and derivatives thereof that dimerize or trimerize. Examples of leucine zipper domains suitable for producing soluble oligomeric proteins are described in PCT application WO 94/10308, and the leucine zipper derived from lung surfactant protein D (SPD) described in Hoppe et al. (FEBS Letters 344:191, 1994). The use of a modified leucine zipper that allows for stable trimerization of a heterologous protein fused thereto is described in Fanslow et al. (Semin. Immunol. 6:267-278, 1994). In one approach, recombinant fusion proteins comprising a soluble IL-4R polypeptide fused to a leucine zipper peptide are expressed in suitable host cells, and the soluble oligomeric IL-4R that forms is recovered from the culture supernatant.

[0113] One example of a heterodimer comprises an IL-4R polypeptide derived from the human IL-4R of SEQ ID NO:2, and an IL-2Rγ polypeptide. IL-2Rγ (also known as IL-2Rγ) is described in U.S. Patent 5,510,259 and in Takeshita et al. (Science 257:379, 17 July 1992). The polypeptides may be in one of the various forms described herein, e.g., soluble fragments, variants, and the like, derived from the indicated proteins. One embodiment of such a heterodimer comprises a soluble IL-4R/Fc fusion protein and a soluble IL-2Rγ/Fc fusion protein. Such heterodimers are described in WO 96/11213, along with IL-4R homodimers.

[0114] Other examples of heterodimers comprise an IL-4R subunit (preferably a soluble fragment of the protein of SEQ ID NO:2) and at least one IL-13 receptor subunit. IL-13 receptor (IL-13R) complexes and IL-13R polypeptides (such as polypeptides designated IL-13Rα1 and IL-13Rα2) are described in Zurawski et al., J. Biol. Chem. 270 (23), 13869, 1995; de Vries, J. Allergy Clin. Immunol. 102(2):165, August 1998; Callard et al. Immunology Today, 17(3):108, March 1996, and U.S. Patent 5,710,023. In one example, a heterodimer comprises a soluble human IL-4R and a soluble IL-13R (preferably a soluble form of the polypeptide described in U.S. Patent 5,710,023 or IL-13Rα1). The components of heterodimers may be any suitable form of the polypeptides that retains the desired activity, such as fragments, variants,
or fusion proteins (e.g., fusions of soluble receptor polypeptides with Fc polypeptides, leucine zipper peptides, peptide linkers, or epitope tags).

[0115] IL-4 receptor polypeptides and fusion proteins described herein may be prepared by any of a number of conventional techniques. IL-4R polypeptides may be purified from cells that naturally express the receptor (such as the cells discussed in Park et al., Proc. Natl. Acad. Sci. USA 84:1669-673, 1987), or may be produced in recombinant expression systems, using well known techniques. Expression systems for use in producing IL-4R include those described in U.S. Patent 5,599,905.

[0116] A variety of expression systems are known for use in producing recombinant proteins. In general, host cells are transformed with a recombinant expression vector that comprises DNA encoding a desired IL-4R polypeptide. Among the host cells that may be employed are prokaryotes, yeast or higher eukaryotic cells. Prokaryotic cells include gram negative or gram positive organisms, for example E. coli or bacilli. Higher eukaryotic cells include insect cells and established cell lines of mammalian origin. Examples of suitable mammalian host cell lines include the COS-7 line of monkey kidney cells (ATCC CRL 1651) (Gluzman et al., Cell 23:175, 1981), L cells, 293 cells, C127 cells, 3T3 cells (ATCC CCL 163), Chinese hamster ovary (CHO) cells, HeLa cells, BHK (ATCC CRL 10) cell lines, and the CVI/EBNA cell line derived from the African green monkey kidney cell line CVI (ATCC CCL 70) as described by McMahan et al. (EMBO J. 10: 2821, 1991). Appropriate cloning and expression vectors for use with bacterial, fungal, yeast, and mammalian cellular hosts are described by Pauwells et al. (Cloning Vectors: A Laboratory Manual, Elsevier, New York, 1985).

[0117] The transformed cells are cultured under conditions that promote expression of the IL-4R, and the polypeptide is recovered by conventional protein purification procedures. One such purification procedure includes the use of affinity chromatography, e.g., over a matrix having IL-4 bound thereto. Expressed IL-4R will be deposited in the cell membrane or secreted into the culture supernatant, depending on the IL-4R DNA selected. Polypeptides contemplated for use herein include substantially homogeneous recombinant mammalian IL-4R polypeptides substantially free of contaminating endogenous materials.

Antibodies

[0118] Antibodies that function as IL-4 antagonists may be employed for use in the methods of the present invention. The antibodies are monoclonal antibodies. Most preferred are human monoclonal antibodies prepared using transgenic mice, as described below. Human antibodies also may be prepared using phage display techniques.

[0119] Examples of suitable antibodies are those that interfere with the binding of IL-4 to an IL-4 receptor. Such antibodies, referred to herein as blocking antibodies, may be raised against IL-4R, and screened in conventional assays for the ability to interfere with binding of IL-4 to IL-4 receptors. Examples of suitable assays are assays that test the antibodies for the ability to inhibit binding of IL-4 to cells expressing IL-4R, or that test antibodies for the ability to reduce a biological or cellular response that results from the binding of IL-4 to cell surface IL-4 receptors.

[0120] It has been reported that IL-4Rα is a component of certain multi-subunit IL-13 receptor complexes (Zurawski et al., J. Biol. Chem. 270 (23), 13869, 1995; de Vries, J. Allergy Clin. Immunol. 102(2):165, August 1998; and Callard et al., Immunology Today, 17(3):108, March 1996). Thus, some antibodies raised against IL-4Rα may interfere with the binding of IL-13 to such receptor complexes.

[0121] In one embodiment, antibodies directed against IL-4R block binding of IL-4 and also IL-13 to cells. The antibodies inhibit IL-4-induced biological activity and also inhibit IL-13-induced activity, and thus may be employed in treating inflammatory conditions induced by either or both cytokines. Examples of such conditions include but are not limited to IgE-mediated conditions, asthma, allergic conditions, allergic rhinitis, and dermatitis including atopic dermatitis.

[0122] Antibodies that bind to IL-4R and inhibit IL-4 binding may be screened in various conventional assays to identify those antibodies that also interfere with the binding of IL-13 to such receptor complexes. Antibodies may be screened in binding assays or tested for the ability to inhibit an IL-4-induced and an IL-13-induced biological activity. An example of a suitable assay is illustrated in Example 5 below.


[0124] Antigen-binding fragments of such antibodies may be produced by conventional techniques. Examples of such fragments include, but are not limited to, Fab, F(ab')2, and Fv fragments. Antibody fragments and derivatives produced by genetic engineering techniques are also contemplated for use. Unless otherwise specified, the terms “antibody” and “monoclonal antibody” as used herein encompass both whole antibodies and antigen-binding fragments thereof.

[0125] Additional embodiments include chimeric antibodies, e.g., humanized versions of murine monoclonal antibodies. Such humanized antibodies may be prepared by known techniques, and offer the advantage of reduced immunogenicity when the antibodies are administered to humans. In one embodiment, a humanized monoclonal antibody comprises the variable region of a murine antibody (or just the antigen binding site thereof) and a constant region derived...
from a human antibody. Alternatively, a humanized antibody fragment may comprise the antigen binding site of a murine monoclonal antibody and a variable region fragment (lacking the antigen-binding site) derived from a human antibody. Procedures for the production of chimeric and further engineered monoclonal antibodies include those described in Riechmann et al. (Nature 332:323, 1988), Liu et al. (PNAS 84:3439, 1987), Larrick et al. (Bio/Technology 7:934, 1989), and Winter and Harris (TIPS 14:139, May, 1993).

A method for producing an antibody comprises immunizing a non-human animal, such as a transgenic mouse, with an IL-4R polypeptide, whereby antibodies directed against the IL-4R polypeptide are generated in said animal. Procedures have been developed for generating human antibodies in non-human animals. The antibodies may be partially human, or preferably completely human. For example, transgenic mice into which genetic material encoding one or more human immunoglobulin chains has been introduced may be employed. Such mice may be genetically altered in a variety of ways. The genetic manipulation may result in human immunoglobulin polypeptide chains replacing endogenous immunoglobulin chains in at least some (preferably virtually all) antibodies produced by the animal upon immunization.

Mice in which one or more endogenous immunoglobulin genes have been inactivated by various means have been prepared. Human immunoglobulin genes have been introduced into the mice to replace the inactivated mouse genes. Antibodies produced in the animals incorporate human immunoglobulin polypeptide chains encoded by the human genetic material introduced into the animal.

Examples of techniques for production and use of such transgenic animals are described in U.S. Patents 5,814,318, 5,569,825, and 5,545,806. Examples 2-4 below provide further description of the preparation of transgenic mice useful for generating human antibodies directed against an antigen of interest.

Antibodies produced by immunizing transgenic non-human animals with an IL-4R polypeptide are provided herein. Transgenic mice into which genetic material encoding human immunoglobulin polypeptide chain(s) has been introduced are among the suitable transgenic animals. Examples of such mice include, but are not limited to, those containing the genetic alterations described in the examples below. One example of a suitable immunogen is a soluble human IL-4R, such as a polypeptide comprising the extracellular domain of the protein of SEQ ID NO:2, or other immunogenic fragment of the protein of SEQ ID NO:2.

Monoclonal antibodies may be produced by conventional procedures, e.g., by immortalizing spleen cells harvested from the transgenic animal after completion of the immunization schedule. The spleen cells may be fused with myeloma cells to produce hybridomas, by conventional procedures.

A method for producing a hybridoma cell line comprises immunizing such a transgenic animal with an IL-4R immunogen; harvesting spleen cells from the immunized animal; fusing the harvested spleen cells to a myeloma cell line, thereby generating hybridoma cells; and identifying a hybridoma cell line that produces a monoclonal antibody that binds an IL-4R polypeptide. Such hybridoma cell lines, and anti-IL-4R monoclonal antibodies produced therefrom, are encompassed by the present invention. Monoclonal antibodies secreted by the hybridoma cell line are purified by conventional techniques. Hybridomas or MAbs may be further screened to identify MAbs with particular properties, such as the ability to block an IL-4-induced activity, and to block an IL-13-induced activity (see the assay in example 5).

Human antibodies that bind IL-4R are provided by the present invention. In one embodiment of the invention, antibodies produced in the animals incorporate human immunoglobulin polypeptide chains encoded by the human genetic material introduced into the animal.

Among the uses of antibodies directed against an IL-4R is use in assays to detect the presence of IL-4R polypeptides, either in vitro or in vivo. The antibodies also may be employed in purifying IL-4R proteins by immunoaffinity chromatography. Those antibodies that additionally can block binding of IL-4 to IL-4R may be used to inhibit a biological activity that results from such binding. Blocking antibodies find use in the present invention. Such antibodies which function as IL-4 antagonists may be employed in treating any IL-4-induced condition, including but not limited to asthma and allergies, e.g., contact dermatitis, and atopic dermatitis. In another embodiment, a human anti-IL-4R monoclonal antibody generated by procedures involving immunization of transgenic mice is employed in treating such conditions.

Antibodies may be employed in an in vitro procedure, or administered in vivo to inhibit an IL-4-induced biological activity. Disorders caused or exacerbated (directly or indirectly) by the interaction of IL-4 with cell surface IL-4 receptors thus may be treated. A therapeutic method involves in vivo administration of a blocking antibody to a mammal in an amount effective for reducing an IL-4-induced biological activity.

Antibodies of the invention include, but are not limited to, fully human monoclonal antibodies that inhibit a biological activity of IL-4 and also inhibit a biological activity of IL-13. One embodiment is directed to a human monoclonal antibody that at least partially blocks binding of IL-4 to a cell, and at least partially blocks binding of IL-13 to a cell.

Antibodies of the present invention include but are not limited to antibodies generated by immunizing a transgenic mouse with an IL-4 receptor immunogen, wherein the transgenic mouse is selected from the mouse strains described in example 3 below. The desired antibodies are human. In one embodiment, the immunogen is a human IL-4 receptor polypeptide. Hybridoma cell lines derived from the thus-immunized mice, wherein the hybridoma secretes a monoclonal antibody that binds IL-4R, also are provided herein. Examples of antibodies produced by immunizing such transgenic mice useful for generating human antibodies directed against an antigen of interest.

The antibodies also may be employed in purifying IL-4R proteins by immunoaffinity chromatography. Those antibodies that additionally can block binding of IL-4 to IL-4R may be used to inhibit a biological activity that results from such binding. Blocking antibodies find use in the present invention. Such antibodies which function as IL-4 antagonists may be employed in treating any IL-4-induced condition, including but not limited to asthma and allergies, e.g., contact dermatitis, and atopic dermatitis. In another embodiment, a human anti-IL-4R monoclonal antibody generated by procedures involving immunization of transgenic mice is employed in treating such conditions.

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mice are the human monoclonal antibodies designated 6-2 (described in example 6); 12B5 (described in example 8); and MABs 63, 1B7, 5A1, and 27A1 (all described in example 9). Monoclonal antibodies 6-2, 12B5, 63, 1B7, 5A1, and 27A1 are fully human antibodies, and are capable of inhibiting activity of both IL-4 and IL-13. MABs 12B5, 63, and 1B7 are preferred antagonists of human IL-4 and human IL-13.

[0137] Certain monoclonal antibodies of the invention compete with 6-2 for binding to a cell that expresses human IL-4R. In one embodiment, the antibody has a binding affinity for human IL-4R that is substantially equivalent to the binding affinity of 6-2 for human IL-4R. MAb 6-2 is an IgM antibody. MABs of other isotypes (including but not limited to IgG1 and IgG4), derived from 6-2, also are encompassed by the present invention. Hybridoma cell lines that produce any such monoclonal antibodies also are provided by the present disclosure. One example of a biological activity of 6-2 is the ability to function as both an IL-4 antagonist and an IL-13 antagonist. In one embodiment, a MAB of the invention possesses IL-4-blocking activity substantially equivalent to that of 6-2; and possesses IL-13-blocking activity substantially equivalent to that of 6-2. Such activity may be measured in any suitable conventional assay (e.g., as measured in the CD23 expression assay described in example 5).

[0138] The DNA sequence of the variable region of the light chain of MAB 6-2 is presented in SEQ ID NO:5, and the encoded amino acid sequence is presented in SEQ ID NO:6. The DNA sequence for the variable region of the heavy chain of MAB 6-2 is presented as SEQ ID NO:7, and the encoded amino acid sequence is presented in SEQ ID NO:8. Antibodies of the present invention include, but are not limited to, monoclonal antibodies that comprise, in their light chain, residues 1 to 107 of SEQ ID NO:6; and antibodies that additionally or alternatively comprise, in their heavy chain, residues 1 to 118 of SEQ ID NO:8.

[0139] Complementarity determining regions (CDRs) of a given antibody may be identified using the system described by Kabat et al. in Sequences of Proteins of Immunological Interest, 5th Ed., US Dept. of Health and Human Services, PHS, NIH, NIH Publication no. 91-3242, 1991). Particular embodiments of antibodies of the present invention comprise, within the variable region of their light chain, at least one of the complementarity determining regions (CDRs), or hypervariable regions, found in the light chain of 6-2. CDRs of 6-2 are discussed in example 6. Thus, among the antibodies provided herein are those comprising from one to all three of the following sequences in the light chain variable region: amino acid residues 24-35 of SEQ ID NO:6; residues 51-57 of SEQ ID NO:6; and residues 90-97 of SEQ ID NO:6. Particular antibodies provided herein comprise, within the variable region of their heavy chain, at least one of the CDRs found in the heavy chain of 6-2. Thus, among the antibodies provided herein are those comprising from one to all three of the following sequences in the heavy chain variable region: residues 31-35; residues 50-66; and residues 99-109 of SEQ ID NO:6.

[0140] Described herein are monoclonal antibodies selected from the group consisting of MAB 12B5; a Mab that is cross-reactive with 12B5; a MAB that binds to the same epitope as 12B5; a MAB that competes with 12B5 for binding to a cell that expresses human IL-4R; a MAB that possesses a biological activity of 12B5; and an antigen-binding fragment of any of the foregoing antibodies. The antibody may have a binding affinity for human IL-4R that is substantially equivalent to the binding affinity of 12B5 for human IL-4R. MAB 12B5 is an IgG1 antibody. MABs of other isotypes, derived from 12B5, also are encompassed by the present invention. The isotype of the MAB may be IgG1, IgG4, or IgM. Hybridoma cell lines that produce any such monoclonal antibodies also are described.

[0141] One example of a biological activity of 12B5 is the ability to function as both an IL-4 antagonist and an IL-13 antagonist. In one example, a MAB described herein possesses IL-4-blocking activity substantially equivalent to that of 12B5, and possesses IL-13-blocking activity substantially equivalent to that of 12B5. Such activity may be measured in any suitable conventional assay (e.g., as measured in the CD23 expression assay described in example 5).

[0142] IgG4 monoclonal antibodies derived from 12B5 are provided herein. Another example is directed to IgM monoclonal antibodies derived from 12B5. Procedures for switching (altering) the subclass or isotype of an antibody are known in the pertinent field. Such procedures may involve, for example, recombinant DNA technology, whereby DNA encoding antibody polypeptide chains that confer the desired subclass is substituted for DNA encoding the corresponding polypeptide chain of the parent antibody.

[0143] The DNA sequence of the variable region of the light chain of MAB 12B5 is presented in SEQ ID NO:9, and the encoded amino acid sequence is presented in SEQ ID NO:10. The DNA sequence for the variable region of the heavy chain of MAB 12B5 is presented as SEQ ID NO:11, and the encoded amino acid sequence is presented in SEQ ID NO:12. Antibodies described herein may include monoclonal antibodies that comprise, in their light chain, residues 1 to 109 of SEQ ID NO:10; and antibodies that additionally or alternatively comprise, in their heavy chain, residues 1 to 115 of SEQ ID NO:12.

[0144] Particular examples of antibodies described herein may comprise, within the variable region of their light chain, at least one of the complementarity determining regions (CDRs), or hypervariable regions, found in the light chain of 12B5. CDRs of 12B5 are discussed in example 8. Thus, among the antibodies described herein are those comprising from one to all three of the following sequences in the light chain variable region: amino acid residues 24-35 of SEQ ID NO:10; residues 51-57 of SEQ ID NO:10; and residues 90-99 of SEQ ID NO:10. Particular antibodies described herein comprise, within the variable region of their heavy chain, at least one of the CDRs found in the heavy chain of 12B5.
Thus, among the antibodies described provided herein are those comprising from one to three of the following sequences in the heavy chain variable region: residues 31-35; residues 50-65; and residues 98-104 of SEQ ID NO:12.

[0145] Particular monoclonal antibodies described herein may be selected from the group consisting of MAb 27A1; a Mab that is cross-reactive with 27A1; a MAb that binds to the same epitope as 27A1; a MAb that competes with 27A1 for binding to a cell that expresses human IL-4R; and a MAb that possesses a biological activity of 27A1; and an antigen-binding fragment of any of the foregoing antibodies. The antibody may be a binding affinity for human IL-4R that is substantially equivalent to the binding affinity of 27A1 for human IL-4R. 27A1 is an IgG1 antibody. MAbs of other isotypes, derived from 27A1, also are described herein. Hybridoma cell lines that produce any such monoclonal antibodies also are described herein.

[0146] One example of a biological activity of 27A1 is the ability to function as both an IL-4 antagonist and an IL-13 antagonist. In one example, a MAb described herein possesses IL-4-blocking activity substantially equivalent to that of 27A1; and possesses IL-13-blocking activity substantially equivalent to that of 27A1. Such activity may be measured in any suitable conventional assay (e.g., as measured in the CD23 expression assay described in example 5).

[0147] The DNA sequence of the variable region of the light chain of MAb 27A1 is presented in SEQ ID NO:13, and the encoded amino acid sequence is presented in SEQ ID NO:14. The DNA sequence for the variable region of the heavy chain of MAb 27A1 is presented as SEQ ID NO:15, and the encoded amino acid sequence is presented in SEQ ID NO:16. Antibodies described herein may include, monoclonal antibodies that comprise, in their light chain, residues 1 to 109 of SEQ ID NO:14; and antibodies that additionally or alternatively comprise, in their heavy chain, residues 1 to 116 of SEQ ID NO:16.

[0148] Particular examples of antibodies described herein may comprise, within the variable region of their light chain, at least one of the complementarity determining regions (CDRs), or hypervariable regions, found in the light chain of 27A1. CDRs of 27A1 are discussed in example 9. Thus, among the antibodies described herein are those comprising from one to all three of the following sequences in the light chain variable region: amino acid residues 24-35 of SEQ ID NO:13; and residues 99-105 of SEQ ID NO:16. Antibodies described herein may include, monoclonal antibodies that comprise, in their heavy chain, residues 1 to 109 of SEQ ID NO:14; and antibodies that additionally or alternatively comprise, in their heavy chain, residues 1 to 116 of SEQ ID NO:16.

[0149] Particular monoclonal antibodies described herein may be selected from the group consisting of MAb 5A1; a Mab that is cross-reactive with 5A1; a MAb that binds to the same epitope as 5A1; a MAb that competes with 5A1 for binding to a cell that expresses human IL-4R; and a MAb that possesses a biological activity of 5A1; and an antigen-binding fragment of any of the foregoing antibodies. The antibody may have a binding affinity for human IL-4R that is substantially equivalent to the binding affinity of 5A1 for human IL-4R. 5A1 is an IgG1 antibody. MAbs of other isotypes, derived from 5A1, also are encompassed by the present invention. Hybridoma cell lines that produce any such monoclonal antibodies also are provided by the present invention.

[0150] One example of a biological activity of 5A1 is the ability to function as both an IL-4 antagonist and an IL-13 antagonist. In one example, a MAb described herein possesses IL-4-blocking activity substantially equivalent to that of 5A1; and possesses IL-13-blocking activity substantially equivalent to that of 5A1. Such activity may be measured in any suitable conventional assay (e.g., as measured in the CD23 expression assay described in example 5).

[0151] The DNA sequence of the variable region of the light chain of MAb 5A1 is presented in SEQ ID NO:17, and the encoded amino acid sequence is presented in SEQ ID NO:18. The DNA sequence for the variable region of the heavy chain of MAb 5A1 is presented as SEQ ID NO:19, and the encoded amino acid sequence is presented in SEQ ID NO:20. Antibodies described herein include, but are not limited to, monoclonal antibodies that comprise, in their light chain, residues 1 to 107 of SEQ ID NO:18; and antibodies that additionally or alternatively comprise, in their heavy chain, residues 1 to 123 of SEQ ID NO:20.

[0152] Particular examples of antibodies described herein may comprise, within the variable region of their light chain, at least one of the complementarity determining regions (CDRs), or hypervariable regions, found in the light chain of 5A1. CDRs of 5A1 are discussed in example 9. Thus, among the antibodies described herein are those comprising from one to all three of the following sequences in the light chain variable region: amino acid residues 24-34 of SEQ ID NO:17; and residues 99-105 of SEQ ID NO:18. Antibodies described herein may include, monoclonal antibodies that comprise, in their heavy chain, residues 1 to 109 of SEQ ID NO:17; and antibodies that additionally or alternatively comprise, in their heavy chain, residues 1 to 123 of SEQ ID NO:20.

[0153] Particular monoclonal antibodies described herein may be selected from the group consisting of MAb 63; a Mab that is cross-reactive with MAb 63; a MAb that binds to the same epitope as MAb 63; a MAb that competes with MAb 63 for binding to a cell that expresses human IL-4R; and a MAb that possesses a biological activity of MAb 63; and an antigen-binding fragment of any of the foregoing antibodies. The antibody may have a binding affinity for human IL-4R that is substantially equivalent to the binding affinity of MAb 63 for human IL-4R. MAb 63 is an IgM antibody. MAbs of other isotypes, derived from MAb 63, also are encompassed by the present invention. Hybridoma cell lines that produce any such monoclonal
antibodies also are provided by the present invention.

[0154] One example of a biological activity of 63 is the ability to function as both an IL-4 antagonist and an IL-13 antagonist. In one example, a MAb described herein possesses IL-4-blocking activity substantially equivalent to that of 63; and possesses IL-13-blocking activity substantially equivalent to that of 63. Such activity may be measured in any suitable conventional assay (e.g., as measured in the CD23 expression assay described in example 5).

[0155] The DNA sequence of the variable region of the light chain of MA b 63 is presented in SEQ ID NO:21, and the encoded amino acid sequence is presented in SEQ ID NO:22. The DNA sequence for the variable region of the heavy chain of MA b 63 is presented as SEQ ID NO:23, and the encoded amino acid sequence is presented in SEQ ID NO:24. Antibodies described herein may include monocl onal antibodies that comprise, in their light chain, residues 1 to 107 of SEQ ID NO:22; and antibodies that additionally or alternatively comprise, in their heavy chain, residues 1 to 117 of SEQ ID NO:24.

[0156] Particular examples of antibodies described herein comprise, within the variable region of their light chain, at least one of the complementarity determining regions (CDRs), or hypervariable regions, found in the light chain of 63. CDRs of 63 are discussed in example 9. Thus, among the antibodies described herein are those comprising from one to all three of the following sequences in the light chain variable region: amino acid residues 24-34 of SEQ ID NO:22; residues 50-56 of SEQ ID NO:22; and residues 89-97 of SEQ ID NO:22. Particular antibodies described herein comprise, within the variable region of their heavy chain, at least one of the CDRs found in the heavy chain of 63. Thus, among the antibodies described herein are those comprising from one to all three of the following sequences in the heavy chain variable region: residues 31-35; residues 50-66; and residues 99-106 of SEQ ID NO:24.

[0157] Certain monoclonal antibodies of the invention compete with 1B7 for binding to a cell that expresses human IL-4R. In one embodiment, the antibody has a binding affinity for human IL-4R that is substantially equivalent to the binding affinity of 1B7 for human IL-4R. MAbs of other isotypes, derived from 1 B7, also are encompassed by the present invention. Hybridoma cell lines that produce any such monoclonal antibodies also are provided by the present disclosure.

[0158] One example of a biological activity of 1B7 is the ability to function as both an IL-4 antagonist and an IL-13 antagonist. In one embodiment, a MA b of the invention possesses IL-4-blocking activity substantially equivalent to that of 1B7; and possesses IL-13-blocking activity substantially equivalent to that of 1 B7. Such activity may be measured in any suitable conventional assay (e.g., as measured in the CD23 expression assay described in example 5).

[0159] MA b 1B7 was derived from MA b 63. The amino acid sequence of the heavy chain of MA b 1B7 is identical to that of MA b 63. The only differences between the two antibodies are in the light chain. The DNA sequence of the variable region of the light chain of MA b 1B7 is presented in SEQ ID NO:25, and the encoded amino acid sequence is presented in SEQ ID NO:26. The DNA sequence for the variable region of the heavy chain of MA b 1B7 is presented as SEQ ID NO:23, and the encoded amino acid sequence is presented in SEQ ID NO:24 (same as for MA b 63). Antibodies of the present invention include, but are not limited to, monoclonal antibodies that comprise, in their light chain, residues 1 to 107 of SEQ ID NO:26; and antibodies that additionally or alternatively comprise, in their heavy chain, residues 1 to 117 of SEQ ID NO:24.

[0160] Particular embodiments of antibodies of the present invention comprise, within the variable region of their light chain, at least one of the complementarity determining regions (CDRs), or hypervariable regions, found in the light chain of 1B7. CDRs of 1B7 are discussed in example 9. Thus, among the antibodies provided herein are those comprising from one to all three of the following sequences in the light chain variable region: amino acid residues 24-34 of SEQ ID NO:22; residues 50-56 of SEQ ID NO:22; and residues 89-97 of SEQ ID NO:22. Particular antibodies described herein comprise, within the variable region of their heavy chain, at least one of the CDRs found in the heavy chain of 1B7. Thus, among the antibodies provided herein are those comprising from one to all three of the following sequences in the heavy chain variable region: residues 31-35; residues 50-66; and residues 99-106 of SEQ ID NO:24.

[0161] Figure 4 presents an alignment of the amino acid sequences for the variable region of the light chain for antibodies 1B7, 63, 12B5, 6-2, 5A1, and 27A1. The first sequence in the alignment is for antibody 1B7. In the sequences for the other antibodies, amino acid residues that differ from the residue found at the corresponding position in the 1B7 sequence are shown, and residues that are identical to the residue found at that position in 1B7 are indicated by "-".

[0162] Figure 5 presents an alignment of the amino acid sequences for the variable region of the heavy chain for antibodies 1B7, 63, 12B5, 6-2, 5A1, and 27A1. The first sequence in the alignment is for antibody 1B7. In the sequences for the other antibodies, amino acid residues that differ from the residue found at the corresponding position in the 1B7 sequence are shown, and residues that are identical to the residue found at that position in 1B7 are indicated by "-".

[0163] As can be seen in Figures 4 and 5, a number of residues are highly conserved, in that the residue appears at a corresponding position in all six of the antibodies. Particular antibodies of the invention may comprise, within the variable region of their light chain, amino acid residues that are found in the corresponding position in at least three of the sequences of Figure 4. In certain examples, the antibodies comprise residues found in a corresponding position in at least four, or at least five, or at least six, of the sequences of Figure 4. Particular antibodies of the invention may comprise, within the variable region of their heavy chain, amino acid residues that are found in the corresponding position in at least three of the sequences of Figure 5. In certain examples, the antibodies comprise residues found in a corre-
spending position in at least four, or at least five, or at least six, of the sequences of Figure 5.

Derivatives of monoclonal antibodies directed against IL-4R may be prepared, and screened for desired properties, by any of a number of known techniques. Certain of the techniques involve isolating DNA encoding a polypeptide chain (or portion thereof) of a MAb of interest, and manipulating the DNA through recombinant DNA technology. The DNA may be fused to another DNA of interest, or altered (e.g., by mutagenesis or other conventional techniques) to add, delete, or substitute one or more amino acid residues, for example.

DNA encoding antibody polypeptides (e.g., heavy or light chain, variable region only or full length) may be isolated from B-cells of mice that have been immunized with IL-4R. The DNA may be isolated by conventional procedures such as polymerase chain reaction (PCR). Phage display is another example of a known technique whereby derivatives of antibodies may be prepared. In one approach, polypeptides that are components of an antibody of interest are expressed in any suitable recombinant expression system, and the expressed polypeptides are allowed to assemble to form antibody molecules.

Single chain antibodies may be formed by linking heavy and light chain variable region (Fv region) fragments via an amino acid bridge (short peptide linker), resulting in a single polypeptide chain. Such single-chain Fvs (scFvs) have been prepared by fusing DNA encoding a peptide linker between DNAs encoding the two variable region polypeptides (VH and VL). The resulting antibody fragments can form dimers or trimers, depending on the length of a flexible linker between the two variable domains (Kortt et al., Protein Engineering 10:423, 1997). Techniques developed for the production of single chain antibodies include those described in U.S. Patent No. 4,946,778; Bird (Science 242:423, 1988); Huston et al. (Proc. Natl. Acad. Sci. USA 85:5879, 1988); and Ward et al. (Nature 334:544, 1989). Single chain antibodies derived from antibodies provided herein (including but not limited to scFvs derived from MAbs 6-2, 12B5, 63, 1B7, 5A1, and 27A1) may be encompassed by the present invention.

Techniques are known for deriving an antibody of a different subclass or isotype from an antibody of interest, i.e., subclass switching. Thus, IgG1 or IgG4 monoclonal antibodies may be derived from an IgM monoclonal antibody, for example, and vice versa. Such techniques allow the preparation of new antibodies that possess the antigen-binding properties of a given antibody (the parent antibody), but also exhibit biological properties associated with an antibody isotype or subclass different from that of the parent antibody. Recombinant DNA techniques may be employed. Cloned DNA encoding particular antibody polypeptides may be employed in such procedures, e.g., DNA encoding the constant region of an antibody of the desired isotype.

In particular examples, antibodies raised against IL-4R have a binding affinity (Ka) for IL-4R of at least 1 x 10^9. In other examples, the antibodies exhibit a Ka of at least 1 x 10^9 or at least 1 x 10^10.

PEGylated derivatives of antibodies (modified with polyethylene glycol) also are contemplated, and may be prepared by conventional techniques. Also described herein are conjugates comprising a detectable (e.g., diagnostic) or therapeutic agent, attached to an antibody directed against IL-4R. Examples of such agents are well known, and include but are not limited to diagnostic radionuclides, therapeutic radionuclides, and cytotoxic drugs. The conjugates find use in in vitro or in vivo procedures.

Particular examples of the invention are directed to novel nucleic acid molecules and polypeptides. DNA and amino acid sequence information has been determined for polypeptides that are components of certain antibodies of the present invention, as discussed in examples 6, 8, and 9 below. Among the nucleic acids is isolated DNA comprising a nucleotide sequence selected from the group consisting of the nucleotide sequence presented in SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, and SEQ ID NO:25. Among the polypeptides is a purified polypeptide comprising an amino acid sequence selected from the group consisting of the amino acid sequence presented in SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, and SEQ ID NO:26. For in vivo use, the polypeptides advantageously are purified. A polypeptide may be purified individually, or in the form of a purified antibody of which the polypeptide is a component.

Further examples of IL-4 antagonists are antibodies that bind IL-4 and inhibit the binding of IL-4 to cell surface receptors. Such antibodies may be prepared, and screened to identify those that are blocking antibodies, by conventional procedures. Antigen-binding fragments of such antibodies find use as antagonists, as do humanized or genetically engineered derivatives thereof.

Examples of procedures for preparing antibodies directed against human IL-4 (including monoclonal antibodies), assays by which blocking antibodies are identified, and techniques for generating humanized or genetically engineered derivatives of anti-IL-4 antibodies, are described in U.S. Patents 5,041,381, 5,863,537, 5,928,904, and 5,676,940. Further examples of antibodies that may be employed as IL-4 antagonists are described in WO 91/09059.

Other antagonists

Antagonists need not completely abolish IL-4-induced biological activity to be useful. Rather, a given antagonist may reduce a biological activity of IL-4.
Therapeutic Methods and Administration of Antagonists

Derivatives, mutants/muteins, and other variants of IL-4 that function as IL-4 antagonists may be employed. Peptides (which may or may not be muteins) derived from IL-4 that bind to an IL-4R without inducing transduction of a biological signal find use herein. Such peptides function as inert blockers, interfering with the binding of biologically active endogenous IL-4 to cell surface receptors. IL-4-induced signal transduction thereby is inhibited. Muteins or other antagonists that induce a biological response at a reduced level or to a lesser degree, compared to the response induced by native IL-4, also find use as IL-4 antagonists.

A mutein of human IL-4 is capable of inhibiting IL-4-induced and IL-13-induced biological activity. Such muteins may be employed in those methods described below that involve use of an antagonist of both IL-4 and IL-13 to treat particular disorders. See, for example, the human IL-4 mutein (a variant polypeptide with two point mutations) described in Fitch et al. (J. Allergy Clin. Immunol., Vol. 107, no. 2, pg. S61, abstract no. 209, 2001).

Further examples of IL-4 antagonists, including IL-4 muteins, and procedures for preparation thereof are described in Muller et al., J. Mol. Biol., 237:423-436, 1994; U.S. Patent 6,028,176, and U.S. Patent 5,723,118.

Other options are antisense molecules (oligonucleotides) that inhibit expression of IL-4. Alternatively, the antisense molecule may suppress expression of other molecules involved in IL-4-induced signal transduction.

Any suitable assay, including in vitro assays, can be utilized to determine whether a given compound inhibits an IL-4-induced biological activity. An antagonist may be assayed for the ability to inhibit 3H-thymidine incorporation in cells that normally proliferate in response to IL-4.

An alternative involves use of conventional binding assay techniques to test an antagonist for the ability to inhibit binding of IL-4 to cells expressing native or recombinant IL-4 receptors. For use in such assays, recombinant human IL-4 can be expressed and purified as described in U.S. Patent 5,017,691, or in Park et al., J. Exp. Med. 166:476 (1987). The purified protein may be labeled with a detectable agent (e.g., radiolabeled) by any of a number of conventional techniques. A commercially available enzyme-linked radioimmunodetection reagent (BioRad) may be employed in radiolabeling IL-4 with 125I for example.

The ability of an IL-4 antagonist to inhibit IL-4-induced damage to epithelium, such as lung epithelium or intestinal epithelium (which may result in loss of barrier function), may be confirmed in any of a number of suitable assays. Among the suitable assay techniques are those described in example 7 below.

Therapeutic Methods and Administration of Antagonists

Methods provided herein in which the antibody of the invention is used comprise administering the IL-4 antagonist antibody to a patient, thereby reducing an IL-4-induced biological response that plays a role in a particular condition. Particular methods involve contacting endogenous IL-4 with the antibody, e.g., in an ex vivo procedure.

Treatment encompasses alleviation of at least one symptom of a disorder, or reduction of disease severity, and the like. An antagonist need not effect a complete "cure", or eradicate every symptom or manifestation of a disease, to constitute a viable therapeutic agent. As is recognized in the pertinent field, drugs employed as therapeutic agents may reduce the severity of a given disease state, but need not abolish every manifestation of the disease to be regarded as useful therapeutic agents. A method comprising administering to a patient an IL-4 antagonist in an amount and for a time sufficient to induce a sustained improvement over baseline of an indicator that reflects the severity of the particular disorder, is described.

Antibodies that inhibit the binding of both IL-4 and IL-13 to cells are discussed herein. A method for suppressing IL-4-induced and IL-13-induced activities in humans comprises administering an effective amount of such an antibody. Conditions induced by IL-4 or by IL-13, or by both cytokines, thus may be treated.

As is understood in the pertinent field, antagonists are administered to a patient in a manner appropriate to the indication. Antagonists may be administered by any suitable technique, including but not limited to parenterally, topically, or by inhalation. If injected, the antagonist can be administered, for example, via intra-articular, intravenous, intramuscular, intradermal, intraperitoneal or subcutaneous routes, by bolus injection, or continuous infusion. Localized administration, e.g. at a site of disease or injury is contemplated, as are transdermal delivery and sustained release from implants. Delivery by inhalation includes, for example, nasal or oral inhalation, use of a nebulizer, inhalation of the antagonist in aerosol form, and the like. Other alternatives include eyedrops; oral preparations including pills, syrups, lozenges or chewing gum; and topical preparations such as lotions, gels, sprays, and ointments.

Use of IL-4 antagonists in ex vivo procedures is contemplated. For example, a patient's blood (bodily fluid containing IL-4) may be contacted with an antagonist that binds IL-4 ex vivo, thereby reducing the amount of IL-4 in the fluid when returned to the patient. The antagonist may be bound to a suitable insoluble matrix or solid support material.

Advantageously, antagonists are administered in the form of a composition comprising at least one IL-4 antagonist antibody in accordance with the invention and one or more additional components such as a physiologically acceptable carrier, excipient or diluent. The present invention provides such compositions comprising an effective amount of an IL-4 antagonist antibody, for use in the methods provided herein.

The compositions contain antagonist(s) in any of the forms described herein. The antagonist may be a whole
Methods described herein involve subcutaneous injection of from 0.5 mg to 10 mg, preferably from 3 to 5 mg, times per week. Involves the subcutaneous injection of 0.4 mg/kg, up to a maximum dose of 25 mg of IL-4R, administered two or three times per week over a period of at least three weeks, though treatment for longer periods may be necessary to induce the desired degree of improvement. For pediatric patients (age 4-17), one suitable regimen involves injecting a dose of about 20-30 mg of a soluble IL-4R once a week, at a dose of 1.5 to 3 mg, to treat pulmonary sarcoidosis, minimal change nephrosis, autoimmune uveitis, sickle cell crisis, Churg-Strauss syndrome, Sjogren’s syndrome, autoimmune lymphoproliferative syndrome, pre-eclampsia, autoimmune hemolytic anemia, Barrett’s esophagus, Grave’s Disease, Kawasaki Disease, and cavitary tuberculosis. Weekly administration of IL-4R is continued until symptoms subside. Treatment may resume as needed, or, alternatively, maintenance doses may be administered.

An antagonist is administered to the patient in an amount and for a time sufficient to induce an improvement, preferably a sustained improvement, in at least one indicator that reflects the severity of the disorder that is being treated. Various indicators that reflect the extent of the patient’s illness may be assessed for determining whether the amount and time of the treatment is sufficient. Such indicators include, for example, clinically recognized indicators of disease severity, symptoms, or manifestations of the disorder in question. In most instances, an improvement is considered to be sustained if the patient exhibits the improvement on at least two occasions separated by two to four weeks. The degree of improvement generally is determined by the patient’s physician, who may make this determination based on signs or symptoms, and who may also employ questionnaires that are administered to the patient, such as quality-of-life questionnaires developed for a given disease.

As one example, in treating benign prostate hyperplasia, an IL-4 inhibitor is administered to the patient in an amount and for a time effective in scar regression or complete healing. Maintenance doses may be given or treatment resumed as needed.

Elevated levels of IL-4 are associated with a number of disorders, as discussed above. Patients with a given disorder may be screened, to identify those individuals who have elevated IL-4 levels, or to identify those with an elevated
TH2-type immune response, thereby identifying the patients who may benefit most from treatment with an IL-4 antagonist. Thus, treatment methods provided herein optionally comprise a first step of measuring a patient’s IL-4 level. An IL-4 antagonist may be administered to a patient in whom IL-4 levels are elevated above normal. Alternatively or additionally, a patient may be pre-screened to determine whether the patient has an elevated TH2-type immune response, prior to administration of antagonist(s) against one or more TH2-type cytokines.

A patient’s levels of IL-4 (and, optionally, of other TH2-type cytokines) may be monitored during and/or after treatment with an IL-4 antagonist, to detect reduction in the levels of the cytokines. For some disorders, the incidence of elevated IL-4 levels, and the balance between TH1-type and TH2-type immune responses, may vary according to such factors as the stage of the disease or the particular form of the disease. Known techniques may be employed for measuring IL-4 levels, e.g., in a patient’s serum, and for assessing TH2-type immune responses. Cytokine levels in blood samples may be measured by ELISA, for example.

Particular examples described herein involve the use of two or more different IL-4 antagonists. In further examples, IL-4 antagonist(s) are administered alone or in combination with other agents useful for treating the condition with which the patient is afflicted. Examples of such agents include both proteinaceous and non-proteinaceous drugs. When multiple therapeutics are co-administered, dosages may be adjusted accordingly, as is recognized in the pertinent art. “Co-administration” and combination therapy are not limited to simultaneous administration, but include treatment regimens in which an IL-4 antagonist is administered at least once during a course of treatment that involves administering at least one other therapeutic agent to the patient.

Examples of other agents that may be co-administered with IL-4 antagonists are other antibodies, cytokines, or cytokine receptors, which are chosen according to the particular condition to be treated. Alternatively, non-proteinaceous drugs that are useful in treating one of the particular conditions discussed above may be co-administered with an IL-4 antagonist.

For treating IgE-mediated conditions, an IL-4 antagonist may be co-administered with an IgE antagonist. One example is an anti-IgE antibody. Humanized anti-IgE monoclonal antibodies are described in Presta et al. (J. Immunol. 151(5):2623-2632, 1993) and Adelroth et al. (J. Allergy Clin. Immunol. 106(2):253-259, 2000), for example.

IL-4 antagonists may be co-administered with an IL-5 antagonist, which may be a molecule that interferes with the binding of IL-5 to an IL-5 receptor, such as an anti-IL-5 antibody (e.g., a human or humanized anti-IL-5 monoclonal antibody), or a receptor such as a soluble human IL-5 receptor polypeptide. IL-5 has been implicated as playing a role in mediating allergic responses. Thus, administration of antagonist(s) of IL-4 and IL-5 is contemplated for treatment of allergic reactions, including but not limited to allergic asthma.

IL-4 antagonists may be employed in conjunction with other agent(s) in treating the particular IL-4-induced conditions discussed above. For example, drugs currently employed in treating the conditions may be co-administered with one or more IL-4 antagonists.

For treating asthma, an IL-4 antagonist may be co-administered with other anti-asthma medications, such as inhaled corticosteroids, beta agonists, leukotriene antagonists, xanthines, fluticasone, salmeterol, albuterol, non-steroidal agents such as cromolyn, and the like. IL-4 antagonists may be co-administered with other anti-allergy medications to treat allergic reactions.

One example described is directed to co-administration of an IL-4 antagonist (such as a soluble human IL-4R) and fluticasone and salmeterol to treat a disorder such as asthma. Compositions comprising an IL-4 inhibitor (e.g., soluble human IL-4R), fluticasone, and salmeterol are provided herein. Advair Diskus (Glaxo Wellcome) comprises fluticasone propionate and salmeterol xinafoate. For treating asthma, Advair Diskus and the IL-4 antagonist preferably are delivered by inhalation.

Another example of combination therapy comprises co-administration of an IL-4 antagonist and an IL-9 antagonist to a patient who has asthma. Any suitable IL-9 antagonist may be employed, such as an IL-9 receptor (preferably a soluble form thereof), an antibody that interferes with binding of IL-9 to a cell surface receptor (wherein the antibody may be raised against IL-9 or an IL-9 receptor polypeptide), or another compound that inhibits IL-9-induced biological activity. IL-9 receptors include those described in WO 93/18047 and U.S. Patents 5,789,237 and 5,962,269.

In an additional example of combination therapy, a method for treating ulcerative colitis comprises co-administration of at least one IL-4 antagonist and at least one IL-1 antagonist. Examples of IL-1 antagonists include type I IL-1 receptor, type II IL-1 receptor, IL-1 receptor antagonist (IL-1Ra), antagonistic (blocking) antibodies directed against IL-1, and antagonistic antibodies directed against an IL-1 receptor. Various forms of the receptors may be employed, such as fragments, variants and fusions analogous to those described above for IL-4 receptor. A preferred IL-1 antagonist is a soluble form of type II IL-1 receptor, which is described in U.S. Patent 5,350,683.

One method comprises co-administering IL-4 antagonist(s) and IL-13 antagonist(s) to a patient who has minimal change nephropathy. Alternative examples involve administering IL-4 antagonist(s) alone, or IL-13 antagonist(s) alone, to a minimal change nephropathy patient. The IL-4 antagonists(s) and/or IL-13 antagonist(s) may be administered to reduce severity of the disease.

Another method described herein is a method for treating various allergic inflammatory conditions, comprising
co-administering IL-4 antagonist(s) and IL-13 antagonist(s). Conditions such as asthma, allergies, and chronic lung
diseases such as cystic fibrosis and chronic obstructive pulmonary disease are treated by such a method.

Any suitable IL-13 antagonist may be employed, including but not limited to IL-13 receptors (preferably soluble
forms thereof), IL-13 receptor antagonists, antibodies directed against IL-13 or an IL-13 receptor, other proteins that
interfere with the binding of IL-13 to an IL-13 receptor, and compounds that inhibit IL-13-mediated signal transduction.
IL-13 receptors and heterodimers comprising IL-13R polypeptides as components thereof are described above. Anti-
body that are raised against IL-4R may be screened for the ability to also function as IL-13 antagonists, as discussed
above.

A method for treating or preventing a condition characterized by reduced epithelial barrier function comprises
co-administering IL-4 antagonist(s) and one or more IL-13 antagonists. Such conditions are discussed above. In one
example, the condition is asthma. Particular examples are directed to co-administering one or more IL-4 antagonists
and one or more IL-13 antagonists to a patient having a condition involving reduction of lung epithelial barrier function
or intestinal epithelial barrier function, wherein both IL-4 and IL-13 play a role in the reduced barrier function. The method
thus inhibits both IL-4-induced reduction of barrier function and IL-13-induced reduction of barrier function. The adverse
effect of IL-13 on lung and intestinal epithelial barrier function can be confirmed using assay techniques such as those
described in example 7 below. (See also Zund et al., J. Biol. Chem. 271(13):7460-7464, 1996.)

Another method provided herein comprises co-administering IL-4 antagonist(s) and interferon-γ (IFN-γ) to a patient
having a condition involving reduction of lung epithelial barrier function. Optionally, such a method further com-
prises co-administering one or more IL-13 antagonists to the patient (i.e., co-administering an IL-4 antagonist, IFN-γ,
and an IL-13 antagonist). Other methods comprise administering IFN-γ as a single agent, or co-administering IFN-γ
and an IL-13 antagonist, to a patient having a condition involving reduction of lung epithelial barrier function. In one example,
the patient has asthma. For treating asthma, the IL-4 antagonist, IFN-γ, and/or IL-13 antagonist preferably are admin-
istered by inhalation.

One method described herein for treating asthma comprises administering an IL-4 antagonist and interferon-
γ to a human who has asthma. Another method for treating asthma comprises co-administering an IL-4 antagonist, IFN-
γ, and an IL-13 antagonist to a human who has asthma. In one example, IFN-γ is co-administered to an asthmatic,
together with an antibody that functions as an antagonist of both IL-4 and IL-13. Such antibodies are described elsewhere
herein.

A single agent may function as an IL-4 antagonist and an IL-13 antagonist, as discussed above. As an example
of such an agent, some antibodies raised against IL-4Rα may interfere with the binding of both IL-4 and IL-13 receptor
complexes, due to the shared IL-4Rα component in such multi-subunit receptor complexes (discussed above). Thus, a
single agent may be employed in a method for inhibiting reduction of barrier function.

Antagonists may be co-administered with one or more leukotriene receptor antagonists to treat disorders such
as allergic inflammatory diseases, e.g., asthma and allergies. Examples of leukotriene receptor antagonists include but
are not limited to montelukast, pranlukast, and zafirlukast. Drugs that function as 5-lipoxygenase inhibitors may be co-
administered with an IL-4 antagonist to treat asthma.

Methods described herein comprise administering one or more of the following to Churg-Strauss Syndrome
patients: IL-4 antagonist(s), IL-5 antagonist(s), IL-13 antagonist(s) or IgE antagonist(s). One example of such a method
involves co-administering IL-4 antagonist(s) and IL-5 antagonist(s) to a Churg-Strauss Syndrome patient. In another
example, IL-4 antagonist(s) and IgE antagonist(s) are co-administered to the patient. In yet another example, IL-4
antagonist(s) and IL-13 antagonist(s) are co-administered to the patient.

The hormone relaxin may be co-administered with an IL-4 antagonist to treat scleroderma (systemic sclerosis),
idiopathic pulmonary fibrosis, or any other disorder characterized by pulmonary fibrosis, such as the conditions involving
fibrosis of the lung that are discussed above. Recombinant human relaxin is preferred for use in treating humans.

A method for treating benign prostate hyperplasia comprises co-administering IL-4 antagonist(s) and one or more
additional anti-inflammatory agents. Examples of agents that inhibit inflammation include tumor necrosis factor
(TNF) antagonists and IL-17 antagonists.

Any suitable IL-17 antagonist may be employed, including but not limited to an IL-17 receptor (preferably soluble
forms thereof), IL-17 receptor antagonists, antibodies directed against IL-17 or an IL-17 receptor, other proteins that
interfere with the binding of IL-17 to an IL-17 receptor, and compounds that inhibit IL-17-mediated signal transduction.
An IL-17 receptor, including soluble forms thereof and oligomers thereof, is described in WO 96/29408, hereby incor-
porated by reference. An alternative method described herein comprises administering an IL-17 antagonist to treat a
patient with benign prostate hyperplasia.

Likewise, any suitable TNF antagonist may be employed, including but not limited to a TNF receptor (preferably
soluble forms thereof), fusion proteins comprising a TNF receptor (or comprising the TNF-binding portion of a TNF
receptor), TNF receptor antagonists, antibodies directed against TNF or a TNF receptor, other proteins that interfere
with the binding of TNF to a TNF receptor, and compounds that inhibit TNF-mediated signal transduction. Further
examples of TNF inhibitors are the drugs thalidomide and pentoxyfylline. The TNF receptor protein known as p75 or
p80 TNF-R preferably is employed. A preferred TNF antagonist is a soluble human TNF receptor (sTNF-R) in dimeric form, such as dimers of sTNF-R/Fc fusion proteins. One such dimer is etanercept (Enbrel®, Immunex Corporation, Seattle, WA). p75/p80 TNF-R, including soluble fragments and other forms thereof, is described in WO 91/03553.

[0222] An IL-4 antagonist may be co-administered with a TNF antagonist to treat any condition in which undesirable IL-4-induced and TNF-induced immune responses play a role, such as inflammation. One method described herein comprises co-administering an IL-4 antagonist and a TNF antagonist to a patient with inflammatory bowel disease, Crohn’s disease, or ulcerative colitis. Other examples are directed to a method comprising co-administering an IL-4 antagonist and a TNF antagonist to a patient who has Kawasaki Disease, autoimmune hemolytic anemia, autoimmune uveoretinitis, autoimmune lymphoproliferative syndrome, Sjogren’s syndrome, chronic fatigue syndrome, or hepatotoxicity induced by a drug such as diclofenac.

[0223] Another method described herein comprises co-administering an IL-4 antagonist and a TNF antagonist to a pregnant woman who has developed pre-eclampsia. Administration of the IL-4 antagonist and TNF-antagonist preferably continues for the duration of the pregnancy.

[0224] Suitable dosages of etanercept (Enbrel®, Immunex Corporation, Seattle, WA) will vary according to the nature of the disease to be treated, disease severity, the size of the patient (e.g., adult or child), and other factors, as is recognized in the pertinent field. In one embodiment of the methods provided herein, Enbrel® is administered twice a week by subcutaneous injection at a dose of from 1 to 25 mg. One example of a pediatric dosage is 0.4 mg/kg. Particular methods provided herein comprise co-administration of an IL-4 antagonist and Enbrel® to a patient who has autoimmune lymphoproliferative syndrome or Sjogren’s syndrome, wherein Enbrel® is given by subcutaneous injection at a dose of from 1 to 25 mg.

[0225] For treating graft versus host disease, an IL-4 antagonist is co-administered with at least one of the following agents: a TNF antagonist, an IL-1 antagonist, steroids, or corticosteroids. The TNF inhibitor preferably is Enbrel®. A preferred IL-1 antagonist is a soluble form of II IL-1 receptor, which is described in U.S. Patent 5,350,683. In one example, the GVHD is associated with (e.g., develops subsequent to) bone marrow transplantation. An IL-4 antagonist may be employed in combination with at least one of the above-listed agents, in methods for suppressing an immune response directed against transplanted cells, tissue, and/or alloantigen.

[0226] A number of cytokine antagonists and other agents/drugs are disclosed herein as being useful for combination therapy (e.g., co-administration with an IL-4 antagonist) in treating particular diseases. It is to be understood that such antagonists, agents, or drugs also find use as single agents in treating those diseases. It also is to be understood that disclosure of methods involving administration of an antagonist to a particular cytokine, to treat a disease, encompasses administration of one type of antagonist, and also encompasses administration of two or more different antagonists for that cytokine, unless specified otherwise.

[0227] The following examples are offered by way of illustration, and not by way of limitation.

**EXAMPLE 1: Preparation of Monoclonal Antibodies**

[0228] IL-4 receptor polypeptides may be employed as immunogens in generating monoclonal antibodies by conventional techniques, e.g., techniques described in U.S. Patent 5,599,905. It is recognized that polypeptides in various forms may be employed as immunogens, e.g., full length proteins, fragments thereof, fusion proteins thereof such as Fc fusions, cells expressing the recombinant protein on the cell surface, etc.

[0229] To summarize an example of such a procedure, an IL-4R immunogen emulsified in complete Freund’s adjuvant is injected subcutaneously into Lewis rats, in amounts ranging from 10-100 μl. Three weeks later, the immunized animals are boosted with additional immunogen emulsified in incomplete Freund’s adjuvant and boosted every three weeks thereafter. Serum samples are periodically taken by retro-orbital bleeding or tail-tip excision for testing by dot-blot assay, ELISA (enzyme-linked immunosorbent assay), or inhibition of binding of 125I-IL-4 to extracts of IL-4R-expressing cells. Following detection of an appropriate antibody titer, positive animals were given a final intravenous injection of antigen in saline. Three to four days later, the animals are sacrificed, splenocytes harvested, and fused to the murine myeloma cell line AG88653. The resulting hybridoma cell lines are plated in multiple microtiter plates in a HAT selective medium (hypoxanthine, aminopterin, and thymidine) to inhibit proliferation of non-fused cells, myeloma hybrids, and spleen cell hybrids.

[0230] Hybridoma clones thus generated are screened for reactivity with IL-4R. Initial screening of hybridoma supernatants utilizes an antibody capture and binding of partially purified 125I-IL-4 receptor. Hybridomas that are positive in this screening method are tested by a modified antibody capture to detect hybridoma cells lines that are producing blocking antibody. Hybridomas that secrete a monoclonal antibody capable of inhibiting 125I-IL-4 binding to cells expressing IL-4R are thus detected. Such hybridomas then are injected into the peritoneal cavities of nude mice to produce ascites containing high concentrations (>1 mg/ml) of anti-IL-4R monoclonal antibody. The resulting monoclonal antibodies may be purified by ammonium sulfate precipitation followed by gel exclusion chromatography, and/or affinity chromatography based on binding of antibody to Protein G.
EXAMPLE 2: Generation of Cmu targeted mice

[0231] This example describes procedures for generating transgenic mice. Additional procedures for generating transgenic mice, and the use of such mice for preparing human antibodies, are described in Examples 3 and 4.

[0232] Construction of a CMD targeting vector. The plasmid pICEmu contains an EcoRI/Xhol fragment of the murine Ig heavy chain locus, spanning the mu gene, that was obtained from a Balb/C genomic lambda phage library (Marcu et al. Cell 22: 187, 1980). This genomic fragment was subcloned into the Xhol/EcoRI sites of the plasmid pCEMl9H (Marsh et al; Gene 32, 481-485, 1984). The heavy chain sequences included in pICEmu extend downstream of the EcoRI site located just 3' of the mu intronic enhancer, to the Xhol site located approximately 1 kb downstream of the last transmembrane exon of the mu gene; however, much of the mu switch repeat region has been deleted by passage in E. coli.

[0233] The targeting vector was constructed as follows. (See Figures 2A-2C, which depict further details.) A 1.3 kb HindIII/Smal fragment was excised from pICEmu and subcloned into HindIII/Smal digested pBluescript (Stratagene, La Jolla, CA). This pICEmu fragment extends from the HindIII site located approximately 1 kb 5' of Cmu1 to the Smal site located within Cmu1. The resulting plasmid was digested with Smal/Spel and the approximately 4 kb Smal/Xba1 fragment from pICEmu, extending from the Sma I site in Cmu1 3' to the Xba1 site located just downstream of the last Cmu exon, was inserted. The resulting plasmid, pTAR1, was linearized at the Smal site, and a neo expression cassette inserted. This cassette consists of the neo gene under the transcriptional control of the mouse phosphoglycerate kinase (pgk) promoter (Xbal/Taq1 fragment; Adra et al. (1987) Gene 60: 65-74) and containing the pgk polyadenylation site (PvuI/HindIII fragment, Boer et al. (1990) Biochemical Genetics 28: 299-308). This cassette was obtained from the plasmid pKJ1 (described by Tybulewicz et al. (1991) Cell 65: 1153-1163) from which the neo cassette was excised as an EcoRI/HindIII fragment and subcloned into EcoRI/HindIII digested pGEM-7Z (+) to generate pGEM-7 (KJ1). The neo cassette was excised from pGEM-7 (KJ1) by EcoRI/Sall digestion, blunt ended and subcloned into the Smal site of the plasmid pTAR1, in the opposite orientation of the genomic Cmu sequences.

[0234] The resulting plasmid was linearized with Not I, and a herpes simplex virus thymidine kinase (tk) cassette was inserted to allow for enrichment of ES clones bearing homologous recombinants, as described by Mansour et al. (1988) Nature 336: 348-352. This cassette consists of the coding sequences of the tk gene bracketed by the mouse pgk promoter and polyadenylation site, as described by Tybulewicz et al. (1991) Cell 65:1153-1163.

[0235] The resulting CMD targeting vector contains a total of approximately 5.3 kb of homology to the heavy chain locus and is designed to generate a mutant mu gene into which has been inserted a neo expression cassette in the unique Smal site of the first Cmu exon. The targeting vector was linearized with PvuI, which cuts within plasmid sequences, prior to electroproportion into ES cells.

[0236] Generation and analysis of targeted ES cells. AB-1 ES cells (McMahon, A. P. and Bradley, A., (1990) Cell 62: 1073-1085) were grown on mitotically inactive SNL76/7 cell feeder layers (ibid.), essentially as described in Teratocarcinosms and Embryonic Stem Cells: a Practical Approach, E. J. Robertson, Ed., Oxford: IRL Press, 1987, pp. 71-112. The linearized CMD targeting vector was electroporated into AB-1 cells by the methods described in Hasty et al. (1991) Nature 350: 243-246. Electroporated cells were plated into 100 mm dishes at a density of 1-2 x 10^6 cells/dish. After 24 hours, G418 (200 micrograms/ml of active component) and FIAU (5 x 10^-7 M) were added to the medium, and drug-resistant clones were allowed to develop over 8-9 days. Clones were picked, trypsinized, divided into two portions, and further expanded. Half of the cells derived from each clone were then frozen and the other half analyzed for homologous recombination between vector and target sequences.

[0237] DNA analysis was carried out by Southern blot hybridization. DNA was isolated from the clones as described by Laird et al., (1991) Nucleic Acids Res. 19:4293). Isolated genomic DNA was digested with Spel and probed with a 915 bp SacI fragment, probe A (Figure 2C), which hybridizes to a sequence between the mu intronic enhancer and the mu switch region. Probe A detects a 9.9 kb Spel fragment from the wild type locus, and a diagnostic 7.6 kb band from a mu locus which has homologously recombined with the CMD targeting vector (the neo expression cassette contains a Spel site).

[0238] Of 1132 G418 and FIAU resistant clones screened by Southern blot analysis, 3 displayed the 7.6 kb Spel I band indicative of homologous recombination at the mu locus. These 3 clones were further digested with the enzymes BglII, BstXI, and EcoRI to verify that the vector integrated homologously into the mu gene. When hybridized with probe A, Southern blots of wild type DNA digested with BglII, BstXI, or EcoRI produce fragments of 15.7, 7.3, and 12.5 kb, respectively, whereas the presence of a targeted mu allele is indicated by fragments of 7.7, 6.6, and 14.3 kb, respectively. All 3 positive clones detected by the Spel digest showed the expected BglII, BstXI, and EcoRI restriction fragments diagnostic of insertion of the neo cassette into the Cmu1 exon.

[0239] The three targeted ES clones, designated number 264, 272, and 408, were thawed and injected into C57BL/6J blastocysts as described by A. Bradley in Teratocarcinosms and Embryonic Stem Cells: a Practical Approach, E. J. Robertson, Ed., Oxford: IRL Press, 1987, pp. 113-151. Injected blastocysts were transferred into the uterus of pseudopregnant females to generate chimeric mice representing a mixture of cells derived from the input ES cells and the host blastocyst. The extent of ES cell contribution to the chimera can be
visually estimated by the amount of agouti coat coloration, derived from the ES cell line, on the black C57BL/6J background. Clones 272 and 408 produced only low percentage chimeras (i.e. low percentage of agouti pigmentation) but clone 264 produced high percentage male chimeras. These chimeras were bred with C57BL/6J females and agouti offspring were generated, indicative of germline transmission of the ES cell genome. Screening for the targeted mu gene was carried out by Southern blot analysis of BglI digested DNA from tail biopsies (as described above for analysis of ES cell DNA). Approximately 50% of the agouti offspring showed a hybridizing BglI band of 7.7 kb in addition to the wild type band of 15.7 kb, demonstrating a germline transmission of the targeted mu gene.

Analysis of transgenic mice for functional inactivation of mu gene. To determine whether the insertion of the neo cassette into Cmu1 has inactivated the Ig heavy chain gene, a clone 264 chimera was bred with a mouse homozygous for the JHD mutation, which inactivates heavy chain expression as a result of deletion of the JH gene segments (Chen et al, (1993) Immunol. 5: 647-656). Four agouti offspring were generated. Serum was obtained from these animals at the age of 1 month and assayed by ELISA for the presence of murine IgM. Two of the four offspring were completely lacking IgM (Table 1). Genotyping of the four animals by Southern blot analysis of DNA from tail biopsies by BglI digestion and hybridization with probe A (Figure 2C), and by Stul digestion and hybridization with a 475 bp EcoRI/Stul fragment (ibid.) demonstrated that the animals which fail to express serum IgM are those in which one allele of the heavy chain locus carries the JHD mutation, the other allele the Cmu1 mutation. Mice heterozygous for the JHD mutation display wild type levels of serum Ig. These data demonstrate that the Cmu1 mutation inactivates expression of the mu gene.

Table 1 presents the level of serum IgM, detected by ELISA, for mice carrying both the CMD and JHD mutations (CMD/JHD), for mice heterozygous for the JHD mutation (+/JHD), for wild type (129Sv x C57BL/6J)F1 mice (+/+), and for B cell deficient mice homozygous for the JHD mutation (JHD/JHD).

### Table 1

<table>
<thead>
<tr>
<th>Mouse</th>
<th>Serum IgM (micrograms/ml)</th>
<th>Ig H chain genotype</th>
</tr>
</thead>
<tbody>
<tr>
<td>42</td>
<td>&lt;0.002</td>
<td>CMD/JHD</td>
</tr>
<tr>
<td>43</td>
<td>196</td>
<td>+/JHD</td>
</tr>
<tr>
<td>44</td>
<td>&lt;0.002</td>
<td>CMD/JHD</td>
</tr>
<tr>
<td>45</td>
<td>174</td>
<td>+/JHD</td>
</tr>
<tr>
<td>129 x BL6 F1</td>
<td>153</td>
<td>+/+</td>
</tr>
<tr>
<td>JHD</td>
<td>&lt;0.002</td>
<td>JHD/JHD</td>
</tr>
</tbody>
</table>

EXAMPLE 3: Generation of transgenic mice

The HCo12 human heavy chain transgene. The HCo12 transgene was generated by coinjection of the 80 kb insert of pHc2 (Taylor et al., 1994, Int. Immunol., 6: 579-591) and the 25 kb insert of pVx6. The plasmid pVx6 was constructed as described below.

An 8.5 kb HindIII/Sall DNA fragment, comprising the germline human VH1-18 (DP-14) gene together with approximately 2.5 kb of 5′ flanking, and 5 kb of 3′ flanking genomic sequence was subcloned into the plasmid vector pSP72 (Promega, Madison, WI) to generate the plasmid p343.7.16. A 7 kb BamHI/HindIII DNA fragment, comprising the germline human VH5-51 (DP-73) gene together with approximately 5 kb of 5′ flanking and 1 kb of 3′ flanking genomic sequence, was cloned into the pBR322 based plasmid cloning vector pGP1f (Taylor et al. 1992, Nucleic Acids Res. 20: 6287-6295), to generate the plasmid p251f.

A new cloning vector derived from pGP1f, pGP1k (the sequence of which is presented in Figures 3A and 3B and SEQ ID NO:4), was digested with EcoRV/BamHI, and ligated to a 10 kb EcoRV/BamHI DNA fragment, comprising the germline human VH3-23 (DP47) gene together with approximately 4 kb of 5′ flanking and 5 kb of 3′ flanking genomic sequence. The resulting plasmid, p112.2RR-7, was digested with BamHI/Sall and ligated with the the 7 kb purified BamHI/Sall insert of p251f. The resulting plasmid, pVx4, was digested with Xhol and ligated with the 8.5 kb Xhol/Sall insert of p343.7.16. A clone was obtained with the VH1-18 gene in the same orientation as the other two V genes. This clone, designated pVx6, was then digested with NotI and the purified 26 kb insert coinjected, together with the purified 80 kb NotI insert of pHc2 at a 1:1 molar ratio, into the pronuclei of one-half day (C57BL/6J x DBA/2J)F2 embryos as described by Hogan et al. (B. Hogan et al., Manipulating the Mouse Embryo, A Laboratory Manual, 2nd edition, 1994, Cold Spring Harbor Laboratory Press, Plainview NY).

Three independent lines of transgenic mice comprising sequences from both Vx6 and HC2 were established from mice that developed from the injected embryos. These lines are designated (HCo12)14881, (HCo12)15083, and (HCo12)15087. Each of the three lines were then bred with mice comprising the CMD mutation described in Example...
Hybridoma Screening. To identify hybridomas secreting human antibodies against the IL-4R, ELISA plates were coated overnight with a non-neutralizing mouse anti-human IL-4R antibody M10 at 2 µg/ml in PBS. Plates were washed with 100 µl/well PBS-Tween (PBST) containing 1% BSA. Fifty µl cell culture supernatent was added followed by a 1.0 hour incubation. Plates were washed and then incubated for one hour with 100 µl/well goat anti-human IgG conjugated to horseradish peroxidase (Sigma #A-3813, or #A-7164). Plates were washed three times in PBS-Tween between each step.

Wells that read positive by ELISA were screened for their ability to block the binding of IL-4 to IL-4R. ELISA plates were coated overnight with a non-neutralizing mouse anti-human IL-4R antibody M10 at 2 µg/ml. Plates were washed 3X with PBST. 100 µl human IL-4R was added at 10 ng/ml in PBST and incubated for 1.0 hour. Plates were washed 4X with PBST and 100 µl supernatant samples were added and incubated for 1.0 hour. Wells were washed 4X with PBST. 5.0 ng/ml biotinylated IL-4 was added in PBST and incubated for 1.0 hour. 100 µl/well poly80 horseradish peroxidase (RDI) was added at 1:5000 in PBST and incubated for 45 minutes. Plates were washed 5X with PBST, and a colorimetric reagent (3,3',5,5' tetramethylbenzidine, available from Kirkegaard and Perry) was added at 100 µl/well. Reaction was stopped with 100 µl phosphoric acid and plates were read at 450nm. Absent or reduced signal was interpreted as the antibody binding to receptor in a manner that blocked IL-4 from binding to receptor.

EXAMPLE 4: Generation of Human Anti-IL-4R Monoclonal Antibodies

Transgenic mice Strain ((CMD)++; (JKD)++; (HCo7)11952/++; (KCo5)9272/++; which is homozygous for disruptions of the endogenous heavy chain (CMD) and kappa light chain (JKD) loci (see example 3), was used to generate IL-4R-reactive monoclonal antibodies. This strain also comprises a human kappa light chain transgene (HCo7) with individual animals either hemizygous or homozygous for insertion #11952. Each of these transgenic strains is homozygous for disruptions of the endogenous heavy chain (CMD) and kappa light chain (JKD) loci. Both strains also comprise a human kappa light chain transgene (HCo7), with individual animals either hemizygous or homozygous for insertion #11952. The two strains differ in the heavy chain transgene used. Mice were hemizygous or homozygous for either the HCo7 or the HCo12 transgene. The CMD mutation is described above in Example 2. The generation of (HCo12)15087 mice is described above (in this example). The JK D mutation (Chen et al. 1993, EMBO J. 12: 811-820) and the (KCo5)9272 (Fishwild et al. 1996, Nature Biotechnology 14: 845-851) and (HCo7)11952 mice, are described in U.S. Patent 5,770,429, which is hereby incorporated by reference.

EXAMPLE 5: Assay for assessing blocking activity

This assay is based on ability of both IL-4 and IL-13 to enhance the expression of the activation-associated surface antigen CD23 on human B cells. Antibodies are tested for the ability to inhibit CD23 expression induced by IL-
Cells that may be employed in preparing and loss of epithelial barrier function. Techniques that may be employed in assessing the ability of an IL-4 antagonist to inhibit IL-4-induced damage to epithelium number of techniques are known for determining whether an epithelial layer is intact. The following are examples but not limited to lung epithelium or intestinal epithelium. Damage to epithelium can result in loss of barrier function. A

EXAMPLE 7: Assays for Measuring Loss of Barrier Function

One hybridoma cell line generated by procedures described above (see example 4) is designated 6-2. The anti-IL-4R monoclonal antibody secreted by this hybridoma is a blocking antibody, as determined in a conventional plate binding assay, and thus functions as an IL-4 antagonist. The monoclonal antibody produced by 6-2 also exhibits the ability to reduce an IL-13-induced biological activity.

Described herein is a hybridoma cell line produced as described above, wherein the hybridoma secretes an isotype IgM MAb directed against human IL-4R. Also provided herein are IgG1 monoclonal antibodies derived from IgM monoclonal antibodies.

The DNA sequence of the variable region of the light chain of MAb 6-2 has been determined, and is presented in SEQ ID NO:7; the amino acid sequence encoded thereby is presented in SEQ ID NO:8. Complementarity determining regions 1 to 3 (CDR 1-3) are believed to correspond to amino acids 31-35, 50-66, and 99-107, of SEQ ID NO:6, respectively.

EXAMPLE 6: Hybridoma Cell Line

One hybridoma cell line generated by procedures described above (see example 4) is designated 6-2. The anti-IL-4R monoclonal antibody secreted by this hybridoma is a blocking antibody, as determined in a conventional plate binding assay, and thus functions as an IL-4 antagonist. The monoclonal antibody produced by 6-2 also exhibits the ability to reduce an IL-13-induced biological activity.

Described herein is a hybridoma cell line produced as described above, wherein the hybridoma secretes an isotype IgM MAb directed against human IL-4R. Also provided herein are IgG1 monoclonal antibodies derived from IgM monoclonal antibodies.

The DNA sequence of the variable region of the light chain of MAb 6-2 has been determined, and is presented in SEQ ID NO:7; the amino acid sequence encoded thereby is presented in SEQ ID NO:8. Complementarity determining regions 1 to 3 (CDR 1-3) are believed to correspond to amino acids 31-35, 50-66, and 99-107, of SEQ ID NO:6, respectively.

EXAMPLE 7: Assays for Measuring Loss of Barrier Function

A method provided herein involves use of IL-4 antagonists to inhibit IL-4-induced damage to epithelium, including but not limited to lung epithelium or intestinal epithelium. Damage to epithelium can result in loss of barrier function. A number of techniques are known for determining whether an epithelial layer is intact. The following are examples of techniques that may be employed in assessing the ability of an IL-4 antagonist to inhibit IL-4-induced damage to epithelium and loss of epithelial barrier function.

Cells that may be employed in preparing in vitro models of epithelium (epithelial barriers) are known. For example, Calu-3 human lung epithelial cells are suitable for use in barrier function studies. Another suitable cell line is the human intestinal epithelial cell line designated T84. T84 cells are cultured under conditions that result in formation of a monolayer of epithelial cells on a permeable support, as described in Madara, J. and K. Dharmasathaphorn (J. Cell Biol., 101:2124-2133, 1985), Madara, J. and J. Stafford (J. Clin. Invest. 83:724-727, 1989), and Youakim, A. and M. Ahdieh (Am. J. Physiol. 276 (Gastrointest. Liver Physiol. 39):G1279-G1288, 1999). The cultured monolayers are tested for properties such as resistance to passive transepithelial ion flow (such resistance indicating an intact monolayer performing a barrier function). The thus-generated epithelial monolayer simulates the intestinal epithelial barrier.

One type of assay determines whether a particular radiolabeled compound is able to cross an epithelial monolayer (e.g., a monolayer generated as described above). Transport of the radiolabeled compound across the monolayer indicates that the barrier is permeable rather than intact. One such procedure is mannitol flux analysis, which assesses movement of radiolabeled mannitol (e.g., ³H mannitol) across a monolayer (see Madara and Stafford, supra).

Methods for imaging a monolayer are identified in Madara and Stafford, supra. Such imaging methods are an alternative for assessing the condition of an epithelial layer, after exposure to IL-4 with or without an antagonist.

Youakim and Ahdieh, supra, discuss proteins that are part of "tight junction" complexes in intact intestinal epithelial barriers, and report studies of the effect of IFN-γ on proteins associated with tight junctions. Other techniques
for studying the effect of a cytokine on barrier function are described. For example, the effect of a cytokine on monolayer permeability may be assessed by transepithelial electrical resistance measurements, using techniques described in the reference.

U.S. Patent 6,033,688 also describes procedures that may be employed in studies of barrier permeability; see especially examples 1 and 4 of the patent. Human tracheal epithelial cells were cultured under conditions that yielded a monolayer exhibiting transepithelial electrical resistance. Transepithelial resistance (indicating an intact barrier) was determined using a voltmeter. The effect of a particular reagent (HGH) on the epithelial monolayer was assessed by exposing the monolayer to HGH, and then measuring ion transport activities in Ussing chambers, by standard methods (column 8, lines 40-56). Similar studies were conducted on monolayers that were generated from bronchial epithelial cells from a human cystic fibrosis patient (example 4, column 11).

Using any of the above-described barrier function assay procedures, an epithelial monolayer is exposed to IL-4 alone, or exposed to IL-4 in the presence of an IL-4 antagonist. The antagonist’s ability to inhibit the IL-4-induced reduction in barrier function thus is assessed.

In one such assay, a monolayer of T84 cells served as an in vitro model of an intestinal epithelial barrier, as discussed above. IL-4 added to the basolateral side of polarized epithelial cells was found to reduce barrier function by 70% within 48-72 hours of treatment. When an IL-4 receptor polypeptide was added at the same time as IL-4, the reduction in barrier function was prevented, and the barrier was maintained at the same level as untreated (control) cells. A soluble human IL-4 receptor polypeptide, consisting of the extracellular domain, was employed in the assay.

The assay procedure also was conducted on a monolayer derived from lung epithelial cells, which served as an in vitro model of a lung epithelial barrier. IL-4 added to the basolateral side of polarized lung epithelial cells was found to reduce barrier function by 50% within 48-72 hours of treatment. When the IL-4 receptor polypeptide was added at the same time as IL-4, the reduction in barrier function was prevented, and the barrier was maintained at the same level as untreated (control) cells.

EXAMPLE 8: Monoclonal Antibody designated 12B5

A human monoclonal antibody directed against human IL-4 receptor was prepared by the following procedure. The monoclonal antibody, which is designated 12B5, is a blocking antibody that functions as an IL-4 antagonist and as an IL-13 antagonist.

The procedure began with immunization of a transgenic mouse with a soluble human IL-4 receptor polypeptide. Mouse strain ((CM)(+)(++)(JKD)(++)((Co7)11952+/+)(KCo5)9272+/+), described in example 3 above, was employed. The antigen for immunization was purified soluble IL-4R, comprising the extracellular domain of human IL-4 receptor (500ug/ml). The mouse was initially immunized with 50 ug antigen emulsified in Complete Freund’s Adjuvant, followed by two more immunizations with Incomplete Freund’s Adjuvant at 50ug and then 25ug. Immunization was every two weeks by intraperitoneal injection. A specific human IgG anti-IL-4R titer from the serum was performed by ELISA, eight days after the last injection. A good titer against the target was detected, and the mouse then was IV/IP boosted 25ug each on day 3 (i.e., 3 days before the mouse was sacrificed) and 15ug IV on day 2.

The mouse was sacrificed, and spleen cells were extracted and fused with the murine myeloma cell line P3x63Ag8.653 (ATCC CRL 1580). A conventional PEG fusion protocol was followed. The fusion was screened for HuIgG and HuKappa, followed by a rescreen for human gamma and kappa specific to IL-4R. The positive clones were evaluated, and clone 12B5 was identified as a blocking antibody. The clone was subcloned; a hybridoma cell line that produces MAb 12B5 was isolated; and MAb 12B5 was purified from the supernatant.

12B5 was determined to be an IgG1 antibody, and to be fully human. Antibodies of other subclasses, such as IgG4 or IgM monoclonal antibodies, may be derived from 12B5. Techniques for altering (switching) the subclass/isotype of an antibody are known. The constant region of 12B5 may be replaced, for example, with a constant region derived from a human IgG4 antibody. Sequence information for a human IgG4 heavy chain is presented, for example, in Ellison et al. (DNA Vol. 1, no. 1, pp 11-18, 1981), which is hereby incorporated by reference herein.

DNA encoding the variable region of the light chain of MAb 12B5 was isolated, and the nucleotide sequence thereof was determined. The DNA sequence for the light chain variable region is presented as SEQ ID NO:9; the amino acid sequence encoded thereby is presented in SEQ ID NO:10. Complementarity determining regions 1 to 3 (CDR 1-3) are believed to correspond to amino acids 24-35, 51-57, and 90-99, of SEQ ID NO:10, respectively.

DNA encoding the variable region of the heavy chain of MAb 12B5 was isolated, and the nucleotide sequence thereof was determined. The DNA sequence for the heavy chain variable region is presented as SEQ ID NO:11; the amino acid sequence encoded thereby is presented in SEQ ID NO:12. CDR-1 of the heavy chain is believed to correspond to amino acids 31-35; CDR-2 to amino acids 50-65; and CDR-3 to amino acids 98-104 of SEQ ID NO:12.
EXAMPLE 9: Additional Monoclonal Antibodies that inhibit both IL-4 and IL-13

[0275] Additional human monoclonal antibodies were raised against human IL-4 receptor, by immunizing transgenic mice with a soluble human IL-4R polypeptide. The transgenic mice employed were selected from the transgenic mouse strains described in example 3.

[0276] Hybridoma cell lines secreting human monoclonal antibodies that specifically bind human IL-4R, and which are capable of functioning as IL-4 antagonists and IL-13 antagonists, were identified and isolated. The MAbs are designated 27A1, 5A1, and 63. Another MAb, designated 1B7, was derived from MAb 63, and differs from the parent antibody only in the light chain. 1B7 retains the ability to bind IL-4R and to function as an IL-4 antagonist and an IL-13 antagonist.

[0277] The DNA sequence of the variable region of the light chain of MAb 27A1 is presented in SEQ ID NO:13, and the encoded amino acid sequence is presented in SEQ ID NO:14. Complementarity determining regions 1 to 3 (CDR 1-3) are believed to correspond to amino acids 24-35, 51-57, and 90-99, of SEQ ID NO:14, respectively.

[0278] The DNA sequence for the variable region of the heavy chain of MAb 27A1 is presented as SEQ ID NO:15, and the encoded amino acid sequence is presented in SEQ ID NO:16. Complementarity determining regions 1 to 3 (CDR 1-3) are believed to correspond to amino acids 31-35, 50-66, and 99-105, of SEQ ID NO:16, respectively.

[0279] The DNA sequence of the variable region of the light chain of MAb 5A1 is presented in SEQ ID NO:17, and the encoded amino acid sequence is presented in SEQ ID NO:18. Complementarity determining regions 1 to 3 (CDR 1-3) are believed to correspond to amino acids 24-34, 50-56, and 89-97 of SEQ ID NO: 18, respectively.

[0280] The DNA sequence for the variable region of the heavy chain of MAb 5A1 is presented as SEQ ID NO:19, and the encoded amino acid sequence is presented in SEQ ID NO:20. Complementarity determining regions 1 to 3 (CDR 1-3) are believed to correspond to amino acids 31-35, 50-65, and 98-112 of SEQ ID NO:20, respectively.

[0281] The DNA sequence of the variable region of the light chain of MAb 63 is presented in SEQ ID NO:21, and the encoded amino acid sequence is presented in SEQ ID NO:22. Complementarity determining regions 1 to 3 (CDR 1-3) are believed to correspond to amino acids 24-34, 50-56, and 89-97 of SEQ ID NO:22, respectively.

[0282] The DNA sequence for the variable region of the heavy chain of MAb 63 is presented as SEQ ID NO:23, and the encoded amino acid sequence is presented in SEQ ID NO:24. Complementarity determining regions 1 to 3 (CDR 1-3) are believed to correspond to amino acids 31-35, 50-66, and 99-106 of SEQ ID NO:24, respectively.

[0283] The DNA sequence of the variable region of the light chain of MAb 1B7 is presented in SEQ ID NO:25, and the encoded amino acid sequence is presented in SEQ ID NO:26. Complementarity determining regions 1 to 3 (CDR 1-3) are believed to correspond to amino acids 24-34, 50-56, and 89-97 of SEQ ID NO:26, respectively.

[0284] MAb 1B7 was derived from MAb 63, and the heavy chains of the two MAbs are identical. Thus, the DNA sequence for the variable region of the heavy chain of MAb 1B7 is presented as SEQ ID NO:23, and the encoded amino acid sequence is presented in SEQ ID NO:24. Complementarity determining regions 1 to 3 (CRDs 1-3) are believed to correspond to amino acids 31-35, 50-66, and 99-106 of SEQ ID NO:24, respectively.

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EP 2 990 420 B1

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Lys Gly Arg Phe Thr Val Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr 240
65 70 75 80

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Leu Glu Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys 288
85 90 95

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24
Claims

1. A human monoclonal antibody capable of inhibiting an IL-4 induced biological activity that competes with a reference antibody for binding to a cell that expresses human IL-4 receptor (IL-4R), wherein:
   a) the light chain of the reference antibody comprises the sequence of SEQ ID NO:6 and the heavy chain of the reference antibody comprises the sequence of SEQ ID NO:8; or
   b) the light chain of the reference antibody comprises the sequence of SEQ ID NO:26 and the heavy chain of the reference antibody comprises the sequence of SEQ ID NO:24.

2. An antibody as claimed in claim 1, wherein said antibody additionally is capable of inhibiting an IL-13-induced biological activity.

3. The antibody as claimed in claim 1 or claim 2 for use in a method of treating dermatitis, atop dermatitis, contact dermatitis, or asthma.

4. The antibody for use as claimed in claim 3, wherein said antibody is in the form of a composition that additionally comprises a diluent, excipient, or carrier.

5. The antibody as claimed in claim 1 or claim 2, or for use as claimed in claim 3 or claim 4, wherein the IL-4R comprises amino acids 1 to 197 of SEQ ID NO:2.

6. A nucleic acid encoding the light chain of the antibody as claimed in claim 1 or claim 2.

7. A nucleic acid encoding the heavy chain of the antibody as claimed in claim 1 or claim 2.

8. A host cell comprising the nucleic acid of claim 6 or claim 7.

Patentansprüche

1. Humaner monoklonaler Antikörper, der zur Hemmung einer IL-4-induzierten biologischen Aktivität fähig ist, welcher mit einem Referenz-Antikörper um die Bindung an eine Zelle konkurriert, die humanen 11-4-Rezeptor (IL-4R) exprimiert, wobei:
   a) die leichte Kette des Referenz-Antikörpers die Sequenz der SEQ ID NO: 6 umfasst und die schwere Kette des Referenz-Antikörpers die Sequenz der SEQ ID NO: 8 umfasst; oder
   b) die leichte Kette des Referenz-Antikörpers die Sequenz der SEQ ID NO: 26 umfasst und die schwere Kette des Referenz-Antikörpers die Sequenz der SEQ ID NO: 24 umfasst.

2. Antikörper, wie in Anspruch 1 beansprucht, wobei dieser Antikörper außerdem zum Hemmen einer IL-13-induzierten biologischen Aktivität fähig ist.

3. Antikörper, wie in Anspruch 1 oder Anspruch 2 beansprucht, zur Anwendung in einem Verfahren zur Behandlung von Dermatitis, atopischer Dermatitis, Kontaktdermatitis oder Asthma.

4. Antikörper zur Anwendung, wie in Anspruch 3 beansprucht, wobei dieser Antikörper in der Form einer Zusammensetzung vorliegt, die außerdem ein Verdünnungsmittel, einen Exzipienten oder einen Träger umfasst.

5. Antikörper, wie in Anspruch 1 oder Anspruch 2 beansprucht, oder zur Anwendung, wie in Anspruch 3 oder Anspruch 4 beansprucht, wobei der IL-4R Aminosäuren 1 bis 197 der SEQ ID NO: 2 umfasst.

6. Nukleinsäure, welche die leichte Kette des Antikörpers, wie in Anspruch 1 oder Anspruch 2 beansprucht, kodiert.

7. Nukleinsäure, welche die schwere Kette des Antikörpers, wie in Anspruch 1 oder Anspruch 2 beansprucht, kodiert.

Revendications

1. Anticorps monoclonal humain capable d’inhiber une activité biologique induite par l’IL-4 qui est en concurrence avec un anticorps de référence pour la liaison à une cellule qui exprime le récepteur de l’IL-4 humaine (IL-4R), dans lequel :
   a) la chaîne légère de l’anticorps de référence comprend la séquence de SEQ ID N° : 6 et la chaîne lourde de l’anticorps de référence comprend la séquence de SEQ ID N° : 8 ; ou
   b) la chaîne légère de l’anticorps de référence comprend la séquence de SEQ ID N° : 26 et la chaîne lourde de l’anticorps de référence comprend la séquence de SEQ ID N° : 24.

2. Anticorps selon la revendication 1, dans lequel ledit anticorps est en outre capable d’inhiber une activité biologique induite par l’IL-13.

3. Anticorps selon la revendication 1 ou la revendication 2 pour utilisation dans un procédé de traitement de la dermite, de la dermite atopique, de la dermite de contact ou de l’asthme.

4. Anticorps pour utilisation selon la revendication 3, dans lequel ledit anticorps se présente sous la forme d’une composition qui comprend en outre un diluant, un excipient ou un vecteur.

5. Anticorps selon la revendication 1 ou la revendication 2, ou pour utilisation selon la revendication 3 ou la revendication 4, dans lequel l’IL-4R comprend les acides aminés 1 à 197 de la SEQ ID N° : 2.

6. Acide nucléique codant pour la chaîne légère de l’anticorps selon la revendication 1 ou la revendication 2.

7. Acide nucléique codant pour la chaîne lourde de l’anticorps selon la revendication 1 ou la revendication 2.

8. Cellule hôte comprenant l’acide nucléique selon la revendication 6 ou la revendication 7.
FIGURE 1A

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Glu Pro Ser Leu Arg Ile Ala Ala Ser Thr Leu Lys Ser Gly Ile 170

TCC TAC AGG GCA CGG GTG AGG GCC TGG GTT CAG TGC TAT AAC ACC 555
Ser Tyr Arg Ala Arg Val Arg Ala Trp Ala Gln Cys Thr Asn Thr 185

ACC TGG AGT GAG TGG AGC CCC AGC ACC AAG TGG CAC AAC TCC TAC 600
Thr Thr Ser Glu Trp Ser Pro Ser Thr Lys Thr Asp Ser Thr Tyr 200

AGG GAG CCC TCC GAG CAG CAC CTC CTG CGC TGG GTC AGC GTT TCC 645
Arg Glu Pro Phe Glu Gln His Leu Leu Leu Gly Val Ser Ser Leu 215

TGC ATT GTC ATC CTG GCC TGC CTG CTG TGG TAT GTC AGC ATC 690
Cys Ile Val Ile Leu Ala Val Cys Leu Leu Cys Tyr Val Ser Ile 230

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CGA AAT GTC CTC CAG CAT GGG GCA GCT GCA GCC CCC GTC TCG ARG 1635
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ACC AGT GGC TAT CAG GAG TTT GTA CAT GCG GTG GAG CAG GGT 1680
Pro Thr Ser Gly Tyr Gln Glu Phe Val His Ala Val Glu Gln Gly 560

GCC ACC CAG GGC AGT GCG GTG GTG GGC TGG GGT CCC CCA GGA GAG 1725
Gly Thr Gln Val Ala Val Ala Val Gly Leu Gly Pro Gly Glu 575

GCT GGT TAC AAG GCC TTC TCA AGC CTG CTT GCC AGC AGT GCT GTG 1770
Ala Gly Tyr Lys Ala Phe Ser Ser Leu Leu Ala Ser Ser Ala Val 590

TCC CCA GAG AAA TGT GGG TTT GGT GCT AGC AGT GGG GAA GAG GGG 1815
Ser Pro Glu Lys Cys Gly Phe Gly Ala Ser Gly Ala Ser Gly Glu 605

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Tyr Lys Pro Phe Gln Asp Leu Ile Pro Gly Cys Pro Gly Asp Pro 620

GCC CTC GTC CCT GTC CCC TCG TTC ACC TTT GGA CTG GAC AGG GAG 1905
Pro Val Pro Leu Pro Val Phe Thr Phe Gly Leu Arg Glu 635

CCA CCT CGC AGT CCG CAG AGC TCA CAT CTC CCA AGC AGC TCC CCA 1950
Pro Pro Arg Ser Pro Gln Ser Ser His Leu Pro Ser Ser Ser 650

GAG CAC CTG GGT CTG GAG CCG GGG GAA AAG GTA GAG GAC ATG CCA 1995
Glu His Leu Gly Leu Glu Pro Gly Glu Lys Val Glu Asp Met Pro 665

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Lys Pro Pro Leu Pro Gln Glu Glu Ala Thr Asp Pro Leu Val Asp 680

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Ser Leu Gly Ser Gly Ile Val Tyr Ser Ala Leu Thr Cys His Leu 695

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Cys Gly His Leu Lys Glu Lys His Gli Gly Glu Asp Gly Gly Glu 710

ACC CCT GTC ATG GCC AGT CCT TGC TGT GCC TGC TGT GGA GAC 2175
Thr Pro Val Met Ala Ser Pro Cys Gly Cys Cys Gly Asp 725

AGG TCC TCG CCC CCT ACA ACC CCC CTG AGG GCC CCA GAC CCC TCT 2220
Arg Ser Ser Pro Pro Thr Thr Pro Leu Arg Ala Pro Asp Pro Ser 740

CCA GGT GGG GCT CTA CAG GAC AGT TCT TGT CCC GCC TCC CTG 2265
Pro Gly Gly Val Pro Leu Glu Ala Ser Leu Cys Pro Ser Leu 755

GCA CCC TCG GCC ATC TCA GAG AGT AAA TCC TCA TCA TCC TCC 2310
Ala Pro Ser Gly Ile Ser Glu Lys Ser Ser Ser Ser Phe 770

CAT CCT GCC CCT GGC AAT GCT CAG AGC TCA AGC CAG ACC CCC AAA 2355
His Pro Ala Pro Gly Asn Ala Gln Ser Ser Ser Ser Ser Ser 785

ATC GTG AAC TTT GTC TCC GTG GGA CCC ACA TAC ATG AGG GTC TCT 2400
Ile Val Asn Phe Val Ser Val Gly Pro Thr Tyr Met Arg Val Ser 800
FIGURE 3A

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CCTACTGTCG CGGGAATGGC GATCCGCTG GATATGATGT ATCCTGAGGT 100
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FIGURE 3B

CCATTTGCTGC AGGCATCGTG GTCGTACGCT CGTCTTTGG TGATGGCTTCA 2450
TTCAGCTCCG GTCCTCAACC ATCAAGGCCA GTTATGATG CCCCAGGTT 2500
GTGCAAAAAA CGGTTGAGCT CTTCCGTCC TCCGATGGT GTCAAGAGTA 2550
AGTTGGCAGC AGTGGTATCA CTCATGGTAA TGGCAGCACT GCATAATTCCT 2600
CTTACTGTCA TGCCATCCGT AAGATGCTTT TCTGTGACTG GTGAGTACTC 2650
AACCAAGTCA TTCTGAGAAT AGTGTAGCCG GCGACGAGCT TGCCTCTGCC 2700
CGGGTTCAAC ACGGATAAAA ACCGGCCACC ATAGCAGAAC TTTAAAATGG 2750
CTCATCATCG TAAAACGTCG TTCGGGCGGA AACTCTGCAAA GGAATTTACC 2800
GCTGGTGAGA TCCAGTTCCG TGTAACCAC TCGTGCAACC AACTGATCTT 2850
CAGCATCTTT TACTTTCACT AGCGTTTCTG GTGAGCAAA AAGAGGAAGG 2900
CAAATGCCC CAAAAGAGG AATAAGGCGC ACAGGAATG GTGAATACCT 2950
CATACTCTTC CTCTTCATAT ATATTGAAG CATTTATCG GGTATTGTC 3000
TCAATGAGCC ATACATATTT GAATGTATTG AGAAAAATAT ACAAAATAGG 3050
GTTCCCCGCA CATTTCCTCC AAAAACTGCC CCTGACGCTC GAAAGACCAT 3100
TATTATCATG ACATTTACCT ATAAAAATAG GCTATCACG AGGCCCTTTC 3150
GTATTCAAG 3159
FIGURE 4

1B7Vk  EIVLTQSPSS VSASVGRVT ITCRASQGIS N.RLAWYQQK PGKAEKLLIY IASILQRGVP
63Vk  D-QM------ -------------- T-W------H-- -------------- V--S--S--
12B5Vk  -----GT L-L-P-E-A- L3------SV-- SSY------ --Q--R--F G--SRAT-I--
6-2Vk  -----GT L-L-P-E-A- L3------SV-- TSY------ --Q--R------ G--SRAT-I--
5A1Vk  -----AT L-L-P-E-A- L3------SV-- .SY------ --Q--R---- H--NRAT-I--
27A1Vk  -----GT L-L-P-E-A- L3------SV-- SSY------ --Q--R------ G--SRAT-I--

1B7Vk  SRFSGSGSST DFTLTITRLQ PEDFATYQCQ Q.ANSFPFTF GFGKVDIK
63Vk  --------------- SS-- -------------- -------------- --------------
12B5Vk  D------ ----S--E ------V-- ------YGS-P-W-- --Q----E--
6-2Vk  D------ ----S--E ------V-- ------YGS-P---- --Q----E--
5A1Vk  A------ ----S--E ------V-- ------RSNW-L-- --G----E--
27A1Vk  D------ ----S--E ------V-- ------YGS-P-W-- --Q----E--
REFERENCES CITED IN THE DESCRIPTION

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